

CODEN [USA]: IAJPBB

ISSN: 2349-7750

INDO AMERICAN JOURNAL OF PHARMACEUTICAL SCIENCES

Available online at: <u>http://www.iajps.com</u>

Review Article

NON-MEDICATION TECHNIQUES FOR WOUND HEALING: CLINICAL CONSIDERATIONS

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Received: 26 March 2017	Accepted: 10 April 2017	Published: 18 April 2017
Abstraat		

Abstract

Objective: Several non-medication techniques have been proposed for the treatment of chronic wounds. Electric and magnetic fields, electromagnetic fields [EMFs], ultrasound [US], and photostimulation are some of these techniques with promising potentials. However, the clinical efficacy of these techniques for different chronic wounds is still not fully understood and standard guidelines on the allowed doses and possible side-effects should be determined. This paper aims to comprehensively review the therapeutic efficacies and clinical considerations of the main non-drug techniques for chronic wounds.

Methods: The databases of PubMed [1985-2016], EMBASE [1985-2016], Web of Sciences [1985-2016], and Google Scholar [1980-2016] were searched using the set terms of "non-medication" OR "non-drug treatment" AND "wound treatment". The obtained results were screened for the title and abstract by two authors and the relevant papers were reviewed for further details.

Results: Pulsed EMFs [PEMFs], non-contact low frequency US or MIST therapy, and Low Level Laser Therapy [LLLT] are the main non-drug techniques with promising effective outcomes for different wounds. PEMFs and MIST therapy have been used in some clinical studies with promising outcomes. In addition different lasers particularly HeNe lasers have shown therapeutic effect of superficial wounds. Despite of rigorous evidence on the therapeutic efficiency of these techniques, the main limit on developing approved clinical protocols of these techniques for wound treatment is the lack of definite dose-response on the clinical trials of these techniques.

Conclusion: The available data showed the therapeutic efficacy of PEMFs, MIST, and LLLT techniques for chronic wounds. Further in vitro and in vivo preclinical and clinical trials are needed to understand the mechanism of actions of these techniques for developing clinical protocols and guidelines of these techniques for treatment of different wounds.

Key words: Non-medication, Wound, Clinical Considerations, Treatment

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Please cite this article in press as Ali Yadollahpour et al, Non-Medication Techniques for Wound Healing: Clinical Considerations, Indo Am. J. Pharm. Sci, 2017; 4(04).

INTRODUCTION:

Wounds are among the most prevalent disorders with significant burden on healthcare systems worldwide. Despite of different medications for wound treatment, high costs of medications, resistant to conventional medications and high prevalence of chronic wounds have necessitated developing non-medication safe and feasible treatments. Nowadays, different non-medication techniques have been developed for the treatment of wounds that they are generally used for treatment and prevention of pressure wounds. Several methods such as light and photostimulation techniques [1-4], direct current [5], electric and magnetic fields [3, 6-9], and ultrasound [US] waves [10-15] have been shown promising therapeutic outcomes for different chronic and acute wounds.

Direct current and electric field, magnetic fields and electromagnetic field [EMFs] are among the first group of non-medication techniques developed for wound treatment. Several animal and human studies have shown that pulsed EMFs [PEMFs] can accelerate wound healing [16-20]. The results of these studies showed that PEMFs promote wound healing by up-regulation of FGF-2–mediated angiogenesis, nerve regeneration, alteration of the cell proliferation rate, changes in the levels of mRNA and protein synthesis, alteration of cellular membrane's permeability, and Ca2+, Na+, K+ ion transfer, effect on the production of melatonin and influence the expression of early-induced genes such as c-myc, c-fos, c-jun [16, 21-25].

Since the discoveries of potential therapeutic effects of US energy, various US technologies have been investigated for treatment of several disorders including skin wounds, malignant tumors, and bone fractures [26, 27]. Advantages of US treatments have made them one of the most promising treatment options for the management of soft tissue injuries [28]. Many experimental studies have shown various physiological efficacies of US on living tissues [12, 14, 29, 30] and also vigorous evidence indicating the beneficial effects of these mechanical waves in the treatment of disorders involving soft tissues [31-33].

Low level laser therapy [LLLT] is an another technique that it has shown clinical efficacy for tissue healing by facilitates collagen synthesis, keratinocyte cell motility, and growth factor release and transforms fibroblasts to myofibroblasts [34-39].

However, the clinical efficacy of these techniques is still not fully understood and further studies are needed to determine the exact mechanism of action and also possible side-effects of these techniques on different chronic wounds.

METHOD:

The databases of PubMed [1985-2016], EMBASE [1985-2016], Web of Sciences [1985-2016], and Google Scholar [1980-2016] were searched using the set terms of "non-medication" OR "non-drug" AND "wound treatment". The obtained results were screened for the title and abstract by at least two authors and the relevant papers were reviewed for further details.

Animal and human studies in both in vivo and in vitro designs that evaluated the effects of any nonmedication treatments in any type of wounds were included for further review. Due to the immense body of literature in this field, this study aims to provide a comprehensive and descriptive overview of the recent advances in applications of nonmedication techniques for treatment of wounds as well as their clinical applications and perspectives. The initial review showed the most common and promising techniques were EMFs, LLLT, and US techniques. Therefore, these three techniques were reviewed in more details.

RESULTS:

Electromagnetic fields and wound

Chronic wounds are caused to high rate of morbidity and mortality and have a profound economic impact for human and health care systems [40-43]. The pathogenesis of wound healing is not completely understood. Evidence from in vitro and in vivo studies models is shown several abnormalities in different phases of the wound-healing process. In particular, in diabetic wound some mechanisms such as inflammatory response, angiogenesis, fibroplasias, defects in deposition and differentiation collagen of extracellular matrix are disturbed [44-47]. Results of several human clinical trials and animal studies have shown that electrical stimulation applied to full thickness excisional wounds produced a reduction in wound size and accelerates wound healing, probably with increasing the endogenous current induced by injury [48-52]. Direct current stimulation has the disadvantage of requiring electrode placement directly on or near the wound, whereas pulsed electromagnetic fields have an inherent advantage that the electromagnetic signal influences the dressing and tissue involved. The basic mechanisms of the clinical effects of pulsed electromagnetic fields are not clear. Based on the results, researchers suggested that PEMFs may cause to specific, measurable cellular responses such as DNA synthesis, transcription, and protein synthesis by altering or augmenting pre-existing endogenous electrical fields [8, 53]. Results of studies have reported that PEMF stimulation

induces differentiation of skin fibroblasts in culture and decreases the doubling time of fibroblasts and endothelial cells and as a result decreases woundhealing time and increases the tensile strength of scar tissue [54-56].

Previous studies showed that PEMFs may contribute to wound healing by increasing collagen synthesis, angiogenesis, and bacteriostasis [20, 54]. These specifications are appropriated PEMFs for delayed wound healing in diabetic patients and accelerating of wound healing and also PEMFs are able to prevent tissue necrosis in diabetic tissue after an ischemic insult [16, 18, 19, 57].

Ultrasound and wound

The current range of frequency and in therapeutic US is 0.75-3MHz. Low frequency US waves have more penetration depth and less focused. The frequencies is choose for deep injuries or superficial lesions based on the penetration depth. For example 1 MHz US has 3-5 cm penetration depth on tissues and is an ideal choice for deeper injuries or 3 MHz US has 1-2 cm penetration depth on tissues and is applied for more superficial lesions [15, 58]. The US waves have high penetration on tissues with high-water content like fat because of its low absorption. Whereas tissues which are rich in protein like skeletal muscle have high US adsorption [59, 60]. With using pulsed waves and moving the transducer during the treatment process can avoid generating a standing wave and its side effects [15, 59, 61]. Low frequency US have shown effectiveness effects on accelerating the healing speed of open wounds and also an effective treatment for suspected deeptissue injuries. However the results have shown therapeutic efficacies of US techniques in different wounds, there is not an exact dose-response for clinical applications of US treatments in different wounds. Therefore to reach a standard treatment, further studies are needed to demonstrate the exact mechanism of action and also to provide exact dose-response of therapeutic US for different wounds.

Non-contact low frequency US

Non-contact low frequency ultrasound [NCLF-US] devices have been used to treat of some kinds of wounds. Results of several studies have shown that ultrasound has an effect on decreasing the bacterial count in wounds, inflammatory cytokines and pain [13, 62, 63]. Yao et al. [2014] in a pilot study evaluated effects of non-contact low-frequency ultrasound and its molecular mechanism on diabetic foot ulcers [DFUs]. One of the aims of their study was to evaluate and explore the correlation between wound healing and change of cytokine, proteinase and growth factor profile. Results showed reduction on pro-inflammatory cytokines [IL-6, IL-8, IL-1 β , TNF- α , and GM-

CSF], matrix metalloproteinase-9 [MMP-9], vascular endothelial growth factor [VEGF] and macrophages in treatment group compared with control group. The results demonstrated that NCLF-US is effective in treating neuropathic with inhibiting prodiabetic foot ulcers inflammatory cytokines in chronic wound and improving tissue regeneration [63]. Honaker et al. [2013] in a retrospective analysis study evaluated the effectiveness of non-contact low-frequency ultrasound on the healing of suspected deep tissue injury [SDTI]. They measured surface area, wound colour/tissue assessment, and skin integrity with potential scores of 3 to 18 [higher scores indicate greater severity] before and after treatment in patients. The results showed reduction in wound severity for the intervention group [1.45] and increase in the non-intervention group [1.06]. They suggested that non-contact low frequency ultrasound is effectiveness on healing of SDTI [13].

Low level laser and wound

Low level laser therapy [LLLT] has been shown beneficial effects on tissue healing and pain relief. However, the results of in vitro and in vivo studies have shown varies reported [1, 35, 64]. Results of several studies have reported the effectiveness of using the helium neon [HeNe] and gallium arsenide lasers in cell proliferation and collagen production [1]. They suggested that the effectiveness of laser therapy may be related to photothermal, photochemical, or photomechanical effects but the exact mechanism is not yet clear [1, 35, 64]. Although the conflicting results have limited and disputed using low level laser therapy but it is widespread used clinically in the treatment of various neurologic, chiropractic, dental, and dermatologic disorders [35, 65-67]. The low level laser therapy [LLLT] has been also used in Dentistry to improve wound healing. Lopes et al. [2001] in a study investigated the effect of LLLT on the in vitro proliferation of gingival fibroblasts. The results of their study showed that a smaller laser exposure time results in higher proliferation and improve the fibroblast proliferation [67]. Medrado et al. [2003] evaluated the effects of low level laser therapy on wound healing and its biological action upon myofibroblasts. Before and after treatment, the tissues were assessed by histology, immunohistochemistry, and electron microscopy. The results showed that low level laser therapy induced increased collagen deposition, reduced the inflammatory reaction, and a greater proliferation of myofibroblasts [64]. Further studies with controlled dose-response design are needed for better understanding of the mechanisms of action of LLLT to develop clinical applications of the technique.

CONCLUSION:

Several techniques with diverse mechanisms have been suggested for treatment of different types of wound [35]. PEMFs have shown beneficial effects on wound healing through the production of small quantities of free radicals within cells, DNA synthesis, transcription, protein synthesis and other several mechanisms [16-19]. US particularly the NCLFUS or MIST therapy is a relatively new technique with promising clinical outcomes in superficial and even deep seated soft injuries. The most US machines are set at the frequency of 1 or 3 MHz. Low frequency US and NCLFUS have shown therapeutic effects on periodic wound debridement and bacterial biofilm destruction that indicated their bright perspectives as adjunctive or alternative wound treatment [15, 60, 61]. These techniques because of their capability for focusable and steerable penetration can be used for deep seated or superficial injuries. LLLTs have been reportedly as effective treatments for different wounds and their main mechanism of actions are reducing the inflammatory reaction, enhancing collagen deposition and pain relieving [35, 64, 65]. Considering the different mechanism of action of the above mentioned techniques, using combined approaches of these techniques has been recently appropriate developed seems using and combinations of the techniques can result in more effective treatment with synergistic therapeutic effects. Further controlled studies in this regard should be conducted to develop such combined techniques.

REFERENCES:

1.Hopkins JT, McLoda TA, Seegmiller JG, Baxter GD. Low-level laser therapy facilitates superficial wound healing in humans: a triple-blind, sham-controlled study. Journal of Athletic training. 2004;39[3]:223.

2.Rashidi S, Yadollahpour A, Mirzaiyan M. Low level laser therapy for the treatment of chronic wound: Clinical considerations. Biomedical and Pharmacology Journal. 2015;8[2]:1121-7.

3.Mostafa J, Ali Y, Zohre R, Samaneh R. Electromagnetic Fields and Ultrasound Waves in Wound Treatment: A Comparative Review of Therapeutic Outcomes. Biosci, Biotech Res Asia. 2015;12[Spl.Edn.1]:185-95.

4.Samaneh R, Ali Y, Mostafa J, Mahmud NA, Zohre R. Laser Therapy for Wound Healing: A Review of Current Techniques and Mechanisms of Action. Biosci, Biotech Res Asia. 2015;12[Spl.Edn.1]:217-23.

5.Carey L, Lepley Jr D, editors. Effect of continuous direct electric current on healing wounds. Surgical Forum; 1962.

6.Yadollahpour A, Jalilifar M. Electromagnetic Fields in the Treatment of Wound: A Review of Current Techniques and Future Perspective. J PURE APPL MICROBIO. 2014;8[4]:2863-77.

7.Zahedi M, Yadollahpour A. Therapeutic Effects Of Static Magnetic Fieldsfor Diabetic Wound Healing: A Review Of The Current Evidence. Biosciences Biotechnology Research Asia. 2016;13[1].

8.Henry SL, Concannon MJ, Yee GJ. The effect of magnetic fields on wound healing. Eplasty. 2008;8:40-5.

9.Jing D, Shen G, Cai J, Li F, Huang J, Wang Y, et al. Effects of 180 mT static magnetic fields on diabetic wound healing in rats. Bioelectromagnetics. 2010;31[8]:640-8.

10.Mostafa J, Ali Y, Zohre R, Samaneh R. Electromagnetic fields and ultrasound waves in wound treatment: A comparative review of therapeutic outcomes. Biosciences Biotechnology Research Asia. 2015;12:185-95.

11.Ali Y, Samaneh R, Kavakebian F. Applications of Magnetic Water Technology in Farming and Agriculture Development: A Review of Recent Advances. Current World Environment. 2014;9[3]:695-703.

12.Byl NN, McKenzie AL, West JM, Whitney J, Hunt T, Scheuenstuhl H. Low-dose ultrasound effects on wound healing: a controlled study with Yucatan pigs. Archives of physical medicine and rehabilitation. 1992;73[7]:656-64.

13.Honaker JS, Forston MR, Davis EA, Wiesner MM, Morgan JA. Effects of non contact low-frequency ultrasound on healing of suspected deep tissue injury: a retrospective analysis. International wound journal. 2013;10[1]:65-72.

14.Webster D, Harvey W, Dyson M, Pond J. The role of ultrasound-induced cavitation in the 'in vitro'stimulation of collagen synthesis in human fibroblasts. Ultrasonics. 1980;18[1]:33-7.

15.Ziskin M, McDiarmid T, Michlovitz S. Therapeutic ultrasound. Thermal agents in rehabilitation Philadelphia: FA Davis. 1990;134.

16.Callaghan MJ, Chang EI, Seiser N, Aarabi S, Ghali S, Kinnucan ER, et al. Pulsed electromagnetic fields accelerate normal and diabetic wound healing by increasing endogenous FGF-2 release. Plastic and reconstructive surgery. 2008;121[1]:130-41.

17.Funk RH, Monsees TK. Effects of electromagnetic fields on cells: physiological and therapeutical approaches and molecular mechanisms of interaction. Cells Tissues Organs. 2006;182[2]:59-78.

18.Goudarzi I, Hajizadeh S, Salmani ME, Abrari K. Pulsed electromagnetic fields accelerate wound healing in the skin of diabetic rats. Bioelectromagnetics. 2010;31[4]:318-23.

19. Tepper OM, Callaghan MJ, Chang EI, Galiano RD, Bhatt KA, Baharestani S, et al. Electromagnetic fields increase in vitro and in vivo

angiogenesis through endothelial release of FGF-2. The FASEB journal. 2004;18[11]:1231-3.

20.Yen Patton G, Patton WF, Beer DM, Jacobson BS. Endothelial cell response to pulsed electromagnetic fields: stimulation of growth rate and angiogenesis in vitro. Journal of cellular physiology. 1988;134[1]:37-46.

21.Tao Q, Henderson A. EMF induces differentiation in HL- 60 cells. Journal of cellular biochemistry. 1999;73[2]:212-7.

22.Islamov B, Balabanova R, Funtikov V, Gotovskii YV, Meizerov E. Effect of bioresonance therapy on antioxidant system in lymphocytes in patients with rheumatoid arthritis. Bulletin of experimental biology and medicine. 2002;134[3]:248-50.

23. Tofani S, Cintorino M, Barone D, Berardelli M, De Santi MM, Ferrara A, et al. Increased mouse survival, tumor growth inhibition and decreased immunoreactive p53 after exposure to magnetic fields. Bioelectromagnetics. 2002;23[3]:230-8.

24.Stronati L, Testa A, Villani P, Marino C, Lovisolo G, Conti D, et al. Absence of genotoxicity in human blood cells exposed to 50 Hz magnetic fields as assessed by comet assay, chromosome aberration, micronucleus, and sister chromatid exchange analyses. Bioelectromagnetics. 2004;25[1]:41-8.

25.Murabayashi S, Yoshikawa A, Mitamura Y. Functional Modulation of Activated Lymphocytes by Time varying Magnetic Fields. Therapeutic Apheresis and Dialysis. 2004;8[3]:206-11.

26.Young S, Dyson M. Effect of therapeutic ultrasound on the healing of full-thickness excised skin lesions. Ultrasonics. 1990;28[3]:175-80.

27.Quan K, Shiran M, Watmough D. Applicators for generating ultrasound-induced hyperthermia in neoplastic tumours and for use in ultrasound physiotherapy. Physics in medicine and biology. 1989;34[11]:1719.

28. Ter Haar G, Dyson M, Oakley E. The use of ultrasound by physiotherapists in Britain, 1985. Ultrasound in medicine & biology. 1987;13[10]:659-63.

29.Dyson M, Luke DA. Induction of mast cell degranulation in skin by ultrasound. Ultrasonics, Ferroelectrics and Frequency Control, IEEE Transactions on. 1986;33[2]:194-201.

30.Harpaz D, Chen X, Francis CW, Marder VJ, Meltzer RS. Ultrasound enhancement of thrombolysis and reperfusion in vitro. Journal of the American College of Cardiology. 1993;21[6]:1507-11.

31.Gam AN, Johannsen F. Ultrasound therapy in musculoskeletal disorders: a meta-analysis. Pain. 1995;63[1]:85-91.

32.Beckerman H, Bouter L, Van der Heijden G, De Bie R, Koes B. Efficacy of physiotherapy for musculoskeletal disorders: what can we learn from research? British Journal of General Practice. 1993;43[367]:73-7.

33.Green S, Buchbinder R, Glazier R, Forbes A. Systematic review of randomised controlled trials of interventions for painful shoulder: selection criteria, outcome assessment, and efficacy. Bmj. 1998;316[7128]:354-60.

34.Chromey P. The efficacy of carbon dioxide laser surgery for adjunct ulcer therapy. Clinics in podiatric medicine and surgery. 1992;9[3]:709-19.

35.Posten W, Wrone DA, Dover JS, Arndt KA, Silapunt S, Alam M. Low- level laser therapy for wound healing: mechanism and efficacy. Dermatologic surgery. 2005;31[3]:334-40.

36.Schindl M, Kerschan K, Schindl A, Schön H, Heinzl H, Schindl L. Induction of complete wound healing in recalcitrant ulcers by low- intensity laser irradiation depends on ulcer cause and size. Photodermatology, photoimmunology & photomedicine. 1999;15[1]:18-21.

37.Yu W, Naim J, Lanzafame R. The effects of photo-irradiation on the secretion of TGF and PDGF from fibroblasts in vitro. Lasers Surg Med Suppl. 1994;6[8].

38.Pourreau-Schneider N, Ahmed A, Soudry M, Jacquemier J, Kopp F, Franquin J, et al. Heliumneon laser treatment transforms fibroblasts into myofibroblasts. The American journal of pathology. 1990;137[1]:171.

39. Abergel RP, Meeker CA, Lam TS, Dwyer RM, Lesavoy MA, Uitto J. Control of connective tissue metabolism by lasers: recent developments and future prospects. Journal of the American Academy of Dermatology. 1984;11[6]:1142-50.

40.Leigh IH, Bennett G. Pressure ulcers: prevalence, etiology, and treatment modalities: a review. The American journal of surgery. 1994;167[1]:S25-S30.

41.Phillips T, Stanton B, Provan A, Lew R. A study of the impact of leg ulcers on quality of life: financial, social, and psychologic implications. Journal of the American Academy of Dermatology. 1994;31[1]:49-53.

42.Robinson K. Digby's receipts. Annals of Medical History. 1925;7:216-9.

43.Wainapel SF. Electrotherapy for acceleration of wound healing: low intensity direct current. Arch Phys Med Rehabil 1985. 1985;66:443-6.

44.Prakash A, Pandit P, Sharman L. Studies in wound healing in experimental diabetes. International surgery. 1974;59[1]:25-8.

45.Goodson WH, Hunt TK. Studies of wound healing in experimental diabetes mellitus. Journal of Surgical Research. 1977;22[3]:221-7.

46.Goodson W. Wound healing and the diabetic patient. Surg Gynecol Obstet. 1979;149:600-8.

47.Fahey TJ, Sadaty A, Jones WG, Barber A, Smoller B, Shires GT. Diabetes impairs the late inflammatory response to wound healing. Journal of Surgical Research. 1991;50[4]:308-13.

48. Thawer H, Houghton P. Effects of electrical stimulation on wound healing in diabetic mice. Wound Rep Reg. 1999;7:A332.

49. Thawer H, Houghton P, Kloth L, Butryn A. Effects of electrical stimulation on wound closure in mice with experimental diabetes mellitus. WOUNDS-A COMPENDIUM OF CLINICAL RESEARCH AND PRACTICE. 2000;12[6]:159-+. 50. Reich JD, Tarjan PP. Electrical stimulation of skin. International journal of dermatology. 1990;29[6]:395-400.

51.Weiss DS, Kirsner R, Eaglstein WH. Electrical stimulation and wound healing. Archives of dermatology. 1990;126[2]:222-5.

52.Mulder GD. Treatment of open-skin wounds with electric stimulation. Archives of physical medicine and rehabilitation. 1991;72[6]:375-7.

53.Goodman R, Henderson AS. Some biological effects of electromagnetic fields. Bioelectrochemistry and Bioenergetics. 1986;15[1]:39-55.

54.Murray JC, Farndale RW. Modulation of collagen production in cultured fibroblasts by a low-frequency, pulsed magnetic field. Biochimica et Biophysica Acta [BBA]-General Subjects. 1985;838[1]:98-105.

55.Rodemann HP, Bayreuther K, Pfleiderer G. The differentiation of normal and transformed human fibroblasts in vitro is influenced by electromagnetic fields.Experimental cell research. 1989;182[2]:610-21.

56.Bouzarjomehri F, Sharafi A, Firouzabadi S, Hajizadeh S. Effects of low-frequency pulsed electromagnetic fields on wound healing in rat skin. 2000.

57.Strauch B, Patel MK, Navarro JA, Berdichevsky M, Yu H-L, Pilla AA. Pulsed magnetic fields accelerate cutaneous wound healing in rats. Plastic and reconstructive surgery. 2007;120[2]:425-30.

58.Gann N. Ultrasound: current concepts. Clin Manage. 1991;11[4]:64-9.

59.Dyson M. Mechanisms involved in therapeutic ultrasound. Physiotherapy. 1987;73[3]:116-20.

60.Williams R. Production and transmission of ultrasound. Physiotherapy. 1987;73[3]:113-6.

61.Hekkenberg R, Reibold R, Zeqiri B. Development of standard measurement methods for essential properties of ultrasound therapy equipment. Ultrasound in medicine & biology. 1994;20[1]:83-98.

62.Escandon J, Vivas AC, Perez R, Kirsner R, Davis S. A prospective pilot study of ultrasound therapy effectiveness in refractory venous leg ulcers. International wound journal. 2012;9[5]:570-8.

63.Yao M, Hasturk H, Kantarci A, Gu G, Garcia- Lavin S, Fabbi M, et al. A pilot study evaluating non- contact low- frequency ultrasound and underlying molecular mechanism on diabetic foot ulcers. International wound journal. 2014;11[6]:586-93.

64.Medrado AR, Pugliese LS, Reis SRA, Andrade ZA. Influence of low level laser therapy on wound healing and its biological action upon myofibroblasts. Lasers in surgery and medicine. 2003;32[3]:239-44.

65.Schlager A, Kronberger P, Petschke F, Ulmer H. Low power laser light in the healing of burns: A comparison between two different wavelengths [635 nm and 690 nm] and a placebo group. Lasers in surgery and medicine. 2000;27[1]:39-42.

66.Schlager A, Oehler K, Huebner K-U, Schmuth M, Spoetl L. Healing of Burns after Treatment with 670Nanometer Low-Power Laser Light. Plastic and reconstructive surgery. 2000;105[5]:1635-9.

67.Almeida- Lopes L, Rigau J, Amaro Zângaro R, Guidugli-Neto J, Marques Jaeger MM. Comparison of the low level laser therapy effects on cultured human gingival fibroblasts proliferation using different irradiance and same fluence. Lasers in surgery and medicine. 2001;29[2]:179-84.