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## Anxiolytic potential of ursolic acid derivative—a stearyl glucoside isolated from *Lantana camara* L. (verbanaceae)

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### ABSTRACT

**Objective:** To investigate the anxiolytic activity of newly isolated compound by our lab called ursolic acid stearyl glucoside (UASG) from the leaves of *Lantana camara* (*L. camara*). **Methods:** Column chromatography was used to isolate UASG. Anxiolytic potential was experimentally proved and demonstrated through Elevated plus-maze, Open field and light and dark test. **Results:** The UASG showed marked increased in time spent (%) and number of frequent movements made by animals in open arm of elevated plus-maze apparatus. In light and dark model, UASG produced marked increase in time spent by animal, number of crossing and reduced duration of immobility in light box. **Conclusions:** UASG showed significant increase in number of rearing, assisted rearing and number of square crossed in open field established test model. UASG showed its anxiolytic effect in dose dependent manner.

## 1. Introduction

Anxiety disorders are the most common psychiatric disorders[1]. They accounts for heavy burden on public health. Present epidemiological data gives a different glimpse of this disorder, as traditionally thought, almost involving half of the population[2]. Every eighth human of total population is linked with anxiety, making this field an important area of research, for psychopharmacologist[3], hence it became need of an hour to develop new anxiolytics. Many secondary plant metabolites are reported in the treatment of psychological disorders. The anti anxiety activities plant metabolites are used in traditional medicine practice, either directly or indirectly

affecting the central nervous system there by altering gamma-amino butyric acid, serotonin noradrenalin, benzodiazepine neurotransmitters activities. It is already proved that ursolic acid and its derivative is used in the treatment of central nerve system (CNS) disorders[4]. Ursolic acid obtained from *Nepeta sibthorpii* reported for CNS depressing, anticonvulsant and analgesic activity[5]. Awad *et al* in 2009 also reported *in-vitro* assays of ursolic acid inhibited GABA-T by 20% at a dose of 100  $\mu$ g/ml[6].

*Lantana camara* (*L. camara*) L. is well known traditional and tropical folk medicine[7–11]. Ursolic acid a stearyl glucoside (UASG), pentacyclic triterpenoid was isolated from *L. camara* L. (family: verbanaceae)[12]. The traditional uses of *L. camara* L. mainly refer for the treatment of asthma, ulcers, measles, chickenpox, eczema, tumors, cancers, high blood pressure, bilious fevers, catarrhal infections, tetanus, rheumatism, malaria, ataxy of abdominal viscera[13,14], epilepsy[15], memory weakness, enhance intellect and cognition[16]. The plant

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is reported to possess anticonvulsant<sup>[17]</sup>, anticancer<sup>[18,19]</sup>, antiulcer<sup>[20]</sup>, antioxidant<sup>[19]</sup>, anti-diabetic<sup>[21,22]</sup>, antifungal, antibacterial<sup>[23–25]</sup>, anti-feedant, larval mortality/repellency<sup>[2]</sup>, antimotility<sup>[29]</sup>, analgesic and anti-inflammatory activities<sup>[30]</sup>.

In Indian traditional herbal medicine, it is reported for its use in treatment of mental disorders and found to be effective as a brain tonic. Hence, in this work we used UASG isolated from *L. camara* L. for its anxiolytic effect against elevated plus-maze test (EPM), open field test and light and dark test induced anxiety in experimental animals.

## 2. Materials and methods

### 2.1. Animals

The experimental mice were obtained from the animal house of Siddhartha Institute of Pharmacy, Dehradun, India, (1435/PO/a/11/CPCSEA). The animals were housed as per the guidelines of CPCSEA. All the animals were provided with standard pellet diet (Lipton rat feed, Ltd., Pune) and water ad libitum throughout the experimental protocol. All the animal studies were approved by the Institutional Animal Ethical Committee of Siddhartha Institute of Pharmacy, Dehradun, India. The animals were randomly segregated into four groups, having six mice in each group. Group I, solvent control (received 0.9% (w/v) of saline); Group II, a positive control (diazepam 1 mg/kg); Group III and IV were received UASG 25 and 50 mg/kg suspended in 1% Tween 80 (v/v). All the animals were administered with above drugs/solvent etc intraperitoneally, 30 min prior to start the experiment.

### 2.2. Plant material

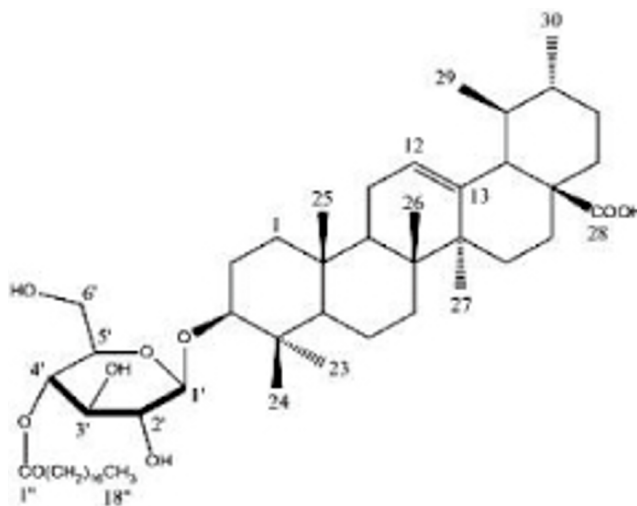
*L. camara* L. leaves were collected from Dehradun and identified by Dr. S. B. Singh, Scientist, NISCAIR, New Delhi. A voucher specimen (NISCAIR/RHMD/consult/-20-09-10/1322/125) was deposited in the herbarium of NISCAIR, India.

### 2.3. Extraction and isolation of UASG

Dried powder of *Lantana camara* leaves (4 kg) was extracted with methanol (12 L) at 50 °C for 1 d. Extract was concentrated to dryness under reduced pressure to obtain slurry (605 g).

The slurry was dissolved in minimum amount of methanol and was adsorbed on silica gel (60–120 mesh). The slurry was subjected to a silica gel column using CHCl<sub>3</sub>/MeOH gradient

system (49:1; 2.0 L for gradient system). It leads to elution of colorless crystals of USAG (yield 11.2 g, 0.28%). It was found to be 100% pure by HPTLC by using solvent system CHCl<sub>3</sub>/MeOH (99:1). Structure of compound was identified by comparison of their spectroscopic data from the reported literature<sup>[12]</sup>. The structure of USAG is depicted in Figure 1.



**Figure 1.** Structure of ursolic acid stearyl glucoside.

### 2.4. Chemicals

Diazepam (Ranbaxy, New Delhi, India) was purchased through institution. All the chemicals were of analytical grade. Tween 80 (1% v/v) in saline was used to suspend UASG and was kept for its use.

### 2.5. Behavioral parameters used to test anxiolytic activity

#### 2.5.1. Elevated plus-maze test (EPM)

The EPM apparatus designed by institute with following dimensions four arms elevated 25 cm from the floor. Each arm is at 90° relative to the adjacent arms. Two arms enclosed with high walls (35 cm×5 cm×20 cm), and the other two linked via a central area (5 cm×5 cm) giving an appearance of plus sign. The maze floor and the walls of enclosed arms were black. An illumination with a 40 W lamp was made in the room. The animals were administered with vehicle, diazepam (1 mg/kg)<sup>[31]</sup> and USAG (25 and 50 mg/kg) intraperitoneally, 30 min prior to the test. Initially the mice were accustomed to lab condition half an hour prior to behavioral testing.

Each animal was individually placed on the central platform. The movement and duration of entries into the closed and open arms were observed for 5 min. Complete entry was considered and counted only when all four paws of the mouse entered an open or closed arm. The time spent (duration) in percentage for the open arms [100 × open/open

+ enclosed)] and percentage of the number of open arm entries (frequency,  $100 \times \text{open}/\text{total entries}$ ) were calculated for each mouse[32].

### 2.5.2. Open field test

The apparatus designed with modification and dimensions of a wooden box (60 cm×60 cm×60 cm), was used. The area of open field was divided into 16 equal squares (15 cm×15 cm). The 12 squares were at the periphery and 4 inner squares in the center. The experimental was performed in, dark room with sound proof facility. The open field area was illuminated with a 40 W lamp, focusing from the height of 75–100 cm. Treatment was started with vehicle, diazepam (1 mg/kg) and USAG (25 and 50 mg/kg), by placing individual animal in one of the corner squares and number of rearing, assisted rearing (forepaws touching the walls of the apparatus) and number of squares crossed were noticed for 5 min[33].

### 2.5.3. Light and dark test

The Light and dark test apparatus is top open wooden box, with two distinct chambers, a black (25 cm long ×35 cm wide ×35 cm deep), and white which is illuminated with 40 W light source as white, was placed approx. 25 cm above the open box. The two chambers were linked with a small opening, (7.5 cm long ×5.0 cm wide) located on the floor level at the center. The individual animal was placed in center of the light box and was observed for 5 min[34].

## 3. Result

As per our results animal treated with vehicle spent (53.29±

1.52) s in open arm and it's time for closed arm was recorded as (135.40±1.62) s with demonstrating the number of (4.86±0.28) entries in open arm and (33.85±0.74) in closed arm. The animal showed the increased stability in the open arm when treated with Diazepam (1 mg/kg) and UASG (25 and 50 mg/kg), significantly ( $P<0.05$  and  $P<0.001$ , respectively), whereas significant ( $P<0.01$  and  $P<0.001$ , respectively) reduction was observed in number of entries and time spent in closed arm. UASG (25 and 50 mg/kg) and diazepam significantly ( $P<0.001$ ) increased the percentage of time spent and entries in open arm (Table 1).

84.85±1.58 squares were crossed by vehicle-treated animal and demonstrated (17.35±0.44) self rearing and 18.90±0.88 assisted rearing during the interval of 5 min. when the animals were treated with Diazepam and UASG (25 and 50 mg/kg), significant ( $P<0.05$  and  $P<0.001$ , respectively) increase was noticed in the number of squares crossed by the animals. The self rearing and assisted rearing was altered significantly ( $P<0.05$  and  $P<0.001$ , respectively) i.e. increased by UASG (25 and 50 mg/kg) and diazepam (Table 2). The animals treated with diazepam (1 mg/kg) and UASG (25 and 50 mg/kg) showed significant change ( $P<0.05$ ,  $P<0.001$  and  $P<0.001$ , respectively) i.e their time spent in the lighted box was increased and decrease the dark box. UASG (25 mg/kg) failed to bring any significant change in the number of crossing and duration of immobility, where as diazepam (1 mg/kg) and UASG (50 mg/kg) really showed significant ( $P<0.05$  and  $P<0.01$ , respectively) increase in the number of crossing and decrease in the duration of immobility (Table 3).

**Table 1**

Effect of UASG on behavior of mice in elevated plus maze test.

Treatment (mg/kg, i.p.)	No. of entries		Time spent (s)		%OAE	TSOA (s)
	Open arm	Closed arm	Open arm	Closed arm		
Vehicle	4.86±0.28	33.85±0.74	53.29±1.52	135.40±1.62	23.74±0.74	35.90±0.78
Diazepam (1)	9.88±0.63 <sup>***</sup>	23.89±0.47 <sup>***</sup>	74.55±1.41 <sup>***</sup>	85.50±1.73 <sup>***</sup>	49.58±1.79 <sup>***</sup>	56.20±1.00 <sup>***</sup>
UASG (25)	6.10±0.45 <sup>*</sup>	17.62±0.24 <sup>**</sup>	57.88±1.20	110.60±1.93 <sup>**</sup>	52.36±2.05 <sup>***</sup>	42.32±0.58 <sup>**</sup>
UASG (50)	8.36±0.52 <sup>***</sup>	20.01±0.38 <sup>***</sup>	74.47±1.19 <sup>***</sup>	86.85±1.71 <sup>***</sup>	52.15±2.20 <sup>***</sup>	54.87±1.16 <sup>***</sup>

Values are the mean ± SEM of 6 mice / treatment, Significant <sup>\*</sup> $P<0.05$ , <sup>\*\*</sup> $P<0.01$  and <sup>\*\*\*</sup> $P<0.001$  compared with control.

**Table 2**

Effect of UASG on behavior of mice in open field test.

Treatment (mg/kg, i.p.)	No. of rearing	No. of assisted rearing	No. of squares crossed
Vehicle	17.35±0.44	18.90±0.88	84.85±1.58
Diazepam (1)	31.67±1.46 <sup>***</sup>	33.34±0.95 <sup>***</sup>	172.80±1.36 <sup>***</sup>
UASG (25)	27.99±0.40 <sup>***</sup>	26.04±0.97 <sup>**</sup>	105.40±1.10 <sup>*</sup>
UASG (50)	28.92±1.64 <sup>***</sup>	29.81±0.74 <sup>***</sup>	147.90±1.34 <sup>***</sup>

Values are the mean ± SEM of 6 mice / treatment, Significant <sup>\*</sup> $P<0.05$ , <sup>\*\*</sup> $P<0.01$  and <sup>\*\*\*</sup> $P<0.001$  compared with control.

**Table 3**

Effect of UASG on behavior of mice in light and dark model.

Treatment (mg/kg, i.p.)	Time spent in lighted box (s)	Time spent in dark box (s)	No. of crossing	Duration of immobility (s)
Vehicle	86.35±1.51	201.10±2.01	24.13±1.98	37.85±2.92
Diazepam (1)	175.00±1.82***	121.40±2.62***	32.53±1.63*	28.17±2.31*
UASG (25)	106.20±1.50*	177.20±2.56**	26.04±1.35	34.16±2.23
UASG (50)	156.60±1.69***	125.30±2.39***	31.94±1.93*	25.63±1.65***

Values are the mean ± SEM of 6 mice/treatment, Significant \* $P<0.05$ , \*\* $P<0.01$  and \*\*\* $P<0.001$  compared with control.

#### 4. Discussion

Elevated plus maze, works on the fact that native mice has tendency to spend more time in closed arms as compared to open one. This may be due to fear generated in the open space. Present study on UASG, is a dose dependent, which induces significant increases both in the number of entries and time spent in the open arms, where as decreases the number of entries and time spent in the closed arm, proving its anxiolytic nature<sup>[35]</sup>.

The open field model evaluates anxiety related activities characterized by the normal aversion of the animal to an open and bright area. Animals when subjected to change in their acclimatized location, expresses anxiety and fear<sup>[36]</sup>. UASG, in a dose dependent manner, significantly changes anxiety parameter that is the increase in the number of self rearing, number of assisted rearing and number of squares crossed, proving its anxiolytic effect by reducing such fearful behavior of animals in open field. The light and dark box method is a natural test for rodents which shows that they avoid bright places<sup>[37]</sup>. UASG may reduce the fear of animal which allows the animal to increase more the time in bright and open space. This effect may be due to the agonistic effect on GABA/benzodiazepine receptor complex. Ursolic acid and its derivatives crosses the blood brain barrier<sup>[38,39]</sup>. UASG is a derivative of ursolic acid. It is known to possess anxiolytic activity<sup>[6]</sup>, and all the activities may be due to ursolic acid only. In conclusion, UASG exhibited significant activity in elevated plus maze, light and dark model and open field test induced anxiety models, proved its anxiolytic action. Exact mechanism of action needs a further detailed study.

#### Conflict of interest statement

We declare that we have no conflict of interest.

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#### References

- [1] Kessler RC, McGonagle KA, Zhao S, Nelson CB, Hughes M, Eshleman S, et al. Lifetime and 12-month prevalence of DSM-III-R psychiatric disorders in the United States: results from the National Comorbidity Survey. *Arch Gen Psychiatry* 1994; **51**: 8–19.
- [2] Moffitt TE, Caspi A, Taylor A, Kokaua J, Milne BJ, Polanczyk G, et al. How common are common mental disorders? Evidence that lifetime rates are doubled by prospective versus retrospective ascertainment. *Psychol Med* 2010; **39**: 899–909.
- [3] Eisenberg RB, Davis SL, Etner S, Appel S, Wilkey M, Rompay V, et al. Trends in alternative medicine used in United States: Results of a follow up national survey. *J Am Med Assoc* 1998; **280**: 1569–1575.
- [4] Martin R, Carvalho-Tavares J, Hernandez M, Arnees M, Ruiz-Gutierrez V, Niet ML. Beneficial actions of oleanolic acid in an experimental model of multiple sclerosis: A potential therapeutic role. *Biochem Pharmacol* 2010; **79**: 198–210.
- [5] Taviano MF, Miceli N, Monforte MT, Tzakou O, Galati EM. Ursolic acid plays a role in *Nepeta sibthorpii* bentham CNS depressing effects. *Phytother Res* 2007; **21**: 382–385.
- [6] Awad R, Muhammad A, Durst T, Trudeau VL. Bioassay guided fractionation of lemon balm (*Melissa officinalis* L.) using an *in vitro* measure of GABA transaminase activity. *Phytother Res* 2009; **23**: 1075–1081.
- [7] Pour BM, Sasidhara S. *In vivo* toxicity study of *Lantana camara*. *Asian Pac J Trop Biomed* 2011; 230–223.
- [8] Kalita S, Kumar G, Karthik L, Rao KV. *In vitro* antioxidant and DNA damage inhibition activity of aqueous extract of *Lantana camara* L. (Verbenaceae) leaves. *Asian Pac J Trop Biomed* 2012; **2**(Suppl 3): S1675–S1679.
- [9] Seth R, Mohan M, Singh P, Haider SZ, Gupta S, Bajpai I, et al. Chemical composition and antibacterial properties of the essential oil and extracts of *Lantana camara* Linn. from Uttarakhand (India). *Asian Pac J Trop Biomed* 2012; **2**(Suppl 3): S1407–S1411.
- [10] Kumar D, Kumar A, Prakash O. Pharmacognostic study of *Lantana camara* Linn. root. *Asian Pac J Trop Dis* 2012; **2**(Suppl 1): S42–S45.
- [11] Kazmi I, Gupta G, Afzal M, Anwar F. Anticonvulsant and depressant-like activity of ursolic acid stearoyl glucoside isolated from *Lantana camara* L. (verbanaceae). *Asian Pac J Trop Dis*

- 2012; **2**(Suppl 1): S453–S456.
- [12]Kazmi I, Rahman M, Afzal M, Gupta G, Saleem S, Afzal O, et al. Anti-diabetic potential of ursolic acid stearyl glucoside: A new triterpenic glycosidic ester from *Lantana camara*. *Fitoterapia* 2012; **83**: 142–146.
- [13]Ghisalberti EL. *Lantana camara* L. (Verbenaceae). *Fitoterapia* 2000; **71**: 467–486.
- [14]Day MD, Wiley CJ, Playford J, Zalucki MP. *Lantana: current management, status and future prospects*. Australian Centre for International Agricultural Research: Canberra; 2003.
- [15]Ganesh T, Sen T, Thilagam E, Thamocharan G, Loganathan T, Chakraborty R. Pharmacognostic and anti-hyperglycemic evaluation of *Lantana camara* (L.) var. aculeate leaves in alloxan-induced hyperglycemic rats. *Int J Res Pharm Sci* 2010; **1**(3): 247–252.
- [16]Adams M, Gmunder F, Hamburger M. Plants traditionally used in age related brain disorders: a survey of ethnobotanical literature. *J Ethnopharmacol* 2007; **113**: 363–381.
- [17]Bisi-Johnson MA, Obi CL, Hattori T, Oshima Y, Li S, Kambizi L, et al. Evaluation of the antibacterial and anticancer activities of some South African medicinal plants. *BMC Complement Altern Med* 2011; **11**: 14–18.
- [18]Gomes de Melo J, de Sousa Araújo TA, Thijian Nobre de Almeida e Castro V, Lyra de Vasconcelos Cabral D, do Desterro Rodrigues M, Carneiro do Nascimento S, et al. Antiproliferative activity, antioxidant capacity and tannin content in plants of semi-arid northeastern Brazil. *Molecules* 2010; **15**: 8534–8542.
- [19]Sathish R, Vyawahare B, Natarajan K. Antilucerogenic activity of *Lantana camara* leaves on gastric and duodenal ulcers in experimental rats. *J Ethnopharmacol* 2011; **134**: 195–197.
- [20]Venkatachalam T, Kumar VK, Selvi PK, Maske AO, Anbarasan V, Kumar PS. Anti-diabetic activity of *Lantana camara* Linn fruits in normal and streptozotocin-induced diabetic rats. *J Pharm Res* 2011; **4**: 1550–1552.
- [21]Garg SK, Shah MA, Garg KM, Farooqui MM, Sabir M. Anti-lymphocytic immunosuppressive effects of *Lantana camara* leave in rats. *Indian J Exp Biol* 1997; **35**: 1315–1318.
- [22]Sinha P, Saxena SK. Effect of treating tomatoes with leaf extract of *Lantana camara* on development of fruit rot caused by *Aspergillus niger* in presence of *Drosophila busckii*. *Indian J Exp Biol* 1987; **25**: 143–144.
- [23]Rwangabo PC, Claeys M, Pieters L, Corthout J, Vanden Berghe DA, Vlietinck AK. Umuhengerin, a new antimicrobially active flavonoid from *Lantana trifolia*. *J Nat Prod* 1988; **51**: 966–968.
- [24]Barreto F, Sousa E, Campos A, Costa J, Rodrigues F. Antibacterial activity of *Lantana camara* Linn and *Lantana montevidensis* Brig extracts from Cariri-Ceará, Brazil. *J Young Pharm* 2010; **2**: 42–44.
- [25]Pandey ND, Singh M, Tewari GC. Antifeeding repellent and insecticidal properties of some indigenous plant material against mustard saw Ify, *Athalia pronuima* klug. *Indian J Ent* 1977; **39**: 60–63.
- [26]Chavan SR, Nikam ST. Investigation of *Lantana camara* Linn (Verbenaceae) leaves for larvicidal activity. *Bull Haffkin Inst* 1982; **10**: 21–22.
- [27]Pandey UK, Srivastava A, Lekha C, Singh A. Efficacy of certain plant extracts against brinjal aphid *Aphis gossypii* Glover. *Indian J Ent* 1983; **45**: 313–314.
- [28]Sagar L, Sehgal R, Ojha S. Evaluation of antimotility effect of *Lantana camara* L. var. aculeata constituents on neostigmine induced gastrointestinal transit in mice. *BMC Complement Altern Med* 2005; **5**: 18–23.
- [29]Ghosh S, Das Sarma M, Patra A, Hazra B. Anti-inflammatory and anticancer compounds isolated from *Ventilago madraspatana* Gaertn., *Rubia cordifolia* Linn. and *Lantana camara* Linn. *J Pharm Pharmacol* 2010; **62**: 1158–1166.
- [30]Commission on Epidemiology and Prognosis, International League Against Epilepsy, 1993. Guidelines for epidemiologic studies on epilepsy. *Epilepsia* 1993; **34**: 592–596.
- [31]Kuribara H, Stavinoha WB, Maruyama Y. Honokiol, a putative anxiolytic agent extracted from magnolia bark, has no diazepam-like side-effects in mice. *J Pharm Pharmacol* 1999; **51**(1): 97–103.
- [32]Adeyemi OO, Yetmitan OK, Taiwo AE. Neurosedative and muscle relaxant activities of ethyl acetate extract of *Baphia nitida* AFZEL. *J Ethnopharmacol* 2006; **106**: 312–316.
- [33]Yadav AV, Kawale LA, Nade VS. Effect of *Morus alba* L. (mulberry) leaves on anxiety in mice. *Ind J Pharmacol* 2008; **40**: 32–36.
- [34]Ambavade SD, Mhetre NA, Tate VD, Bodhankar SL. Pharmacological evaluation of the extracts of *Sphaeranthus indicus* flowers on anxiolytic activity in mice. *Ind J Pharmacol* 2006; **38**: 254–259.
- [35]Hellion-Ibarrola MC, Ibarrola DA, Montalbetti Y, Kennedy ML, Heinichen O, Campuzano M, et al. The anxiolytic-like effects of *Aloysia polystachya* (Griseb) Moldenke (Verbenaceae) in mice. *J Ethnopharmacol* 2006; **105**: 400–408.
- [36]Mechan AO, Moran PM, Elliott M, Young AJ, Joseph MH, Green R. A comparison between dark agouti and Sprague-Dawley rats in their behavior on the elevated plus-maze, open field apparatus and activity meters and their response to diazepam. *Psychopharmacology* 1995; **121**: 38–56.
- [37]Bourin M, Hascoet M. The mouse light/dark box test. *Eur J Pharmacol* 2003; **463**: 55–65.
- [38]Tsai S, Yin MC. Antioxidative and anti-inflammatory protection of oleanolic acid and ursolic acid in PC12 cells. *J Food Sci* 2008; **73**(7): H174–H178.
- [39]Martin R, Carvalho-Tavares J, Hernández M, Arnes M, Ruiz-Gutiérrez V, Nieto ML. Beneficial actions of oleanolic acid in an experimental model of multiple sclerosis: A potential therapeutic role. *Biochem Pharmacol* 2010; **79**(2): 198–208.