RESEARCH ARTICLE

Exploitation of green biomass for the preparation of leaf protein concentrates

Wadaskar SB1*, Manwatkar VG2 and Gogle DP3

¹Department of Botany, G.B.TathaTatyasahebKhare Commerce, Parvatibai Gurupad Dhere Arts &

Shri. M. J. Bhosale Science College, Guhagar, Dist.:-Ratnagiri-415703.

²Department of Botany, VidyaVikas Arts, Commerce and Science College, Samudrapur, Dist-Wardha-442305. ³PGTD of Botany, Rashtrasant Tukadoji Maharaj Nagpur University, Nagpur-440033 *Corresponding Author Email: <u>sai.wadaskar@gmail.com</u>

Manuscript details:

ABSTRACT

Available online on http://www.ijlsci.in

ISSN: 2320-964X (Online) ISSN: 2320-7817 (Print)

Editor: Dr. Arvind Chavhan

Cite this article as:

Wadaskar SB, Manwatkar VG and Gogle DP (2015) Exploitation of green biomass for the preparation of leaf protein concentrates, Int. j. of Life Sciences, Special issue A3: 109-114.

Copyright: © Author, This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derives License, which permits use and distribution in any medium, provided the original work is properly cited, the use is noncommercial and no modifications or adaptations are made. The available food resources are always remaining to be constrained for the ever increasing population. Hence the non-conventional food resourced is to be considered in the coming year. The present study includesgreen foliages from 7 plants to prepare leaf protein concentrate viz., Berseem (*Trifoliuma lexandriumL.*), *Alysicarpus vaginalis* L. var. stocksii., *Alternanthera paronychioides* St. Hil., Cabbage (*Brassica oleracea* L. var. capitata), Radish (*Raphanus sativusL.*), Adulsa (*Adhatoda vasica* Nees.), and Bauchi (*Psoralia corylifolia* L.). The selected plants were fractionated and subjected to prepare LPC as suggested by Pirie (1971). The maximum yield of juice and fiber are reported in green foliages of *Brassica oleracea* L. (550 ml/Kg) and *Alysicarpus vaginalis* L.(540 g/Kg) respectively. Maximum dry weight of juice and 70.22 g respectively) Maximum dry weight of fiber was reported in green foliage of *Alhatoda vasica* Nees. (120 and 70.22 g respectively) Maximum dry weight of fiber was reported in green foliages of *Psoralia corylifolia* L. (80.52 g).

Keywords: leaf protein concentrates, non-conventional food resources, DPJ, LPC, fiber, biomass.

INTRODUCTION

In our country huge amount of green biomass are available in the form of weeds, fodder crops and by-products of the vegetable material. The utilization of this material can be enhanced for the benefit of human being by the extracting available protein in this green biomass. To extract proteins from green leaves numerous technologies have been developed over last 50 years. For this purpose, a technique of fractionation has been proposed by Pirie from United Kingdom. This technique of fractionation recommended by Pirie (1942) has now become popular as "Green Crop Fractionation (GCF)" which involves the separation of proteins from the indigestible fibrous material of leaves.

The process of GCF is basically consists of mechanical operations of grinding and pressing, which enable the fresh green leafy foliages to be separated into two fractions; a protein rich juice and the pressed crop residue

(PCR). For this purpose the crop is macerated with IBP pulper (Davys and Pirie, 1969a) or with simple grinder mixer and then pressed with IBP press (Davys et al.,1969b) or by hand pressing method. After thorough maceration of the crop, almost all cells in it are damaged and the proteins get liberated in the juice expressed after pressing the pulp. After releasing the juice a fibrous material left behind called as pressed crop which is also nutritionally sufficient to the cattle. The juice or leaf extract contain, along with the proteins, sugars, lipids, vitamins and other soluble components of the cell protoplasm. When this fraction is either heated or acidified, a green curd is produced due to the precipitation of protein. This protein rich curd is referred as leaf protein concentrates (LPC). The LPC can be separated from remaining part of the juice, known as deproteinized juice (DPJ) by filtration through a simple cotton or muslin cloth. Thus, the process of GCF results into four fractions i.e. Leaf extract or juice, Pressed crop residue (PCR) or Fiber, Leaf protein concentrate (LPC), Deproteinized juice (DPJ). All these four products can be used in different ways as suggested by several workers.

First suggestion on the use of protein extracted from leaves was made by Ereky (1927). Slade (1937) argued that the use of protein from grass was more economical than meat obtained after feeding the grass to animals. Chibnall (1939) and his co-workers provided information on the quantity and properties of protein in leaves.

Pirie (1942) put forth the real potential of leaf protein (LP) and its use as human food in during II World War. After two decades of war several workers took interest in leaf protein (Davies *et al.*, 1952; Carpenter *et al.*, 1954; Mcdonald, 1954; Tilley *et al.*, 1954; Anandaswamy and Date, 1956; Cowlishaw *et al.*, 1956; Raymond and Harris, 1957; Guha, 1960; and Chayen *et al.*, 1961).

Byers (1961) evaluated the extractability of leaf protein from leaves of 60 tropical spp. growing in Ghana. His results showed that some common weeds offered promising results, yielding more protein than the leaves of many crops with good quality. Devi *et al.* (1964) studied the isolation and composition of leaf protein from certain species of Indian flora and reported that some of the proteins isolated are potentially useful for supplementation of cereal diets deficient in lysine and methionine. Carlsson and Clarke (1983) studied the suitability of *Atriplexhortensis* L. as a source of leaf protein concentrate and showed that it could be utilized for the preparation of LPC as it was found superior to *Spinaceaoleracea* L. The cultivation of berseem for the measurement of LP shown that it could give a yield of 700kg per hectare extractable protein in 140 days (Mungikar, 1974; Tekale, 1975; Mungikar *et al.*, 1978; Mungikar *et al.*, 1976b; Patil and Mungikar, 1992). The yield of extractable protein from leaves taken at the time of harvest of the edible part (root, tuber) from brassicas, beet root, turnip and radish ranged between 76 to 171kg/ha (Tekale, 1975; Deshmukh*et al.*, 1974; Tekale and Joshi,1976; Giri and Nagpal, 1984; Giri *et al.*, 1983). Leafy vegetables were also found suitable for producing good quality of leaf protein concentrates (Mungikar and Ajaykumar, 1995).

MATERIALS AND METHODS

Selection of plants: For the present work green foliages from 7 plants were selected to prepare leaf concentrate viz., Berseem (Trifolium protein alexandriumL.), Alysicarpus vaginalis L. var. stocksii., Alternanthera paronychioides St. Hil., Cabbage (Brassica oleracea L. var. capitata), Radish (Raphanus sativus L.), Adulsa (Adhato davasica Nees.), and Bauchi (Psoralia corylifolia L.). These plant material were authentified at Department of Botany, RTM Nagpur University, Nagpur. These plants were collected from different places. Berseem was collected from Walu Sangopan Kendra, Nagpur. Bauchi was collected from Krishi Vidyapith Campus, Nagpur. Cabbage and Radish were collected from local vegetable market. Alternanthera, Alysicarpus, and Adhatoda were collected from RTM Nagpur University campus, Nagpur.

Preparation of leaf protein concentrates: The selected plants were fractionated and subjected to prepare LPC as suggested by Pirie (1971). These plants were first washed well with water and pulped with grinder mixer. The juice was then expressed by hand pressing method. The amount of juice and fiber obtained per Kg of green foliages was recorded. The juice obtained was employed for the preparation of LPC. LPC was prepared by heat coagulation method.

Heat coagulation method: For this purpose, a sample of 100 ml juice was slowly added to 20 ml boiling water with continuous stirring, as a result proteins in juice coagulated resulting into green colour curd called as leaf protein concentrate (LPC). During whole process the temperature was maintained at 95°C. This

heated juice was then filtered through preweighed Whatmann filter paper No.1. During filtration the yellowish filtrate was obtained which is called as deproteinized leaf juice (DPJ) or whey. The green coloured curd (LPC) along with filter paper and DPJ was dried at 55°C in hot air oven. The amount of the dried LPC and DPJ was recorded per Kg of fresh green foliages.

RESULT AND DISCUSSION

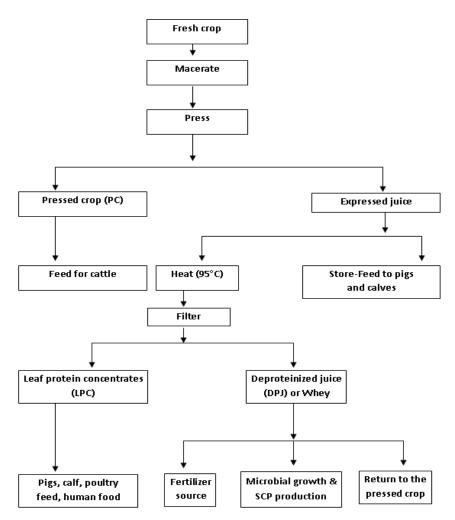
I. Fresh weight of fiber and amount of juice from different plants obtained during green crop fractionation.

The yield of juice and fibers of various plants obtained during GCF are summarized and shown in table 1 and fig. 1a, 1b respectively. The maximum yield of juice and fiber are reported in green foliages of *Brassica oleraceaL.* (550 ml/Kg) and *Alysicarpus vaginalis* L.(540 g/Kg) respectively, whereas, the

minimum yield of juice and fiber are reported in green foliages of *Adhatoda vasica* Nees. (217 ml/Kg) and *Brassica oleracea* L. (250 g/Kg) respectively.

During present investigation 7 plants were considered for fractionation. From these, two are vegetable crops (radish and cabbage), a fodder crop (berseem) and four wild plants (Adhatoda vasica Nees., Psoralia corylifolia L., Alysicarpus vaginalis L., and Alternanthera paronychioides St. Hil.). The green foliages (leaves and tender stem) of these plants were employed for the preparation of LPC. Though the foliages of these wild plants are not used as source of vegetables, however, the young shoot part of Psoralia Alysicarpus corylifolia L., vaginalis L., and Alternanthera paronychioides St. Hil. were generally consumed by animals and Adhatoda vasica Nees.is well known for its medicinal value. A large variation in both juice as well as fiber content was observed due to their time ofharvesting and vegetative growth. The coefficient of variation of the obtained values of juice and fiber was 35.60 and 30.81% respectively.

PROCESS OF GREEN CROP FRACTIONATION (GCF)



Sr. No.	Name of the plant's	Yield of wet fractions of plant materials.		
		Juice (ml / Kg)	Fiber (g / Kg)	
1	Alysicarpus vaginalis L.	265	540	
2	Trifolium alexandrium L.	390	354	
3	Alternanthera paronychioidesSt. Hil.	407 343		
4	Raphanus sativus L.	520	258	
5	Brassica oleracea L.	550	250	
6	Psoralia corylifolia L.	252	494	
7	Adhatoda vasica Nees.	217	508	
-	Mean	372	392	
-	Std. Deviation	132	121	
-	Std. Error	50.00	45.70	
-	Coefficient of variation	35.60%	30.81%	

Table 1: Fresh weight of fiber and amount of	iuice from various	plants obtained durin	g green crop fractionation
Table 1. Tresh weight of fiber and amount of	juice monin various	plants obtained durin	6 Siccil ci op nachonahon.

Table 2: Dry weight of various fractions of plants obtained during green crop fractionation.

Sr.	Name of the plant's	Dry weight of fractions, g / Kg of fresh green foliages.			
No.		Juice	Fiber	LPC	DPJ
1	Alysicarpus vaginalis L.	118.00	305.00	71.15	38.95
2	Trifolium alexandrium L.	62.75	88.00	31.64	23.36
3	Alternanthera paronychioides St. Hil.	115.00	94.74	72.41	31.42
4	Raphanus sativus L.	53.40	80.00	20.83	17.83
5	Brassica oleracea L.	62.23	68.34	21.51	36.33
6	Psoralia corylifolia L.	117.24	160.30	80.52	30.53
7	Adhatoda vasica Nees.	120.00	175.00	42.42	70.22
	Mean	92.70	139.00	48.60	35.50
	Std. Deviation	31.20	84.00	25.60	16.90
	Std. Error	11.80	31.80	9.67	6.40
	Coefficient of variation	33.71%	60.53%	52.58%	47.65%

The fresh green foliages from all these plants were fractionated for the measurement of the yield of juice (fresh and dried), fiber (fresh and dried), LPC (dried) and DPJ (dried) per unit weight of foliage. The fractionation of foliages was undertaken using mechanical mixer for pulping and hand pressing method was employed for expressing juice. To justify the usefulness of wild plants in green crop fractionation process, the yields of the wild plants have been compared with *Brassica oleraceaL*. since this plant have been extensively utilized for the green crop fractionation process by various workers.

The result obtained showed that the extractability of juice and fiber varied from species to species. The

results also showed that the quantity of juice is inversely proportional to the quantity of fiber; if the one fraction was more than other would be low or vice versa. As far as yield of expressed juice is concerned, all the wild plants have relatively low extractability rate than that of *Brassica oleraceaL*. (table 1). The variation in extractability of juice might be due to nature of the foliages and their moisture content.

Patil and Salve (2000) has been reported the yield of juice for radish as 584ml/ Kg, (fresh wt. basis) respectively. Gogle (2000) mentioned the yield of juice from cabbage and radish as 628 and 711ml/Kg fresh material, respectively. However, in present study, the yield of juice from cabbage and radish was found 550 and 520ml/Kg respectively. This variation in extractability of juice might be due to equipment used for expressing the juice, stage of harvesting and also because of difference in region. In present study the juice was expressed by hand press method.

Yield of fiber from all wild plants are comparatively higher than that of *Brassica oleracea*L. (table 1). Bhande and Mungikar (1990) reported the yield of fiber from lucerne as 426g/Kg (fresh wt. basis), whereas in present study it ranged from 250 to 540g/Kg, fresh material. The yields of fiber from various crops are depends upon their maturity level and also it varied from specie to species.

II. Dry weight of various fractions of plants obtained during green crop fractionation (g/Kg, fresh green foliages).

The results obtained for the various dried fractions are represented and illustrated in table 2 and fig. 2 respectively. Maximum dry weight of juice and DPJ was reported in green foliage of *Adhatoda vasica* Nees. (120 and 70.22 g respectively) and minimum in green foliage of *Raphanus sativus* L. (53.4 and 17.83g respectively). Maximum dry weight of fiber was reported in green foliage of *Alysicarpus vaginalis* L. (305 g) and minimum in *Brassica oleracea* L. (68.34 g). Maximum dry weight of LPC was reported in green foliages of *Psoralia corylifolia* L. (80.52 g) and minimum in *Raphanus sativus* L. (20.83 g).

The yield of various fractions from wild plants was compared with *Brassica oleracea*L. to check their usefulness in green crop fractionation process. The present results revealed that the dry weight of juice, fiber and LPC of all wild plants are relatively higher than that of *Brassica oleracea*L. The higher yield of LPC in wild plants may be attributed to high crude protein content in the juice extracted from their foliages. In general, leguminous plants gave better yield of LPC. However, the yield of DPJ from *Alysicarpus vaginalis* L. and *Adhatoda vasica* Nees. was found higher than that of *Brassica oleracea*L., but it was lower in *Alternanthera paronychioides* St. Hil. and *Psoralia corylifolia* L.

Several workers have been reported the yield of LPC from radish as 40.9 g/Kg (Patil and Salve, 2000), 23.77 g/Kg (Gogle, 2000) and 20.86 g/Kg (Madhekar, 2008). Gogle (2000) have been reported the yield of LPC from cabbage as 43.94 g/Kg. While, in present investigation radish and cabbage showed LPC yield as 20.83 and 21.51 g/Kg respectively. This variation in

LPC yield might be due to equipment, regional difference and the duration of processing. Singh (1969) suggested that a plant can be selected for the preparation of LPC, when the yield of dry LPC exceed 10 g/Kg fresh green foliage and the resulting LPC contains more than 5% nitrogen. In this view all plants species which is chosen in present study could be considered suitable for the preparation of LPC.

CONCLUSION

The present results revealed that the yield of expressed juice from all the wild plants was relatively low as compared to *Brassica oleraceaL.*; among all these wild plants *Adhatoda vasica* Nees. yielded minimum amount of juice. However all wild plants show comparatively more yield of fiber than that of *Brassica oleraceaL*.

Similarly the dry weight of juice, fiber, LPC and DPJ of all wild plants are relatively higher than that of *Brassica oleracea*L. except *Alternanthera paronychioides* St. Hil. and *Psoralia corylifolia* L. which showed lower yield of dry DPJ.

Thus from the present study it is concluded that the green biomass can be successfully used for the extraction of protein and the preparation of leaf protein concentrate. However, the studies regarding its nutritional aspects are to be considered in future.

REFERENCES

- Anandaswamy B and Date WB (1956) Bull.Cent. Fd. Technol. Res. Inst., 5: 105.
- Bhande SD and Mungikar AM (1990) Distribution of dry matter and crude protein in pressed crop and expressed juice obtained during fractionation of lucerne. *Res. J. Pl. Environ.*, 6 (2): 37-42.
- Byers M (1961) Extraction of protein from the leaves of some plants growing in Ghana. *Journal of the Science of Food and Agriculture*, 12: 20-30.
- Carlsson R and Clarke EMW (1983) *Atriplexhortensis* L. as a leafy vegetable and as a leaf protein concentrate plant.*Qual Plant Plant Foods Human Nutr.*,33: 127-133.
- Carpenter KJ, Duckworth J and Ellinger GM (1954) Proc. European Grassland Conf. Paris, pp. 243.
- Chayen IH, Smith RS, Tristram GR, Thirkell D and Webb T (1961) The isolation of leaf components.*J. Sci. Fd. Agric.*, 12: 502.
- Chibnall AC (1939) *Protein metabolism in the plant*". Yale Univ. Press.New Haven.
- Cowlishaw SJ, Eyles DE, Raymond WF and Tilley JMA (1956) Nutritive value of leaf protein concentrates. I.

Effect of addition of cholesterol and amino-acids. *J. Sci. Fd. Agric.*, 7: 768.

- Davies Davy, M, Evans WC and Parr WH (1952) Biological values and digestibilities of some grasses, and protein preparations from young and mature species, by the Thomas-Mitchell method, using rats.*Biochem. J.*, 52: xxiii.
- Davys MNG and Pirie NW (1969a) A laboratory-scale pulper for leafy plant material. *Biotechnol. Bioeng.*, 11: 517-528.
- Davys MNG, Pirie NW and Street G (1969b) A laboratory scale press for extracting juice from leaf pulp. *Biotechnol.Bioeng.*,11: 528-529.
- Deshmukh MG, Gore SB, Mungikar AM and Joshi RN (1974) The yields of leaf protein from various shortduration crops *J. Sci. Fd. Agric.*, 25: 717.
- Devi AV, Rao NAN and Vijayaraghavan PK (1964) Isolation and composition of leaf protein from certain species of Indian flora. *Journal of the Science of Food and Agriculture*, 16 (2): 116-120.
- Ereky K (1927) Process for the manufacture and preservation of green fodder pulp or other plant pulp and of dry products made there from. Brit. Pat. 270629.
- Giri P and Nagpal V (1984) In "*Current trends in life sciences*", Vol. XI, Progress in leaf protein research (N. Singh Ed.), pp. 35-39. Today and Tomorrow's Printers and Publishers, New Delhi.
- Giri P, Putan S and Nagpal V (1983) Indian Bot. Reptr., 2: 147.
- Gogle DP (2000) "Studies on various products obtained during green crop fractionation". Ph.D. Thesis, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad.
- Guha BC (1960) Leaf protein as a human food. *Lancet*, 1: 705.
- Madhekar RD (2008) Preperation of leaf protein concentrates (LPCs) from green leafy vegetables. *Bioinfolet*, 5 (3A): 177-180.
- McDonald ANC (1954) Some experiments on the mechanical extraction of leaf protein. *Natl. Inst.Agr.Eng., Rep.* 43: 30.
- Mungikar AM and Ajaykumar K (1995) Dr. B. A. M. U. J. Sci. 28 (Sci. 21): 8.
- Mungikar AM, Batra UR, Tekale NS and Joshi RN (1976b). Effects of nitrogen fertilization on the yields of extracted protein from some crops. *Expl. Agric.*, 12: 353.
- Mungikar AM, Prasad VL, Rangnekar DV and Joshi AL (1978) Studies on preservation of lucerne juice. Presented at the 21st Veterinary Conference, Cuttack.
- Mungikar AM (1974) *Agronomic studies in Leaf protein production IV.* Ph.D. Thesis, Marathwada University, Aurangabad.
- Patil PR and Mungikar AM (1992). Geobios, 19: 58-61.
- Patil VS and Salve US (2000) Preparation of leaf protein concentrate from dark green leafy vegetables.

Proc.Plant resource development.Ed. Mungikar, A. M. and Bhuktar, A. S. pp. 194-201.

- Pirie NW (1942) Green leaves as a source of proteins and other nutrients. *Nature*, 149: 251.
- Pirie NW (1971) "The machinery for LPC production." in Leaf Protein: its agronomy, preparation, quality and use", Pirie, N. W. Ed., IBP Handbook No. 20, (Oxford, United Kingdom: Blackwell's Scientific Publishers), pp. 56-62.
- Raymond WF and Harris CE (1957) The value of the fibrous residue from leaf-protein extraction as a feeding stuff for ruminants. *J. British Grassland Society*, 12: 166.
- Singh N (1969) Leaf protein research in India.J. Fd. Sci. Technol., 6: 165.
- Slade RE (1937) *Grass and the national food supply*. A. Rep. Br. Assn. Advmt. Sci., pp. 457-473.*
- Tekale NS and Joshi RN (1976) Extractable protein from by-product vegetation of some kale and root crops.*Ann. Appl. Biol.*, 82: 155-157.
- Tekale NS (1975) *Agronomic studies on leaf protein production V.* Ph. D. Thesis, Marathwada University, Aurangabad.
- Tilley JMA, Barnes ML and Raymond WF (1954) *Fm. Machanig.*, 6: 487.

© 2015 | Published by IJLSCI