

Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Disease



journal homepage: www.elsevier.com/locate/apjtd

Original article doi: 10.1016/S2222-1808(15)60983-5

©2016 by the Asian Pacific Journal of Tropical Disease. All rights reserved.

Antibacterial activity of garlic (*Allium sativum*) againts Gram-positive bacteria isolated from tiger shrimp (*Penaeus monodon*)

Supiana Dian Nurtjahyani^{1*}, Fitria Hadra²

¹Department of Education Biology, Faculty of Teaching and Education Science, University of PGRI Ronggolawe, Tuban, East Java, Indonesia

²Department of Biology, Faculty of Mathematics and Natural Science, University of PGRI Ronggolawe, Tuban, East Java, Indonesia

ARTICLE INFO

Article history: Received 5 Nov 2015 Received in revised form 30 Nov, 2nd revised form 11 Dec, 3rd revised form 14 Dec 2015 Accepted 26 Dec 2015 Available online 30 Dec 2015

Keywords: Gram-positive bacteria Garlic Inhibition zone Tiger shrimp

ABSTRACT

Objective: To report the effectiveness of various concentrations of garlic juice on inhibition zone of Gram-positive bacteria isolated from tiger shrimps.

Methods: Four concentrations of garlic juice used in this research were: 0%, 12.5%, 25% and 50%. Two different bacteria were successfully isolated from tiger shrimps categorized as Gram-positive. The bacterial inhibition zone revealed that garlic juice could inhibit the growth of these bacteria in the form of clear zone on media.

Results: The average diameters of inhibition zone from those two bacteria were significantly increased in garlic juice receiving group compared with the control (untreated) group (P < 0.05). For bacteria B, this increase was dose dependent manner.

Conclusions: These findings demonstrated the potential of garlic as a natural alternative to currently used antibacterial agent for Gram-positive infection in the tiger shrimp.

1. Introduction

Tiger Shrimp (*Penaeus monodon*) is one of the fishery and marine products that become the important commodity especially in Southeast Asia, because it has high economic value^[1]. This shrimp has several morphological characteristics such as hard skin and large bluish green motif on the body colour. Besides that, tiger shrimp is usually cultivated in a pond with the average length of 20–25 cm and weight of 140 g. The highest nutrient content in a tiger shrimp is protein^[2].

The nutrient content of a tiger shrimp could be contaminated by several microorganisms which can endanger human health. Several studies had proved that fungi could cause a disease in a shrimp especially the genus of *Fusarium*[3]. This pathogen may lead to high mortalities by disturbing the osmoregulation of shrimp and this is often found in shrimps cultivated in a pond with poor water quality management[4]. Besides that, other microorganisms that also become the major disease problem in shrimps are bacteria frequently causing a great loss in economics of aquaculture products. In the recent study, several researchers observed the major

decline in commercial production of the tiger shrimp due to the disease outbreaks caused by bacteria^[5].

However, this bacteria growth could be inhibited through several ways. Based on its processes, the processes are grouped as physical and chemical processes. Many antibiotics and synthetic chemicals used in shrimp cultivation to prevent the bacterial infection have a residual side effect. The development and discovery of alternative methods to overcome the infection have become the main focus and challenge to keep the sustainability of aquaculture production[3]. Other problem caused by the usage of antibiotics is the growth of antibiotic resistant bacteria[6]. Therefore, the use of natural substances especially from herbs or plants is preferable as an antimicrobial agent to prevent and treat the bacterial infection. This is also recommended by World Health Organization[1].

Garlic is one of the herbs that sparks great interests, since it possesses the antibacterial and antifungal activities. It contains organosulphur compounds (*S*-allylcysteine, *S*-allylcysteinine sulfoxide, *S*-methylcysteine, and *S*-ethylcysteine) and several phenolic compounds^[7,8]. The antibacterial compound can inhibit the bacterial growth and is especially used to cure infection. Garlic can be used as antibacterial agent because it contains allicin. Allicin is the main compound that plays an important role in producing a garlic flavour and one of the active compounds which could eliminate germs by impairing the cell walls and inhibiting the protein synthesis^[9]. Here, we reported the effect of garlic juice on Gram-positive bacteria isolated from tiger shrimp (*Penaeus monodon*).

^{*}Corresponding author: Dr. Supiana Dian Nurtjahyani, Department of Education Biology, Faculty of Teaching and Education Science, University of PGRI Ronggolawe, Tuban, East Java, Indonesia.

Tel: +62 356 322233

Fax: +62 356 331578

E-mail: diantbn@yahoo.co.id

Foundation Project: Supported by University of PGRI Ronggolawe (Grant No. 06/ SP2H/LEMLIT UNIROW/IX/2014).

2. Materials and methods

2.1. Sample preparation

Totally 24 tiger shrimps were used in this research. The tiger shrimps were obtained from tiger shrimp cultivation at Dengok Kandangsemangkon Village, Tuban, East Java. The tiger shrimps obtained from cultivation place were weighed about 25 g and then threshed till smooth and put into 225 mL sterile aquades $(10^{-1} \text{ dilution})$, continued with serial dilution from 10^{-2} to 10^{-6} . In this research, only dilution 10^{-5} and 10^{-6} were used.

2.2. Garlic juice preparation

Garlic juice was made using simple method. Garlic was shredded and squeezed to obtain the liquid content. Then it was filtered using filter paper to clean the garlic debris from the garlic juice. Several concentrations were made by adding sterile aquades for dilution. Three concentrations of garlic juice used here were 12.5%, 25%, and 50%.

2.3. Bacteria culture

Bacteria were grown using nutrient agar (Sigma) media. About 1 mL sample with dilution 10^{-5} and 10^{-6} were taken and put into Petri dishes, followed by pouring the media into the Petri dishes and shaken for a while to make it homogenous. Bacteria were grown for 24 h. Bacterial colonies observed were purified and continued with Gram staining. Those bacteria were used for bacterial inhibition zone test.

2.4. Bacterial inhibition zone test

One dose of bacteria pure culture was put into reaction tube containing 10 mL nutrient agar liquid. Solution was poured into sterile Petri dish and homogenized. After agar became solid, Petri dish was divided into four areas for four different concentrations (0%, 12.5%, 25% and 50%) of garlic juice treatment. Culture was incubated for 24 h and observed for the appearance of clear zone and the diameter of the clear zone was measured.

2.5. Statistical analysis

Data analysis and processing were done using SPSS for windows. ANOVA test was performed to test the data significance.

3. Results

Two bacteria randomly taken were used in this research. Complete analysed was not conducted to determine the bacteria species. Only morphology analysis and Gram staining were conducted to observe the differences between those two bacteria. First bacteria (bacteria A) had morphological characteristics such as red colour, round shape, flat edge, and convex elevation. Second bacteria (bacteria B) had morphological characteristics that were similar to those of the bacteria A; the only difference was the colour, in which bacteria B had white milk colour. After Gram staining analysis, both bacteria had blue colour that made them classified as Gram-positive bacteria (Figure 1).

Figure 2 summarizes the diameter of clear zone bacteria culture. For bacteria A, the diameter of clear zone was significantly greater in all doses of garlic juice group compared with the untreated control group (P < 0.05). There was no significant difference between the effects of these two higher doses. For bacteria B, the diameter of clear zone was significantly greater in all doses of garlic juice group compared with the untreated control group (P < 0.05).



Figure 1. Observation of bacterial growth inhibition analysis. The appearance of clear zone on culture of bacteria A (A) and bacteria (B). The area seems increasing due to the increased garlic juice concentration. 1: 0% Garlic juice concentration; 2: 12.5% Garlic juice concentration; 3: 25% Garlic juice concentration; 4: 50% Garlic juice concentration.



Figure 2. Measurement results of clear zone average diameter. Clear zones were quantified by measuring the diameter whether in culture of bacteria A (A) or bacteria B (B). The average diameter was also increased in line with the increased concentration. P1: Treatment with 0% garlic juice concentration; P2: Treatment with 12.5% garlic juice concentration; P3: Treatment with 25% garlic juice concentration; P4: Treatment with 50% garlic juice concentration. Significant different (P < 0.05).

4. Discussion

In the recent years there have been significant increasing number of researches that focus on the knowledge of the disease infection in the shrimps. However, fishery products are still considered as a commodity with high risk of pathogen content, natural toxins, and many other potential contaminants^[10,11]. Although some researches are focusing on the infection caused by Gram-negative bacteria especially from genus *Vibrio*^[12], the role of the Gram-positive bacteria on disease infection also could not be ruled out. Generally, the diseases caused by this pathogen are through their invasion and growth within tissues rather than toxin production and dissemination^[10,11].

In our study, we have isolated two Gram-positive bacteria from tiger shrimp. Previous studies found that several different bacteria strains are isolated from tiger shrimp muscles such as *Aureobacterium faciens*, *Aeromicrobium erythreum*, *Bacillus subtilis*, *Escherichia* *coli, Vibrio cholerae, Enterobacter aerogenes, Micrococcus luteus, Pseudomonas putida, Pseudomonas aeruginosa* and *Enterococcus pseudoavium*^[13]. Although, we did not further investigate the bacteria species, according to their the morphologies, we characterize the two Gram-positive bacteria from tiger shrimps which were closely related to some of the above bacteria.

We used garlic in our study, since its nutritional effect has been extensively investigated as antibacterial agent against a variety of Gram-negative and Gram-positive bacteria. Treatment was conducted using garlic juice or the water extract of juice. Garlic is mainly composed of water. The bacterial growth inhibition using garlic juice generated the positive effect. This result was consistent with several previous studies that showed the inhibitory effect of garlic aqueous extract on numerous bacterial species[14-16].

The inhibition zone analysis was marked by clear zone and increasing average diameter due to the increasing concentrations of garlic juice. We hypothesized that the antibacterial activity of garlic juice on Gram-positive bacteria involves several mechanisms. Anion compounds including nitrates, chlorides, sulfides and organosulphur compounds can be easily resolved in water and are responsible for antibacterial properties[17]. Allicin in garlic is the most important substance that produces antibacterial properties and restricts the speed of RNA synthesis. Lipid is a part of Gram-positive membrane, which helps the easy penetration of allicin into the membrane and consequently influences the RNA[18]. Allicin has been reported to kill pathogens through partial inhibition of DNA and protein synthesis, alteration of the electrochemical ability, induce apoptosis in cells, affect microbial lipid biosynthesis, signal transduction, as well as react with thiol-containing proteins[19-21]. Our finding is consistent with previous studies stating that garlic extract showed a clear zone in the culture of Gram-positive bacteria[22,23].

In conclusion, these findings demonstrate the potential of garlic as a natural alternative to currently used antibacterial agent for Grampositive infection in the tiger shrimp. Further researches related to the species identification of those two bacteria and increasing number of Gram-positive bacteria need to be carried out.

Conflict of interest statement

We declare that we have no conflict of interest.

Acknowledgments

We thank laboratory staff of Fisheries and Marine Faculty, University of PGRI Ronggolawe, Tuban, East Java, Indonesia for technical supports. This work was supported by University of PGRI Ronggolawe (Grant No. 06/SP2H/LEMLIT UNIROW/IX/2014).

References

- Flegel TW. Historic emergence, impact and current status of shrimp pathogens in Asia. J Invertebr Pathol 2012; 110: 166-73.
- [2] Balia RRL, Abun A, Aisjah TT, Darana SS. Bioprocessed of tiger prawn (*Penaeus monodon*) waste product by deproteination and demineralization on nutrient product. *Lucrări Științifice - Universitatea de Științe Agricole şi Medicină Veterinară, Seria Zootehnie* 2010; **54**(15): 60-2.
- [3] Citarasu T. Herbal biomedicines: a new opportunity for aquaculture Industry. *Aquacult Int* 2010; 18: 403-14.
- [4] Ramaiah N. A review on fungal disease of algae, marine fish, shrimp and corals. *Indian J Mar Sci* 2006; 35(4): 380-7.
- [5] Tanticharoen M, Flegel TW, Meerod W, Grudloyma U, Pisamai N. Aquacultural biotechnology in Thailand: the case of the shrimp industry. *Int J Biotechnol* 2009; 10: 588-603.
- [6] Nimrat S, Vuthiphandchai V. In vitro evaluation of commercial probiotic products used for marine shrimp cultivation in Thailand. Afr J Biotechnol

2011; 10(22): 4643-50.

- [7] Tsai DC, Liu MC, Lin YR, Huang MF, Liang SS. A novel reductive amination method with isotopic formaldehydes for the preparation of internal standard and standards for determining organosulfur compounds in garlic. *Food Chem* 2016; **197**: 692-8.
- [8] Raghavendra CK, Srinivasan K. Potentiation of anti-cholelithogenic influence of dietary tender cluster beans (*Cyamopsis tetragonoloba*) by garlic (*Allium sativum*) in experimental mice. *Indian J Med Res* 2015; 142(4): 462-70.
- [9] Li G, Ma X, Deng L, Zhao X, Wei Y, Gao Z, et al. Fresh garlic extract enhances the antimicrobial activities of antibiotics on resistant strains *in vitro. Jundishapur J Microbiol* 2015; 8(5): e14814.
- [10] Rungrassamee W, Klanchui A, Maibunkaew S, Karoonuthaisiri N. Bacterial dynamics in intestines of the black tiger shrimp and the Pacific white shrimp during *Vibrio harveyi* exposure. *J Invertebr Pathol* 2015; 133: 12-9.
- [11] Rungrassamee W, Klanchui A, Chaiyapechara S, Maibunkaew S, Tangphatsornruang S, Jiravanichpaisal P, et al. Bacterial population in intestines of the black tiger shrimp (*Penaeus monodon*) under different growth stages. *PLoS One* 2013; 8(4): e60802.
- [12] Chaiyapechara S, Rungrassamee W, Suriyachay I, Kuncharin Y, Klanchui A, Karoonuthaisiri N, et al. Bacterial community associated with the intestinal tract of *P. monodon* in commercial farms. *Microbiol Ecol* 2012; 63(4): 938-53.
- [13] Narasimhan N, Ravimanickam T, Sukumaran M, Ravichelvan R, Ravichandran R, Madhavan D. Pathogenic bacteria isolated from tiger prawn *Penaeus monodon* in shrimp culture ponds at east coast of Thanjavur District Tamil Nadu, India. *Int J Res Biol Sci* 2013; 3(2): 98-101.
- [14] Mnayer D, Fabiano-Tixier AS, Petitcolas E, Hamieh T, Nehme N, Ferrant C, et al. Chemical composition, antibacterial and antioxidant activities of six essentials oils from the Alliaceae family. *Molecules* 2014; 19(12): 20034-53.
- [15] Lin L, Wang J, Yu J, Li Y, Liu G. Effects of allicin on the formation of *Pseudomonas aeruginosa* biofinm and the production of quorum-sensing controlled virulence factors. *Pol J Microbiol* 2013; **62**(3): 243-51.
- [16] Andualem B. Combined antibacterial activity of stingless bee (*Apis mellipodae*) honey and garlic (*Allium sativum*) extracts against standard and clinical pathogenic bacteria. *Asian Pac J Trop Biomed* 2013; 3(9): 725-31.
- [17] Shobana S, Vidhya VG, Ramya M. Antibacterial activity of garlic varieties (ophioscordon and sativum) on enteric pathogens. *Curr Res J Biol Sci* 2009; 1(3): 123-6.
- [18] Deresse D. Antibacterial effect of garlic (Allium sativum) on Staphylococcus aureus: An in vitro study. Asian J Med Sci 2010; 2(2): 62-5.
- [19] Gruhlke MCH, Portz D, Stitz M, Anwar A, Schneider T, Jacob C, et al. Allicin disrupts the cell's electrochemical potential and induces apoptosis in yeast. *Free Radic Biol Med* 2010; **49**: 1916-24.
- [20] Lu X, Liu Q, Wu D, Al-Qadiri HM, Al-Alami NI, Kang DH, et al. Using of infrared spectroscopy to study the survival and injury of *Escherichia coli* O157:H7, *Campylobacter jejuni* and *Pseudomonas aeruginosa* under cold stress in low nutrient media. *Food Microbiol* 2011; 28: 537-46.
- [21] Lu X, Rasco BA, Jabal JMF, Aston DE, Lin M, Konkel ME. Investigating antibacterial effects of garlic (*Allium sativum*) concentrate and garlicderived organosulfur compounds on *Campylobacter jejuni* by using Fourier transform infrared spectroscopy, Raman spectroscopy, and electron microscopy. *Appl Environ Microbiol* 2011; **77**: 5257-69.
- [22] Viswanathan V, Phadatare AG, Mukne A. Antimycobacterial and antibacterial activity of *Allium sativum* Bulbs. *Indian J Pharm Sci* 2014; 76(3): 256-61.
- [23] Abubakar EM. Efficacy of crude extracts of garlic (Allium sativum Linn.) against nosocomial Escherichia coli, Staphylococcus aureus, Streptococcus pneumoniea and Pseudomonas aeruginosa. J Med Plants Res 2009; 3(4): 179-85.