

Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Disease

journal homepage: www.elsevier.com/locate/apjtd



Document heading

doi:10.1016/S2222-1808(14)60575-2

© 2014 by the Asian Pacific Journal of Tropical Disease. All rights reserved.

Effect of henna and roselle extracts on pathogenic bacteria

Rafat Khalaphallah¹, Wagdi Saber Soliman²

¹Botany Agriculture Department, Faculty of agriculture, South Valley University, Qena, Egypt

PEER REVIEW

Peer reviewer

Dr. Mohamed Abou-Ellail, Cytology and Genetics Department, Genetic Engineering and Biotechnology Division, National Research Center, 12622, Al-Bohos st., Dokki, Cairo, Egypt.

Tel: 002 0100 3269386 E-mail: abou_ellail@yahoo.com

Comments

The study is good whereas the authors isolated the pathogens from wastewater and they tested the antimicrobial activity of water and ethanol extracts from henna and roselle against some pathogens. The results revealed that ethanolic extract had more antimicrobial activity than water extracts. Moreover, the ethanolic extracted from roselle showed highest antibacterial activity against all tested pathogens than the ethanolic extracted from henna.

Details on Page 295

ABSTRACT

Objective: To investigate the antibacterial effects of water and ethanolic extracts of henna leaves and roselle calyxes against pathogenic bacteria isolated from domestic wastewater.

Methods: The antimicrobial activity was determined in the extracts using agar disc diffusion method. The antibacterial activities of extracts (2.5%, 5.0% and 10.0% w/v) of both henna and roselle were tested against one Gram-positive *Bacillius subtilis*; two Gram-negative *Escherichia coli, Pseudomonas aeruginosa* human pathogenic bacteria.

Results: Ethanolic extracts had more antimicrobial activity than water extracts. Ethanolic extract of roselle had the highest antibacterial activity against all tested organisms, followed with ethanolic extract of henna. *Pseudomonas aeruginosa* was the most sensitive bacteria to plant extracts.

Conclusion: The results of this study suggested that roselle contains more phyto-chemicals with antimicrobial activity than henna on the bacteria strains under study, and these phyto-chemicals were more effective when extracted by ethanol rather than water.

KEYWORDS

Antimicrobial activity, Henna, Roselle, Bacillius subtilis, Escherichia coli, Pseudomonas aeruginosa

1. Introduction

Henna, *Lawsonia alba* Lam. (*L. alba*), is a medicinal plant belonging to the family Lythraceae. Powdered leaves of henna is commonly used as cosmetic for staining palm, hands, hairs and other body parts^[1,2]. Lawsone, 2–hydroxy 1,4–naphthoquinone, is the chief coloring component of henna leaves which acts as a substantive dye for keratin and imparts orange color due to the presence of hydroxyl group in naphthoquinone structure^[3,4]. Henna is found

to have several pharmacological uses such as antitumor, anthelmintic, antioxidant, immunomodulatory, burn wound healing, UV protective, and antimicrobial properties[5–9]. Henna is a natural product with low health risk potential[10–12]. Leaves of henna contain major phyto–chemicals such as glycosides, phytosterol, steroids, saponins, tannins and flavonoids. Flavonoids and glycosides are commonly known to posse antimicrobial activity[13].

Roselle (*Hibiscus sabdariffa* L.) (*H. sabdariffa*) belongs to the Malvaceae family^[14]. Roselle calyx is a famous source

*Corresponding author: Rafat Khalaphallah, Botany Agriculture Department, Faculty of Agriculture, South Valley University, Qena 83523, Egypt.

Tel: +20 965 211 835

E-mail: r.shipat@agr.svu.edu.eg

Article history: Received 10 May 2014

Received 10 May 2014 Received in revised form 25 May, 2nd revised form 30 May, 3rd revised form 1 Jun 2014 Accepted 10 Jun 2014

Available online 27 Jun 2014

²Faculty of Agriculture and Natural Resources, Aswan University, Aswan, Egypt

of anthocyanins which are the largest group of water—soluble pigments in plants. Anthocyanins are highly appreciated in the food industry for their coloring properties, which can give foods various hues of red and violet[15–17]. Also, there are several reports on the benefits of Roselle calyxes as an excellent source of natural antioxidants[18]. The extracts of calyx are also used in folk medicine against many complaints that include high blood pressure, liver diseases and fever[19,20]. In addition to, roselle extracts are reported to have antimicrobial activity against different pathogenic microorganisms[21–23].

Bacteria are the most abundant group of organisms, and bacterial pathogens are the major source of disease and mortality to human populations, worldwide^[24]. Treatment by antibiotics to face such pathogens is facing major challenges as bacteria are continued to evolve resistance to the synthesized antibiotics^[25–27]. Nowadays, with taking into the account the side effects of chemical antibiotics, the use of plant extracts as pharmaceutical purposes is increased. In this study, we investigate the antimicrobial activity of henna and roselle extracted by two types of solvent (water and ethanol) against different pathogenic bacteria; *Bacillius subtilis* (*B. subtilis*), *Escherichia coli* (*E. coli*), and *Pseudomonas aeruginosa* (*P. aeruginosa*).

2. Materials and methods

2.1. Plant materials and extractions preparation

The plant materials used in this study were leaves of henna (L. alba) and calyxes of roselle (H. sabdariffa). The plant materials were collected from plants grown in the Agricultural Experimental Farm, Faculty of Agriculture, South Valley University, Qena, Egypt. The leaves and calyxes were left to dry at room temperature for 48 hours. The dried materials, henna leaves and roselle calvxes, were ground to a fine powder with an electric mill, then the materials were sterilized under UV-lighting for an hour. Two types of extract were prepared in this study; alcoholic and water-based extracts. The alcoholic extracts were prepared by mixing plant materials with 95% ethanol for four hours at concentrations of 2.5%, 5.0% and 10% (w/v). The extracts were filtered through filter paper (Whatmann No. 1) and the filtrates were collected for further using. Water based henna extract was prepared in the same way except that distilled water was used instead of alcohol. All tools used for the extraction were sterilized to avoid contamination.

2.2. Pathogenic bacteria

Microbial strains of *B. subtilis* (Gram positive), *E. coli*, and *P. aeruginosa* (Gram negative) were isolated from wastewater and used throughout the study. Wastewater samples were collected from South Valley University. The microorganism species were isolated and grown in nutrient broth medium containing (peptone 10 g/L and yeast extract 5 g/L at 37 °C). The cells were grown in a 100 mL Erlenmeyer flask in a shaking incubator at 300 r/min at 37 °C.

2.3. Antimicrobial assay

The antimicrobial activity of investigated extracts was determined using Agar diffusion dilution method. Nutrient agar was used with different diluted extract concentrations (10–0.03 $\mu g/mL$). 0.1 mL containing 10 5 CFU/mL (0.5 McFarland) was spread on the agar as described in[28,29]. Discs of 5 mm in diameter were made using filter paper Whatmann No. 1. The discs were soaked in the prepared extractions for two hours, and then two discs were applied in Petri dish and incubated for 48 h at 37 °C. After the incubation period, the antimicrobial activity was evaluated by measuring the diameter of inhibition zone including the disc.

2.4. Statistical analysis

The statistical differences were tested by analysis of variance (ANOVA) for each treatment. The analysis was carried out using JMP (version 4; SAS Institute, Cary, NC, USA).

3. Results

3.1. Diagnosis of bacterial isolates

After isolating bacteria from wastewater, diagnosed bacteria was grown on nutrient agar for 24 h depending on the characteristics of the colony such as cell shape and biochemical characteristics (gram stain, oxidase, catalysis and dyes), as shown in Table 1.

Table 1 Characteristics of different bacterial isolates; *B. subtilis*, *E. coli* and *P. aeruginosa*.

Bacteria type	Cell shape	Biochemical characteristics			
		Gram stain	Oxidase	Catalysis	Dyes on media
B. subtilis	Rod-shaped	+	_	+	Gray/yellow
E. coli	Rod-shaped	-		+	
P. aeruginosa	Rod-shaped	_	+	+	Yellow/green

3.2. Microbial activity

One—Way ANOVA showed that the antimicrobial activity of both henna leaves and roselle calyxes increased significantly with extract concentrations (Table 2). In this study, the antimicrobial activity was determined by measuring the diameter of inhibition zone of discs soaked in the plant extractions. Plant materials extracted with both water and ethanol showed considerable antimicrobial activity on all of the tested microorganisms especially at 5.0% and 10.0% concentrations (Figure 1). The differences among extractions were not shown at low concentration (2.5%), and were shown clearly at high concentrations (5.0% and 10.0%).

Table 2
Analysis of variance (F-value) of extract concentrations effect on the antimicrobial activity expressed by inhibition zone against different types of bacteria.

Plant materials	B. subtilis	E. coli	P. aeruginosa
Water henna	154.5***	67.4**	75.8**
Water roselle	58.5 ^{**}	34.9**	40.3**
Ethanol henna	79.4**	62.2**	56.8**
Ethanol roselle	63.4**	35.3**	637.0***

^{**} P<0.01, *** P<0.001

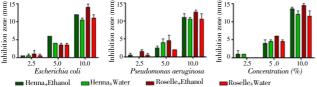


Figure 1. Antibacterial activity of different concentrations of henna and roselle extracts against three different types of bacteria.

Figure 2 showed that roselle extracted by ethanol showed the highest antimicrobial activity against the three types of bacteria, followed with henna extracted by ethanol. In general, ethanolic extracts showed higher activity than those obtained by water extracts, but this different was not significant. *P. aeruginosa* was the most sensitive to plant extracts, followed with *B. subtilis* and *E. coli*, respectively.

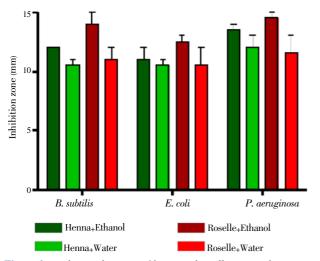


Figure 2. Antibacterial activity of henna and roselle extracts by water and ethanol at concentration of 10% (w/v) against three different types of bacteria; *B. subtilis, E. coli*, and *P. aeruginosa*.

4. Discussion

Medicinal and herbal plants have been used as remedies for human diseases for centuries; this is because they contain components with therapeutic properties in their parts. The antibacterial effect of several plant extracts has been proved previously[30-33]. In this study, henna leaves and roselle calvxes extracted by either water or ethanol increased the inhibition zone on all of the tested bacteria; this result confirmed their antibacterial activity. This is consistent with previous studies which concluded that the extracts of L. alba (henna) and H. sabdariffa (roselle) were shown to have promising antibacterial properties[34,35]. Some studies suggested that henna has a wide spectrum of antimicrobial activity including antibacterial, antiviral, antimycotic and antiparasitic activities. With the ever increasing resistant strains of microorganisms to the already available and synthesized antibiotics, the naturally available henna could be a potential alternative [5,36]. Antimicrobial activity may be due to numerous free hydroxyl ions that have the capability to combine with the carbohydrates and proteins in the bacterial cell wall. They may get attached to enzyme sites rendering them inactive[37]. Also, it is suggested that roselle extracts could be a source of therapeutically useful products[38].

On the other hand, this study showed that ethanolic extract is more efficiency than water extract for both henna and roselle. This result is consistent with[38] who found slight higher inhibition of the roselle ethanol extract against Bacillus subtilis and Staphylococcus aureus than that of water extract. Also, antibacterial activity of henna extracted by alcohol or oil was more effective than that extracted by water[39]. This may be due to the lack of solvent the solvent properties which plays an important role in antibacterial efficacy[40]. roselle extracted by ethanol showed the highest efficiency. These results suggested that roselle had more phyto-chemicals with biological activity than henna, and these phyto-chemicals were more effective when extracted by ethanol rather than water. We recommend studying how to take advantage of these results and its counterparts in the application of wastewater treatment for re-use in the field of agriculture.

Conflict of interest statement

We declare that we have no conflict of interest.

Comments

Background

Wastewater is defined as any water used which may contain pathogens such as bacteria. It is having adverse health implications, natural and ecological affects. So that it is important to treat this before reusing or treatment the diseases which may be caused with the contamination by this water. The researcher observed that several medical plants like roselle and henna that produce secondary metabolites, have biological activities including antimicrobial effects.

Research frontiers

The authors performed a pilot study to investigate the antimicrobial effects of two types of roselle and henna extracts prepared using water and ethanol against both gram positive (*B. subtilis*) and gram negative (*E. coli* and *P. aeruginosa*) bacterial strains isolated from wastewater.

Related reports

The results of the highest antimicrobial activities of roselle and henna ethanolic extracts compared with water extracts are in agreement with (El-Kamali and EL-Karim, 2009; reviewed in Borade, 2011 and Al-Hashimi, 2012). The two extracted used in this study showed variable antibacterial activities it may be due to the differences in the phytochemical constituents extracted

Innovations & breakthroughs

Both of water or ethanol extracts for henna and roselle plants showed high significant antimicrobial effects against some pathogenic bacteria strains.

Applications

It is noteworthy that the wastewater acts as a problem because it contains a lot of pathogens and other components which are hazard for human and the environment. So it is important to treat this water before throw away or reusing in agriculture. This study could be contributed in wastewater treatment, in sterilization and in the therapy of diseases caused by the pathogens under study.

Peer review

The study is good whereas the authors isolated the pathogens from wastewater and they tested the antimicrobial activity of water and ethanol extracts from henna and roselle against some pathogens. The results revealed that ethanolic extract had more antimicrobial activity than water extracts. Moreover, the ethanolic extracted from roselle showed highest antibacterial activity against all tested pathogens than the ethanolic extracted from henna.

References

- Dweck AC. Natural ingredients for colouring and styling. Int J Cosmet Sci 2002; 24: 287–302.
- [2] Rao YM, Shayeda, Sujatha P. Formulation and evaluation of commonly used natural hair colorants. *Nat Prod Rad* 2008; 7: 45– 48.
- [3] Bechtold T. Natural colorants-quinoid, naphthoquinoid and anthraquinoid dyes. In: Bechtold T, Mussak R, editors. *Handbook* of natural colorants. UK: John Wiley & Sons Ltd.; 2009, p. 153–156.
- [4] Mondal MIH, Sayeed MA, Newaz MO, Salam MA. Isolation of a natural dye isolated from the leaves of *Lawsonia inermis* Linn. and its dyeing characteristics onto silk. *Colourage* 2009; 56: 52– 56.
- [5] Rahiman FA, Mahmad N, Taha RM, Elias H, Zaman FH. Antimicrobial properties of *Lawsonia inermis* syn. *Lawsonia alba* in vivo and in vitro. J Food Agric Environ 2013; 11: 502-504.
- [6] Al-Rubiay KK, Jaber NN, Al-Mhaawe BH, Alrubaiy LK. Antimicrobial efficacy of henna extracts. *Oman Med J* 2008; 23(4): 253–256.
- [7] Bairagi GB, Kabra AO, Mandade RJ. Anthelmintic activity of Lawsonia inermis L. leaves in Indian adult earthworm. Int J Res Pharm Biomed Sci 2011; 2: 237–240.
- [8] Ozaslan M, Zumutdal ME, Daglioglu K, Kilic IH, Karagoz ID, Kalender ME, et al. Antitumoral effect of *Lawsonia inermis* in mice with EAC. *Int J Pharmcol* 2009; 5: 263–267.
- [9] Saadabi MAA. Evaluation of *Lawsonia inermis* Linn. (Sudanese henna) leaf extracts as an antimicrobial agent. *Res J Biol Sci* 2007; 2: 419–423.
- [10] Marzin D, Kirkland D. 2-Hydroxy-1,4-naphthoquinone, the natural dye of henna is non-genotoxic in the mouse bone marrow micronucleus test and does not produce oxidative DNA damage in Chinese hamster ovary cells. *Mutat Res* 2004; **560**: 41-47.
- [11] Kirkland D, Marzin D. An assessment of the genotoxicity of 2-hydroxy 1,4-naphthoquinone, the natural dye ingredient of henna. *Mutat Res* 2003; 537: 183-199.
- [12] Kirkland DJ, Henderson L, Marzin D, Muller L, Parry JM, Speit G, et al. Testing strategies in mutagenecity and genetic toxicology: an appraisal of the guidelines of the European Scientific Committee for cosmetics and non-food products for

- the evaluation of hair dyes. Mutat Res 2005; 588: 88-105.
- [13] Raja W, Ovais M, Dubey A. Phytochemical screening and antibacterial activity of *Lawsonia inermis* leaf extract. *Int J Microbiol Res* 2013; 4(1): 33-36.
- [14] Wong PK, Yusof S, Ghazali HM, Man YBC. Physico-chemical characteristics of roselle (*Hibiscus sabdariffa* L.). Nutr Food Sci 2002; 32: 68-73.
- [15] Mertz C, Cheynier V, Gunata Z, Brat P. Analysis of phenolic compounds in two blackberry species (Rubus glaucus and Rubus adenotrichus) by high-performance liquid chromatography with diode array detection and electrospray ion trap mass spectrometry. J Agric Food Chem 2007; 55: 8616-8624.
- [16] Prodanov MP, Dominguez JA, Blazquez I, Salinas MR, Alonso GL. Some aspects of the quantitative/qualitative assessment of commercial anthocyanin-rich extracts. *Food Chem* 2005; 90: 585-596.
- [17] Reyes LF, Cisneros-Zevallos L. Degradation kinetics and colour of anthocyanins in aqueous extracts of purple- and red-flesh potatoes (Solanum tuberosum L.). Food Chem 2007; 100: 885-894.
- [18] Carvajal-Zarrabal O, Hayward-Jones PM, Orta-Flores Z, Nolasco-Hipolito C, Barradas-Dermitz DM, Aguilar-Uscanga MG, et al. Effect of *Hibiscus sabdariffa* L. dried calyx ethanol extract on fat absorption-excretion, and body weight implication in rats. *J Biomed Biotechnol* 2009; 2009: 394592.
- [19] Ross IA. Hibiscus sabdariffa. In: Medicinal plants of the world, vol 1: chemical constituents, traditional and modern medicinal uses. New Jersey: Humana Press; 2003, p. 267–275.
- [20] Wang CJ, Wang JM, Lin WL, Chu CY, Chou FP, Tseng TH. Protective effect of Hibiscus anthocyanins against tert-butyl hydroperoxideinduced hepatic toxicity in rats. Food Chem Toxicol 2000; 38: 411-416.
- [21] Fullerton M, Khatiwada J, Johnson JU, Davis S, Williams LL. Determination of antimicrobial activity of sorrel (*Hibiscus sabdariffa*) on *Escherichia coli* O157:H7 isolated from food, veterinary, and clinical samples. *J Med Food* 2011; 14: 950-956.
- [22] Kamali HHE, Mohammed MF. Antibacterial activity of Hibiscus sabdariffa, Acacia seyal var. seyal and Sphaeranthus suaveolens var. suaveolens against upper respiratory tract pathogens. Sudan J Med Sci 2006; 1: 121–126.
- [23] Morales-Cabrera1 M, Hernández-Morales J, Leyva-Rúelas G, Salinas-Moreno Y, Soto-Rojas L, Castro-Rosas J. Influence of variety and extraction solvent on antibacterial activity of roselle (Hibiscus sabdariffa L.) calyxes. J Med Plants Res 2013; 7(31): 2319-2322.
- [24] Wilson DJ. Insights from genomics into bacterial pathogen populations. *PLoS Pathog* 2012; **8**(9): e1002874.
- [25] Davies J, Davies D. Origins and evolution of antibiotic resistance.

- Microbiol Mol Biol Rev 2010; 74: 417-433.
- [26] Livermore DM. Fourteen years in resistance. Int J Antimicrob Agents 2012; 39: 283–294.
- [27] Schmieder R, Edwards R. Insights into antibiotic resistance through metagenomics approaches. *Future Microbiol* 2012; 7: 73– 89.
- [28] Al-Waili NS, Saloom KY. Effects of topical honey on post-operative wound infections due to gram positive and gram negative bacteria following caesarean sections and hysterectomies. *Eur J Med Res* 1999; **4**(3): 126–130.
- [29] Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing. Tenth informational supplement. Wayne, Pennsylvania: CLSI; 2000.
- [30] Aljamali NM. Study effect of medical plant extracts in comparison with antibiotic against bacteria. J Sci Innov Res 2013; 2: 843–845.
- [31] Habbal O, Hasson SS, El-Hag AH, Al-Mahrooqi Z, Al-Hashmi N, Al-Bimani Z, et al. Antibacterial activity of *Lawsonia inermis* Linn (Henna) against *Pseudomonas aeruginosa*. *Asian Pac J Trop Biomed* 2011; 1: 173–176.
- [32] Masoud EA, Gouda HA. Effect of some natural plant extracts against gram negative bacteria in Njran Area, Saudi Arabia. Egypt Acad J Biolog Sci 2012; 4: 85–92.
- [33] Yaouba A, Tchikoua R, Tatsadjieu NL. Antibacterial effect of plant extracts against some pathogenic bacteria. Int J Nat Prod Res 2012; 1: 83–87.
- [34] Elmanama AA, Alyazji AA, Abu-Gheneima NA. Antibacterial, antifungal and synergistic effect of *Lawsonia inermis*, *Punica granatum* and *Hibiscus sabdariffa*. Ann Alquds Med 2011; 7: 33-41.
- [35] Santhanamari T, Meenakshi PR, Velayutham S. In vitro antibacterial activity of extracts of Lawsonia inermis and Punica grantum against clinically isolated antibiotic resistant Pseudomonas aeruginosa and Staphylococcus aureus. Asian J Pharm Clin Res 2011; 4(1): 62-65.
- [36] Dinesh Babu P, Subhasree RS. Antimicrobial activities of *Lawsonia inermis* a review. *Acad J Plant Sci* 2009; **2**(4): 231–232.
- [37] Harborne JB, Baxter H, editors. Phytochemical dictionary, a handbook of bioactive compounds from plants. London: Tylor and Francis; 1995.
- [38] Jung E, Kim Y, Joo N. Physicochemical properties and antimicrobial activity of roselle (*Hibiscus sabdariffa* L.). J Sci Food Agric 2013; 93: 3769-3776.
- [39] Borade AS, Kale BN, Shete RV. A phytopharmacological review on Lawsonia inermis Linn.). Int J Pharm Life Sci 2011; 2(1): 536– 541.
- [40] Arun P, Purushotham KG, Jayarani JJ, Kumari V. In vitro antibacterial activity and flavonoid contents of Lawsonia inermis (Henna). Int J PharmTech Res 2010; 2(2): 1178-1181.