

Research Article

Biological activity of *Terminalia arjuna* on Human Pathogenic Microorganisms.

Tariq Javed*¹, Sana Riaz², Muhammad Uzair¹, Ghulam Mustafa³, Ayesha Mohyuddin⁴, Bashir Ahmad Ch¹.

¹ Department of Pharmacy, Bahauddin Zakariya University (BZU), Multan, 60000 Pakistan

² The Women University, Multan, 60000 Pakistan.

³ Lahore Pharmacy College, (LMDC) Lahore, 54000 Pakistan

⁴The University of Management and technology Lahore, Pakistan

ABSTRACT

Received: Aug 11, 2015

Revised: Oct 10, 2015

Accepted: Jan 11, 2016

Online: Jan 27, 2016

World's population relies chiefly on traditional medicinal plants, using their extracts or active constituents. *Terminalia arjuna* of family Combretaceae reported to be effective as aphrodisiac, expectorant, tonic, styptic, anti-dysenteric, sweet, acrid, purgative, laxative, diuretic, astringent, cirrhosis, cardio protective and cancer treatment. In present study, antibacterial, antifungal, brine shrimp lethality and phytotoxic effect of *Terminalia arjuna* was performed. Our results showed that methanolic extract of *Terminalia arjuna* leaves has moderate antifungal effect against *Microsporm canis* and fruit extract possess good antibacterial activity against *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Moreover, Dichloromethane extract of *Terminalia arjuna* bark and fruit possess moderate phytotoxic activity.

Keywords: Antibacterial, Antifungal, Phytotoxicity, *Terminalia arjuna*

INTRODUCTION:

The knowledge of medicinal plants has been accumulated in the course of many centuries based on different medicinal systems such as Ayurveda, Unani and Siddha. In India, it is reported that traditional healers use 2500 plant species and more than hundred plant species serve as regular sources of medicine (Sheng-Ji, 2001). Even in the developed countries, like USA 25% of prescription for pharmaceutical drugs contains one or more substances of plant origin (Dev *et al.*, 1997). A large number of plants with therapeutic properties are quite astonishing. It is estimated that around 70,000 plant species, from lichens to trees, have been used at one time or another for medicinal purposes. (Das *et al.*, 2003). The value of ethno-medicine and traditional pharmacology is gaining recognition because the search for potential medicinal plants is successful if the

plants are chosen on ethno-pharmacological basis. Phytochemical components are important sources of potential pharmacological, chemotherapeutic and toxic activities (Lawal *et al.*, 2005).

The continuous development of antibiotic resistant strains of microbial pathogens, such as methicillin resistant *Staphylococcus aureus* (MRSA), penicillin-resistant *Streptococcus pneumoniae* (PRSP) and vancomycin-resistant enterococci (VRE), is a growing problem and it is therefore, extremely important to discover and develop new antimicrobial compounds (Tally *et al.*, 1999). The antimicrobial compounds from plants may inhibit bacteria through different mechanisms than conventional antibiotics and could therefore be of clinical value in the treatment of resistant microbes (Eloff *et al.*, 1998a). The selection of the plant group for this investigation was based on uses in traditional medicine, since many *Combretum* and *Terminalia* species are well known medicinal plants both in Africa and Asia (Watt and Breyer-Brandwijk, 1962; Hedberg *et al.*, 1982).

Alcoholic extract of *Terminalia arjuna* increased the force of contraction of frog heart (Gupta *et al.*,

*Corresponding Author: Dr. Tariq Javed

Address: Faculty of Pharmacy, Bahauddin Zakariya University, Multan, Pakistan

Phone# +92 32 14960825

e-mail: tjavedpk@gmail.com

1974) and had a marked reduction in total cholesterol level in rabbits (Tiwari *et al.*, 1990). The aqueous extract of *Terminalia arjuna* resulted in dose-dependent decrease in blood pressure (Srivastava *et al.*, 1992) and produced hypotensive effects (Takahashi *et al.*, 1997). Another study reveals that an Injection of bark extract in isolated rabbit heart preparation increased the coronary flow (Bhatia *et al.*, 1998) and tannins of the leaves have anticancer activity (Kandil and Nassar, 1998). The bark powder is reported to exert hypocholesterolaemic and antioxidant effect in humans (Gupta *et al.*, 2001), antimutagenic effect (Kaur *et al.*, 2001) and decrease in total cholesterol and triglycerides (Dwivedi *et al.*, 2000; Dwivedi and Gupta, 2002). While, the bark extract has antioxidant compounds that prevent from oxidative stress (Gauthaman *et al.*, 2001) and to exert free radical scavenging activities in human polymorphonuclear cells (Pawar and Bhutani., 2005). Furthermore, the aqueous extract of *Terminalia arjuna* bark protected the oxidant damage to the liver and kidney following carbon tetrachloride challenge in mice (Manna *et al.*, 2006).

Terminalia arjuna extract prevails significant improvement in diastolic dysfunction (Dwivedi *et al.*, 2002) and exhibit an outstanding antioxidant activity (Sultana *et al.*, 2007). Oral administration of abana, a compound formulation containing *Terminalia arjuna*, 30 mg per tablet, resulted in significant reduction of the systolic blood pressure; echocardiographic left ventricular internal diameter, posterior wall thickness and interventricular septal thickness (Dwivedi *et al.*, 2007). An oleanane triterpenoid, arjunolic acid, exhibited cardiac protective action in isoproterenol-induced myocardial necrosis in rats (Sumitra *et al.*, 2001), has profound effects on human hepatoma cell line (HepG2) for cytotoxicity (Sivalokanathan *et al.*, 2006).

MATERIALS AND METHODS

Plant Material Collection and Extraction

Plant material was collected in the vicinity of Bahauddin Zakariya University, Multan and identified by Dr. Mumtaz Bokhari (Taxonomist), Department of Biological Sciences, Bahauddin Zakariya University, Multan. Different parts (fruit, leaves, bark) of *Terminalia arjuna* were collected and kept under shade for drying. The plant material was ground in a grinding mill and weighed. Extraction from the different parts of *Terminalia arjuna* was carried out by simple maceration process. 100 g of the ground parts of *Terminalia arjuna* was taken in extraction bottle and 250 ml of Dichloromethane (DCM) was added. The mixture was occasionally shaken and homogenized using ultrasonic bath. After 24 hours, the mixture was filtered and the marc was again macerated by the solvent using the same procedure. Extracts of DCM and methanol were concentrated by using the Rota vapor (Buchi-rotavapor R-200) and weighed.

Agar Diffusion Assay

Test sample was dissolved in sterile DMSO to serve as stock solution. Sabouraud dextrose agar was prepared by mixing sabouraud 4% glucose agar in distilled water. Test tubes containing media were autoclaved at 121 °C for 15 minutes. Tubes were then allowed to solidify in slanting position at room temperature. All culture containing tubes were inoculated at optimum temperature of 28-30 °C for growth for 7-10 days. Humidity (40% to 50%) was controlled by placing an open pan of water in incubator. After the incubation for 7-10 days, the test tube with no visible growth of the microorganism was taken to represent the minimum inhibitory concentration (MIC) of the test sample (Okeke *et al.*, 2001; Aneja *et al.*, 2009).

Brine-Shrimp Lethality Assay

Bioactive compounds are often toxic to brine shrimp larvae *Artemia salina* (Leech). Artificial "Sea water" was prepared by dissolving by *ca.* 3.8 g sea salt per liter and filtered. "Sea water" was placed in a small unequally divided tank and

shrimp eggs added to the larger compartment of tank which was darkened by covering it with aluminum foil. After 2 days (when the brine shrimp larvae had matured), added 5 ml Sea water to each vial and added 10 shrimps per vial with the help of Pasteur pipette (30 shrimps per dilution). The vials were maintained under illumination. After 24 hours had elapsed, counted and recorded the number of surviving shrimps, with the aid of a 3 x magnifying glass (Rahman *et al.*, 2001).

Lemna Bioassay for Phytotoxicity

Inorganic medium (E-Medium) was prepared: add KOH pellets to attain pH 5.5-6.0. Prepared vials for testing: 10 vials per dose (500, 50, 5 ppm, control). (a) Weighed 15 mg of compound or extract and dissolved in 15 ml of solvent. (b) Added 1000, 100, and 10 μ l solutions of vials for 500, 40, and 5 ppm; allowed solvent to evaporate overnight. (c) Added 2 ml of E-Medium and then a single plant containing a rosette of the fronds to each vial (Rahman *et al.*, 2001). Vials were placed in a glass dish filled with about 2 cm water, seal container with stopcock grease and glass plate. Placed dish with vials in growth chamber for seven days at 26 °C under fluorescent and incandescent light. Counted and recorded number of fronds per vial on day 3 and day 7.

Results and Discussion

A wide variety of medicinal plants used traditionally have not yet been scientifically investigated against diverse microbial pathogens. Previous reports reveal only few studies on antibacterial and antifungal activity of various parts of *Terminalia arjuna*, such as bark (Kadam and Ghosh, 2008), leaves, root and fruit (Dwivedi *et al.*, 2007; Chouksey *et al.*, 2001). The present study was carried out to validate the antimicrobial potential of *T. arjuna* leaves, fruit and bark against the bacterial and fungal isolates. It was found that methanol extract of *T. arjuna* fruit showed good antibacterial activity against *Staphylococcus aureus*, and *Pseudomonas*

aeruginosa (Table 2). The DCM extract of different plant parts like leaves, bark and fruit showed no antibacterial activity. As it was evident from the biological screening results of methanol extract of *T. arjuna* fruit showed good antibacterial activity against *Staphylococcus aureus* (Gram positive) and *Pseudomonas aeruginosa* (Gram negative) has a potential to combat the bacterial pathogens in human and animals (Table 2). Therefore, substitute the findings of some (Cowan *et al.*, 1999) but concurs with findings of (Aneja *et al.*, 2012)

Furthermore, the samples were tested for antifungal activity of *T. arjuna* and found that methanol extract of *T. arjuna* leaves have moderate activity against *Microsporum canis* and non-significant activity against *Aspergillus flavus*, whereas *T. arjuna* fruit (methanol) extract showed non-significant activity against *Aspergillus flavus*, *Microsporum canis* and *Fusarium solani* (Table 2). *T. arjuna* bark (methanol) showed no antifungal activity. *T. arjuna* leaves, stem bark and fruit (DCM) extract showed no antimicrobial activity (Table 2). The narrow spectrum of antimicrobial activity may be due to active compound present in insufficient amount in the crude extract to show activity with the dose level employed or present in high quantities in the extract exerting antagonistic effects of the bioactive compounds of leaves (Nair *et al.*, 2005). Antimicrobial activity of *T. arjuna* is reported to be present due to of secondary metabolites such as arjunic acid, arjungenin, arjunetin and luteolin (Kadam and Ghosh, 2008; Dwivedi *et al.*, 2007).

Moreover, the samples were also tested for phytotoxicity by using lemna minor bioassay and found that methanol extract of *Terminalia arjuna* showed good phytotoxic activity while leaves and bark (Methanol) showed moderate phytotoxic activity. *Terminalia arjuna* fruit (DCM) showed moderate phytotoxic activity while leaves and bark (DCM) showed no phytotoxic activity. However, the samples tested for brine shrimp lethality bioassay showed no

cytotoxic activity (Table 2). The extraction of whole plant by using with dichloromethane and methanol is shown in Table 1.

CONCLUSION

Terminalia arjuna fruit extract (methanol) has shown good antimicrobial activity against Gram positive and Gram negative bacteria and inhibition is more for Gram positive than Gram negative bacteria. Extensive investigations are required on antimicrobial activity and plausible medicinal applications of phytochemicals. Hence, more studies on this plant are necessary to explore the bioactive principles for therapeutic efficacy.

Tables

Table 1: Extraction of plant material with dichloromethane and methanol

Plant Name	Part used	Solvent	Extract (Wt)
<i>Terminalia arjuna</i>	Leaves (100 g)	Dichloromethane	2.38 gm
		Methanol	14.70 gm
	Bark (100 g)	Dichloromethane	0.40 gm
		Methanol	12.20 gm
	Fruit (100 g)	Dichloromethane	0.30 gm
		Methanol	8.90 gm

Table 2: Results of biological activities of DCM and Methanol extracts of *Terminalia arjuna*

<i>Terminalia arjuna</i> Extract	Antibacterial Activity		Antifungal Activity			Lemena Phytotoxicity Bioassays	Brine shrimp lethality
	<i>Staphylococcus aureus</i> (NCTC # 6571) (Gram-positive)	<i>Pseudomonas aeruginosa</i> (NCTC # 10662) (Gram-negative)	<i>Microsporm canis</i> (ATCC # 11622)	<i>Aspergillus Flavus</i> (ATCC # 32611)	<i>Fusarium solani</i> (ATCC # 9846)		
TALD	-	-	-	-	-	-	-
TALM	-	-	+	-	-	+	-
TABD	-	-	-	-	-	-	-
TABM	-	-	-	-	-	+	-
TAFD	-	-	-	-	-	+	-
TAFM	+	+	-	-	-	+	-

List of Abbreviations

TALD: Dichloromethane extract of *T. arjuna* leaves
 TALM: Methanol extract of *T. arjuna* leaves
 TABD: Dichloromethane extract of *T. arjuna* stem bark
 TABM: Methanol extract of *T. arjuna* stem bark
 TAFD: Dichloromethane extract of *T. arjuna* fruit
 TAFM: Methanol extract of *T. arjuna* fruit

REFERENCES

Aggarwal R and Dutt S. (1936). Chemistry, pharmacology and therapeutic actions of *Terminalia arjuna*. Proc Nat Acad Sci 6, 305.
 Aneja K.R and Joshi R. (2009). Evaluation of antimicrobial properties of fruit extracts of *Terminalia chebula* against dental caries pathogens. Jundishapur J Microbiol 2: 105-111.
 Aneja K.R, Sharma, C and Joshi, R. (2012). Antimicrobial activity of *Terminalia arjuna* Wight & Arn.: An ethnomedicinal plant against pathogens causing ear infection. Brazilian Journal of otorhinolaryngology 78: 68-74.
 Atta-ur-Rahman, Choudhary M.I and Thomsen W.J. (2001). Bioassay techniques for drug development, Vol 16 (Harwood academic publishers The Netherlands).

- Bhatia J. (1998). Study of the possible cardioprotective role of *Terminalia arjuna* in experimental animals and its clinical usefulness in coronary artery disease. MD (Pharmacology) thesis, India: University of Delhi.
- Chouksey B and Srivastava S. (2001). New constituent from the roots of *Terminalia arjuna*: Antifungal agent. Indian Journal of chemistry section B 40: 354-356.
- Cowan M.M (1999). Plant products as antimicrobial agents. Clinical microbiology reviews 12: 564-582.
- Das P.N, Purohit S, Sharma A and Kumar T. (2003). A handbook of medicinal plants. Jodhpur, India, Agrobios 118.
- Dev S. (1997). Ethnotherapeutics and modern drug development: the potential of Ayurveda. Current science Bangalore 73: 909-928.
- Dwivedi S. (2007). *Terminalia arjuna* Wight & Arn. A useful drug for cardiovascular disorders. Journal of ethnopharmacology 114: 114-129.
- Dwivedi S and Gupta D. (2002). Efficacy of *Terminalia arjuna* in chronic stable angina. Indian heart journal 54, 441; author reply 441.
- Dwivedi S, Gupta D, Sharma K, Kumar S, Kukreja A, Dwivedi S, and Singh A. (2000). Modification of coronary risk factors by medicinal plants. Paper presented at: Journal of Medicinal and Aromatic Plant Sciences (Central Institute of Medicinal and Aromatic Plants).
- Eloff J. (1998). Which extractant should be used for the screening and isolation of antimicrobial components from plants? Journal of ethnopharmacology 60: 1-8.
- Gauthaman K, Maulik M, Kumari R, Manchanda S, Dinda, A and Maulik S. (2001). Effect of chronic treatment with bark of *Terminalia arjuna*: a study on the isolated ischemic-reperfusion rat heart. Journal of ethnopharmacology 75, 197-201.
- Gupta L. (1974). Studies on cardiac muscle regeneration under the influence of certain indigenous drugs (Ph. D. thesis, Banaras Hindu University, Varanasi, India).
- Gupta R, Singhal S, Goyle A and Sharma V. (2001). Antioxidant and hypocholesterolaemic effects of *Terminalia arjuna* tree-bark powder: a randomised placebo-controlled trial. The Journal of the Association of Physicians of India 49:231-235.
- Hedberg I, Hedberg O, Madati P.J, Mshigeni K.E, Mshiu E, and Samuelsson G. (1982). Inventory of plants used in traditional medicine in Tanzania. I. Plants of the families Acanthaceae-Cucurbitaceae. Journal of ethnopharmacology 6, 29-60.
- Kadam U, and Ghosh S. (2008). Antibacterial principles from the bark of *Terminalia arjuna*. Current Science 94, 27.
- Kandil F.E and Nassar M.I. (1998). A tannin anti-cancer promotor from *Terminalia arjuna*. Phytochemistry 47 :1567-1568.
- Kaur S, Grover I.S and Kumar S. (2001). Antimutagenic potential of extracts isolated from *Terminalia arjuna*. Journal of environmental pathology, toxicology and oncology 20.
- Lawal M, Wasagu R, and Ladan M. (2005). Hepatotoxicity risk assessment of neem (*Azadirachta indica*) seed extract using albino rats. Biological and Environmental Sciences Journal for the Tropics 2: 36-38.
- Manna P, Sinha M and Sil P.C. (2006). Aqueous extract of *Terminalia arjuna* prevents carbon tetrachloride induced hepatic and renal disorders. BMC Complementary and Alternative Medicine 6, 33.
- Nair R, Kalariya T, and Chanda S. (2005). Antibacterial activity of some selected Indian medicinal flora. Turk J Biol 29: 41-47.
- Okeke M, Iroegbu C, Eze E, Okoli, A and Esimone C. (2001). Evaluation of extracts of the root of *Landolphia owerrience* for antibacterial activity. Journal of ethnopharmacology 78: 119-127.
- Pawar R and Bhutani K. (2005). Effect of oleanane triterpenoids from *Terminalia arjuna*, A cardioprotective drug on the process of respiratory oxyburst. Phytomedicine 12: 391-393.
- Sheng Ji P. (2001). Ethnobotanical approaches of traditional medicine studies: some experiences from Asia. Pharmaceutical biology 39: 74-79.
- Sivalokanathan S, Vijayababu M.R and Balasubramanian M.P. (2006). Effects of *Terminalia arjuna* bark extract on apoptosis of human hepatoma cell line HepG2. World journal of gastroenterology: WJG 12: 1018-1024.
- Srivastava R, Dwivedi S, Sreenivasan K and Chandrashekar C. (1992). Cardiovascular effects of *Terminalia* species of plants. Indian drugs 144.
- Sultana B, Anwar F, and Przybylski R. (2007). Antioxidant activity of phenolic components present in barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. trees. Food Chemistry 104: 1106-1114.
- Sumitra M, Manikandan P, Kumar D.A, Arutselvan N, Balakrishna K, Manohar B.M, and Puvanakrishnan R. (2001). Experimental myocardial necrosis in rats: role of arjunolic acid on platelet aggregation, coagulation and antioxidant status. Molecular and Cellular Biochemistry 224: 135-142.
- Takahashi S, Tanaka H, Hano Y, Ito K, Nomura T, and Shigenobu K. (1997). Hypotensive effect in rats of hydrophilic extract from *Terminalia arjuna* containing tannin-related compounds. Phytotherapy research 11: 424-427.
- Tally F.P. (1999). Researchers reveal ways to defeat 'superbugs'. Drug discovery today 4: 395-398.
- Tiwari A, Gode J and Dubey G. (1990). Effect of *Terminalia arjuna* on lipid profiles of rabbits fed hypercholesterolemic diet. Pharmaceutical biology 28: 43-47.
- Watt J.M, and Breyer-Brandwijk M.G. (1962). The Medicinal and Poisonous Plants of Southern and Eastern Africa: Being an Account of Their Medicinal and Other Uses, Chemical Compositions, Pharmacological Effects and Toxicology in Man and Animal (Livingstone).