

Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Medicine

journal homepage:www.elsevier.com/locate/apjtm

Document heading doi: 10.1016/S1995-7645(14)60016-X

# Clinical implications and treatment of dengue

# Chawla Pooja<sup>\*</sup>, Yadav Amrita, Chawla Viney

Faculty of Pharmacy–Pharmaceutical Chemistry, BBDNITM, Sector–1, Dr. Akhilesh Das Nagar, Faizabad Road, Chinhut Lucknow Lucknow Uttar Pradesh 226028, India

### ARTICLE INFO

ABSTRACT

Article history: Received 10 September 2013 Received in revised form 15 November 2013 Accepted 15 January 2014 Available online 20 March 2014

Keywords: Dengue DHF DSS Aedes Flavivirus

# **1. Introduction**

Dengue (DEN) virus, a member of the genus *Flavivirus* (family Flaviviridae), is one of the most rapidly spreading mosquito-borne human pathogens in the tropics (Figure 1). DEN viruses are classified into four serotypes (DEN-1, DEN-2, DEN-3 and DEN-4), causing a spectrum of illnesses ranging from a flu-like disease dengue fever (DF) to dengue hemorrhagic fever (DHF), a fulminating illness which can progress to dengue shock syndrome (DSS) and death[1]. Today DF and DHF/DSS are considered the most important arthropod-borne viral diseases in terms of morbidity and mortality<sup>[2]</sup>. The incidence of dengue has grown dramatically around the world in recent decades. Over 2.5 billion people-over 40% of the world's population are now at risk from dengue. WHO currently estimates there

Tel: +919838658557

Dengue is a common pathogenic disease often proving fatal, more commonly affecting the tropics. Aedes mosquito is the vector for this disease, and outbreaks of dengue often cause mass damage to life. The current review is an effort to present an insight into the causes, etiology, symptoms, transmission, diagnosis, major organs affected, mitigation and line of treatment of this disease with special emphasis on drugs of natural origin. The disease has a potential to spread as an endemic, often claiming several lives and thus requires concerted efforts to work out better treatment options. Traditional medicine offers an alternative solution and could be explored as a safer treatment option. Development of a successful vaccine and immunization technique largely remains a challenge and a better antiviral approach needs to be worked out to complement the supportive therapy. No single synthetic molecule has found to be wholly effective enough to offer curative control and the line of treatment mostly utilizes a combination of fluid replacement and antipyretics–analgesics like molecules to provide symptomatic relief.

may be 50–100 million dengue infections worldwide every year. Before 1970, only nine countries had experienced severe dengue epidemics. The disease is now endemic in more than 100 countries in Africa, the Americas, the Eastern Mediterranean, South–east Asia and the Western Pacific. South–east Asia and the Western Pacific regions are the most seriously affected. Since 2003, dengue cases have risen dramatically in Singapore. In 2004, there was a record 9 459 cases notified with eight deaths, with the highest incidence of disease in young adults aged 15–24 years. More than three quarters of all dengue cases in Singapore were hospitalized<sup>[3]</sup>.

The African and Eastern Mediterranean regions have also recorded more outbreaks of the disease in the last ten years. In a recent publication, 22 countries in Africa have reported sporadic cases or outbreaks of dengue from 1960–2010<sup>[4]</sup>. In 2010, indigenous transmission of dengue was also reported in two countries of Europe. Urbanization, rapid movement of people and goods, favorable climatic conditions and lack of trained staff have all contributed to the global increase of dengue. The "official" number of dengue cases in India is 30 000 the actual number, according to the New York Times,

<sup>\*</sup>Corresponding authors: Chawla Pooja, Faculty of Pharmacy – Pharmaceutical Chemistry, BBDNITM, Sector-1, Dr. Akhilesh Das Nagar, Faizabad Road, Chinhut Lucknow Lucknow Uttar Pradesh 226028, India.

Fax: +915223911111

E-mail: pvchawla@gmail.com, pj\_abrol@yahoo.com

is more than 1 000 times that. But the actual dengue cases in India are more than 37 million, according to experts<sup>[5]</sup>.



Figure 1. Mosquito A. aegypti.

# 1.1. Structure

Dengue viruses are spherical, lipid-enveloped that contain a positive strand RNA genome of approximately 10 200 nucleotides coding for three structural proteins (capsid, membrane and envelope) and seven nonstructural proteins (NS1, NS2a, NS2b, NS3, NS4a, NS4b, NS5) (Figures 2 and 3). The envelope protein (E) plays a key role in several important processes including receptor binding, blood cell hemagglutination, induction of a protective immune response, membrane fusion and virion assembly[6]. This is a glycoprotein of approximately 55 kDa, with 3 distinct domains. Using X-ray crystallography, these domains have been characterized for DENV-2 and DENV-3. Domain I is located in the center, and domain II contains an internal fusion loop which is involved in membrane fusion and dimerization of the E protein. Domain III is an immunoglobulin-like domain thought to be involved in cell receptor binding. Domain II is important, as it contains mostly flavivirus group and sub-group crossreactive epitopes. The M-protein, which is important in the formation and maturation of the viral particle, consists of seven antiparallel  $\beta$  –strands stabilized by three disulphide bonds[7].



Figure 2. Structure of dengue virus.



Figure 3. Dengue virus genome.

## 1.2. Transmission

All four serotypes of dengue virus have a similar natural history, including humans as the primary vertebrate host and Aedes mosquitoes of the subgenus Stegomyia [especially Aedes aegypti (A. aegypti), Aedes albopictus (A. albopictus) and Aedes polynesiensis] as the primary mosquito vectors<sup>[8]</sup>. In Africa, and the Indian subcontinent, dengue viruses also exist in enzootic and epizootic forest cycles with nonhuman primates as the vertebrate host<sup>[9,10]</sup>. Other vertebrate species are generally not susceptible to dengue viruses, with the exception of neonatal mice, challenged intracerebrally. After ingestion of a blood meal containing virus, there is infection of the epithelial cells lining the midgut. The virus then escapes from the midgut epithelium into the haemocele and infects the salivary gland. Finally, virus is secreted in the saliva, causing infection during probing. The genital tract is also infected and virus may enter the fully developed egg at the time of oviposition<sup>[11]</sup>. For transmission to occur, the female A. aegypti must bite an infected human during the viraemic phase of the illness which generally lasts 4 to 5 days but may last up to 12 days<sup>[12]</sup>. A. aegypti may be infected with 2 different viruses without affecting the yield of either virus<sup>[13]</sup>. The extrinsic incubation period refers to the time required from itself becomes infective. This period is about 8 to 12 days<sup>[14]</sup>. The feeding behaviour of the mosquito is characterized by easily interrupted feeding and repeated probing of one or several hosts<sup>[15]</sup>.

Whilst the *A. aegypti* has a generally low susceptibility to oral infection with dengue virus, it remains the most important vector because of its highly domesticated habits<sup>[16]</sup>.

The persistence of dengue virus therefore depends on the development of high viral titres in hosts to ensure transmission in mosquitoes. This vector/virus relationship may be a major factor in selecting and propagating pathogenic strains of dengue in the urban setting<sup>[17]</sup>.

# 1.3. Signs and symptoms

In endemic areas, most patients with DF are either asymptomatic or present with mild febrile illness<sup>[18]</sup>. The illness ranges from asymptomatic infection, through undifferentiated fever and benign DF to severe haemorrhagic fever with or without shock syndrome (Figure 4)<sup>[19]</sup>.

Others have more severe illness (5%), and in a small proportion it is life-threatening<sup>[20]</sup>. The incubation period ranges from 3–14 days, but most often it is 4–7 days<sup>[21]</sup>. Therefore, travellers returning from endemic areas are unlikely to have dengue if fever or other symptoms start more than 14 days after arriving home<sup>[22]</sup>. Children often experience symptoms similar to those of the common cold and gastroenteritis (vomiting and diarrhoea)<sup>[23]</sup> and generally have less severe symptoms than adults, but are more susceptible to the severe complications.



Figure 4. Schematic depiction of the symptoms of dengue fever.

#### 1.4. Clinical course

The characteristic symptoms of dengue are sudden-onset fever, headache (typically located behind the eyes), muscle and joint pains and rashes. The alternative name for dengue, "break-bone fever", comes from the associated muscle and joint pains<sup>[24]</sup>. The course of infection is divided into three phases: febrile, critical and recovery. The febrile phase involves high fever, often over 40  $^{\circ}$ C (104  $^{\circ}$ F), and is associated with generalized pain and headache; this usually lasts for two to seven days<sup>[25]</sup>. At this stage, rashes occur in 50-80% of those with symptoms. It occurs in the first or second day of symptoms as flushed skin, or later in the course of illness (days 4-7), as measles-like rashes[11]. Some petechiae (small red spots that do not disappear when the skin is pressed, which are caused by broken capillaries) can appear at this point as well as mild bleeding from the mucous membranes of the mouth and nose. The fever itself is classically biphasic in nature, breaking and then returning for one or two days, although there is wide variation in how often this pattern actually happens[12,13].

In some people, the disease proceeds to a critical phase, which follows the resolution of the high fever and typically lasts for one to two days. During this phase, there may be significant fluid accumulation in the chest and abdominal cavity due to increased capillary permeability and leakage. This leads to depletion of fluid from the circulation and decreased blood supply to vital organs. During this phase, organ dysfunction and severe bleeding, typically from the gastrointestinal tract, may occur. Shock (DSS) and hemorrhage (DHF) occur in less than 5% of all cases of dengue, however those who have previously been infected with other serotypes of dengue virus ("secondary infection") are at an increased risk[<sup>26</sup>].

The recovery phase occurs next, with resorption of the leaked fluid into the bloodstream. This usually lasts for two to three days. The improvement is often striking, but there may be severe itching and a slow heart rate. Another rash may occur with either a maculopapular or a vasculitic appearance, which is followed by peeling of the skin. During this stage, a fluid overload state may occur; if it affects the brain, it may cause a reduced level of consciousness or seizures. A feeling of fatigue may last for weeks afterwards<sup>[8]</sup>.

# 2. Dengue diagnosis

Three factors have been fundamental in dengue diagnosis:

# 2.1. Development of ELISAs for dengue-specific IgM detection

The diagnostic test employed most commonly is IgG and IgM antibodies assay by ELISA. This test is based on an increase in the IgG titre by a factor of four, is difficult in routine clinical care because a second blood sample is required at the convalescent stage<sup>[27]</sup>. Primary infections are characterized by an increase in dengue-specific IgM antibodies four to five days after the onset of fever and by an increase in IgG antibodies. Only after seven to ten days, IgM antibodies are detectable for three to six months, whereas IgG antibodies remain detectable for life. In secondary infections, the level of IgM antibodies is lower than in primary infections and the antibodies are sometimes absent, whereas levels of IgG antibodies rise rapidly in secondary infections, even during the acute phase. Thus, the presence of high titers of IgG early in the course of the disease is a criterion for secondary infection. The sensitivity of IgM ELISA ranges from 90%-97% as compared with the gold standard haemagglutination inhibition test. Some false positive reactions can be observed in less than 2% of cases and a low or negative IgM reaction in secondary infections<sup>[28]</sup>. A number of commercial kits are available in the market used for detection of anti-dengue antibodies[29-36].

# 2.2. Mosquito cell lines and monoclonal antibody development for viral isolation and identification

This method depends on the availability of host-cell cultures or mice that serve as an indicator of virus infection, ie. cytopathic effects in cell culture, sign of illness or death in mice. This is the most common method for virus isolation<sup>[37]</sup>.

Several continuous mosquito cell lines have been known to be highly susceptible to dengue virus infection. The C6/36 clone of *A. albopictus* cells was chosen for virus isolation because it high sensitivity to dengue virus infection and ease of handling<sup>[38,39]</sup>.

Mosquito cell cultures are a recent addition to dengue virus isolation methodologies. Three cell lines of comparable sensitivity are most frequently used. The C6/36 clone of *A. albopictus* cells is less sensitive than the mosquito inoculation method. Use of these cell lines has provided a rapid, sensitive and economical method for dengue virus isolation. The sensitivity mosquito cell lines may vary with the strain of the virus. Even though cell cultures are less sensitive than the mosquito inoculation, large number of samples can be processed in a relatively short time<sup>[40]</sup>. Mammalian cell cultures have many of the same advantages as baby mice for isolation of dengue viruses, such as being expensive, slow and intensive<sup>[41]</sup>.

# 2.3. Reverse transcriptase PCR

In recent years, reverse transcriptase-polymerase chain reaction (RT-PCR), has been developed for a number of RNA viruses, including dengue viruses. Unlike most other techniques which require screening of pools of mosquitoes to detect viruses, RT-PCR carries out the job with solitary specimen. The technique allows for the multifold biological amplification of viral nucleic acid and has been used to rapidly diagnose viral diseases<sup>[42–45]</sup>. The primary advantage of this molecular tool lies in the speed at which specimens can be screened for the presence of dengue viruses and also by its highly sensitive and specific detection. It is able to monitor the infection rate in mosquitoes, both adults and larvae, with a high degree of precision<sup>[46]</sup>. RT–PCR can also detect small quantities of virus-3. This method has also been employed for detecting and typing dengue virus RNA in the field caught Aedes mosquitoes, besides determining the infection rate in local Aedes mosquitoes[47].

These three methods cover the serological, virological, and molecular diagnosis of dengue<sup>[48]</sup>.

# 3. Tissues/Organs affected by dengue

Several body organs are affected by dengue in many ways.

# 3.1. Liver

The spectrum of hepatic involvement in dengue varies from jaundice to elevation in liver enzymes<sup>[49]</sup>. Hepatomegaly and increased serum liver enzymes are the two most clinical evidence of the involvement of liver in dengue infection. It has been reported that among the two, hepatomegaly is more frequent in patients with DHF than in those with DF. Several studies document raised serum transaminase levels in dengue infection (higher in DHF/DSS than in DF), however these tend to return to normal in 14-21 days after infection<sup>[50]</sup>. The enzymes, aspartate aminotransferase (AST) and alanine aminotransferase (ALT), are believed to be highly sensitive indicator of liver damage. The studies conducted by Kuo and colleagues on nearly 240 dengue patients from the 1987-1988 outbreak in Taiwan showed elevated levels of AST and ALT in 93.3% and 82.2% of cases respectively<sup>[51]</sup>. Presence of fever, jaundice and hepatomegaly in endemic areas should arouse the suspicion of dengue hepatitis.

# *3.2. Heart*

Dengue continues to cause significant global morbidity and mortality. Severe disease is characterized by cardiovascular compromise from capillary leakage. Cardiac involvement in dengue has also been reported but has not been adequately studied. Studies have reported patients with severe dengue had septal and right ventricular wall being predominantly affected leading to systolic and diastolic cardiac impairement<sup>[52]</sup>. During episodes of DHF, cardiac rhythm disorders such as atrioventricular blocks, atrial fibrillation, sinus node dysfunction and ectopic ventricular beats have been reported. Cases of complicated dengue viral infection with acute myocarditis involving young male adults have been reported. Two cases were studied of which one was fatal. The first case presented with typical signs of myocardial disease: chest pain and diaphoresis with myocardial depression. The second case deteriorated rapidly and demised within the first day of admission<sup>[53]</sup>. However, the incidence of cardiac complications in patients with dengue illness varies greatly<sup>[54,55]</sup>.

Further studies carried out by Wali *et al* showed that 70% of 17 patients with DHF/DSS who underwent myocardial scintigraphic study suffered diffuse left ventricular hypo kinesis with a mean ejection fraction of 40%<sup>[55]</sup>. Whereas Kabra *et al* reported that 16.7% of 54 children with dengue illness had a decreased left ventricular ejection fraction of <50%<sup>[56]</sup>.

These studies suggest that cardiac complications in dengue illness are not uncommon, but have been under-diagnosed as cases with cardiac complications are clinically mild and self-limited. Although severe cardiac impairment is not commonly reported in dengue infection, it can be life threatening.

# 3.3. Kidney

Dengue shock syndrome is the most severe form of dengue that can be fatal. Nonresponders to standard therapy need intensive care. A report on the clinical features, complications, and outcomes of DSS not responding to standard therapies and needing supportive care in a tertiary referral intensive care unit of a developing country indicated that nearly one-third died within 3 days of admission to ICU. Peritoneal dialysis increased the risk to 100%[57]. Acute renal failure is rare in dengue fever and it mainly presents as shock induced acute tubular necrosis. It has been observed as a complication of dengue fever in French Guiana and was found to occur in 0.3% of cases in a series of 6 154 patients with DHF. Acute renal failure and multiple organ failure can also be a manifestation of rhabdomyolysis. The role of immune complex in development of renal failure in dengue infection is still unclear. Wiwanitkit discovered that the diameter of dengue virus-immunoglobulin complex is much smaller than the diameter of glomerulus. Thus, he postulated

that immune complex can be entrapped only if a previous glomerular lesion causes narrowing of the glomerulus's diameter, and concluded that the immune complex does not play a significant role in pathogenesis of renal failure in dengue infection. Reports of renal failure because of haemolytic uraemic syndrome are rare<sup>[58]</sup>.

# 3.4. Eye

Studies on ocular complication in dengue fever have been reported in literature. These ocular manifestations may be a result of inflammatory changes in vascular endothelium resulting in vascular leakage, haemorrhage and ischaemia. Ocular manifestations reported in DHF are subconjunctival hemorrhage, intraretinal haemorrhage, macular haemorrhage, vitreous hemorrhage, Roth spots, cotton wool spots, retinal oedema, disc oedema or choroidal effusion. A characteristic maculopathy has only rarely been reported and may be due to actual infection of macular tissue. This maculopathy may manifest itself reduced visual acuity, abnormal visual fields or electrophysiological studies. Good recovery is the norm but exceptions have been seen in patients with macular hemorrhages<sup>[59]</sup>. The spectrum of ophthalmologic manifestations would lead one to conclude that several pathophysiologic processes are involved. The first and most obvious pathogenesis would be the thrombocytopenic state, with its resultant bleeding tendency, which gives rise to increased incidence of hemorrhage. These hemorrhages manifest as retinal blot hemorrhages in the macula and retinal periphery. Most patients with dengue-related ophthalmic complications recover spontaneously without any treatment. Despite this, isolated reports of cases of dengue ophthalmic complications with poor visual acuity refractory to treatment have been reported<sup>[60]</sup>.

# 3.5. Blood

In severe condition of dengue, the fibrinolytic system has also been found to get greatly disturbed. Such fibrinolytic abnormalities suggest a stressed haemostatic system that may ultimately result in overt disseminated intravascular coagulation<sup>[61]</sup>.

Thrombocytopenia is common in DF and always found in dengue hemorrhagic fever/ dengue shock syndrome<sup>[62]</sup>. However, the pathogenesis of thrombocytopenia is unknown and it is believed that dengue- virus-induces bone marrow suppression leading to depressed platelet synthesis and thrombocytopenia<sup>[63]</sup>.

Another study reported that dengue-2 virus can bind to human platelets in the presence of virus-specific antibodies, and proposed that the immune-mediated clearance of platelets was involved in the pathogenesis of thrombocytopenia in DHF/DS<sup>[64]</sup>. The titre of IgM antiplatelet antibodies is higher in DHF/DSS than in DF patients. The presence of these auto antibodies not only induces platelet lysis via complement activation, but also inhibits ADP-induced platelet aggregation<sup>[65]</sup>. The production of anti-platelet autoantibodies whose affinities are enhanced in secondary infection not only explains the immune-mediated destruction of platelets, but also raises an important issue of the long-term safety of a dengue vaccine.

# 3.6. Brain

Encephalopathy in DHF is an atypical manifestation and may occur because of various reasons such as intracranial haemorrhage, cerebral oedema, hyponatremia, cerebral anoxia, fulminant hepatic failure with portosystemic encephalopathy, renal failure or release of toxic products. The signs and symptoms of neurological involvement appear in various forms, including depressed sensitivity, convulsions, neck rigidity, pyramidal signs, headache, papilloedema, myoclonus and behavioural disorders. Post– infectious sequelae are mainly amnesia, dementia, manic psychosis, Reye's syndrome and meningo encephalitis. Pathophysiology may include the following factors: direct tissue lesion caused by the virus because of its neurotropicity, capillary haemorrhage, disseminated intravascular coagulation and metabolic disorders<sup>[66]</sup>.

In a study, it was reported that dengue virus was observed in the cerebrospinal fluid (CSF) in five of six patients presenting with encephalitis, indicating that the virus may cross the blood-brain barrier and directly invade the brain. In another study, samples of 150 individuals suspect of an infection disease and with fatal outcomes were investigated for evidence of the presence of DENV. The sampling was made up of 150 CSF, 120 tissue samples, and 109 blood specimens. Out of 150 studied patients, 84 were dengue positive. Evidence of the presence of DENV was found in 41 CSF, showing the following neurologic diagnosis: 46.3% encephalitis, 34.1% meningoencephalitis, and 19.5% meningitis, giving a frequency of 48.8% of the 84 dengue-positive cases. The major clinical manifestations observed on these individuals were fever, headache, mental irritability, breathless, vomiting, muscle pain, tiredness, abdominal pain, somnolence, restlessness, dizziness, cough, seizure, coma, and neck stiffness[67,68].

# 3.7. Pancreas

Acute pancreatitis is a rare complication of dengue fever. Pancreas may be affected due to direct viral invasion or hypotension in dengue hemorrhagic fever. Case studies of dengue lead to an emphasis on the possibility of dengue related pancreatitis in patients with abdominal pain and fever. This will help the patient to receive adequate monitoring and supportive care directed at the management of dengue along with the management of acute pancreatitis. The exact pathogenesis in the development of acute abdomen from infection with dengue virus is unknown. However, it might be due to virus invasion of the gallbladder wall, appendicular wall or pancreas, which causes oedematous change. Recently a case study was reported wherein acute pancreatitis as the complication of DHF was studied. Although rare, this complication can cause more severe fatal condition, and difficulties in treatment. Acute pancreatitis is an uncommon but life-threatening complication of dengue fever<sup>[69]</sup>. Early diagnosis and quick management of dengue related complications is necessary to improve the patient's condition and survival<sup>[70]</sup>.

# 3.8. Effect on Endothelial cells

The endothelium is the primary fluid barrier of the vasculature and ultimately the effects of dengue virus infection that bring about capillary leakage impact endothelial cell barrier functions. Infection of endothelium by dengue virus alters capillary permeability, permits virus replication and induces responses that recruit immune cells to the endothelium<sup>[71]</sup>. In a postmortem study, Jessie et al reported dengue virus antigen in sinusoidal ECs in the liver as well as macrophages, lymphoid cells in the spleen, the vascular endothelium of the lung, monocytes within the blood, and kidney tubules<sup>[72]</sup>. Salgado et al. also demonstrated the presence of viral antigen in endothelial cells within the heart and small myocardial vessels of a patient post-mortem[73]. Leakage of the vascular endothelium is a central component of dengue virus disease and studies discussed here suggest that the dengue infected endothelium may contribute to pathogenic immune responses and immune targeting of the endothelium.

# 3.9. Pregnant patients

Dengue in pregnancy must be carefully differentiated from preeclampsia. An overlap of signs and symptoms, including thrombocytopenia, capillary leak, impaired liver function, ascites and decreased urine output may make this clinically challenging. Pregnant women with dengue fever respond well to the usual therapy of fluids, rest and antipyretics. An awareness of the clinical and laboratory manifestations of dengue in pregnancy should allow its early recognition and the institution of appropriate treatment. If the mother acquires infection in the peripartum period, newborns should be evaluated for dengue with serial platelet counts and serological studies<sup>[74,75]</sup>. Dengue fever in pregnancy most often is treated conservatively. Complications like preeclampsia, pre-term labour, increased risk of caesarean section and fetal transmission have been noted. Platelet count may fall rapidly but no active intervention required unless patient is in labour or has bleeding disorder<sup>[76]</sup>.

## 4. Treatment against dengue

Treatment of dengue may involve following four parameters which may or may not be successful.

# 4.1. Vaccination

The development of a dengue vaccine has unique challenges. The four dengue serotypes circulate globally and infection with one dengue serotype confers life-long protection against re-infection with the same serotype, but only short-term protection against the other 3 serotypes. Moreover, dengue is unique in that sequential infections with different serotypes increase the risk of developing severe and potentially lethal disease[77]. There is limited understanding of how the virus interacts with the immune system and how certain types of pre-existing immunity can exacerbate disease. Therefore, a safe and effective dengue vaccine must be tetravalent and induce strong and longlived protection against all 4 serotypes simultaneously in order to avoid the risk of sensitizing the vaccine recipient to severe disease<sup>[78]</sup>. There are a number of dengue vaccine candidates in different stages of development. The more advanced consist of tetravalent mixtures of live attenuated viruses representing each serotype. Different attenuation mechanisms have been used to develop three of the leading candidates:

a) Chimerization with yellow fever 17D vaccine strain, developed by Sanofi Pasteur.

b) Combinations of defined mutations/deletions and chimeras, developed by NIH.

c) Chimerization with dengue 2 PDK53 virus, attenuated by cell culture passage, developed by Inviragen.

But several disadvantages are associated with all live attenuated vaccine:

1)Single inoculation is not sufficient to induce protection to all 4 serotypes, probably due to viral interference among the live components of the vaccine.

2)Booster doses are not effective when administered less than 6 months apart.

Therefore, live attenuated vaccines require three immunizations over an extended dosing schedule of 12 months to elicit balanced neutralizing antibody responses to all 4 serotypes. As a result, there is a risk that an incomplete response induced by the initial immunizations will enhance disease if infection occurs during the window between the first and the last immunization.

# 4.2. Medication /Treatment

Dengue fever is usually a self-limited illness. There is no specific antiviral treatment currently available for dengue fever. Supportive care with analgesics, fluid replacement and bed rest is usually sufficient<sup>[79]</sup>.

No medication has been found to be useful in treatment

dengue and its association disorder or complication. However, Acetaminophen may be used to treat fever and relieve other symptoms. Aspirin, nonsteroidal antiinflammatory drugs (NSAIDs) and corticosteroids should be avoided. Management of severe dengue requires careful attention to fluid management and proactive treatment of hemorrhage. Single-dose methylprednisolone showed no mortality benefit in the treatment of dengue shock syndrome in a prospective, randomized, double-blind, placebo-controlled trial<sup>[80]</sup>.

The Novartis Institute for Tropical Diseases (NITD) in Singapore is carrying out research to find inhibitors of dengue viral target proteins to reduce the viral load during active infection<sup>[81]</sup>.

Some measures can be taken as a supportive care in dengue fever. They can be classified in to two categories:

#### 4.2.1. For suspected dengue

1) Patients with moderate dehydration caused by high fever and vomiting are recommended oral rehydration therapy.

2) Should have their platelet count and hematocrit measured daily from the third day of illness until 1–2 days after defervescence.

3) Patients with clinical signs of dehydration and a rising hematocrit level or falling platelet count should have intravascular volume deficits replaced under close observation<sup>[82]</sup>.

## 4.2.2. For severe dengue

1)Severe dengue requires careful attention to fluid management and proactive treatment of hemorrhage. Admission to an intensive care unit is indicated for patients with dengue shock syndrome.

2)Patients may need a central intravenous line for volume replacement and an arterial line for accurate blood pressure monitoring and frequent blood tests<sup>[83]</sup>.

3)Intravascular volume deficits should be corrected with isotonic fluids such as Ringer lactate solution.

4)Boluses of 10–20 mL/kg should be given over 20 minutes and may be repeated.

5)If this fails to correct the deficit, the hematocrit value should be determined. If it is rising, limited clinical information suggests that a plasma expander may be administered. Starch, dextran 40, or albumin 5% at a dose of 10–20 mL/kg may be used. One study has suggested that starch may be preferable because of hypersensitivity reactions to dextran<sup>[84]</sup>.

## 4.3. Natural treatment options for dengue fever

Natural drugs possess activity against *A. aegypti* by their antiviral mechanism, larvicidal and mosquitocidal action and mosquito repellants property. Some of the important natural cures are enumerated below.

1) Eupatorium perfoliatum (Boneset): Bonoset is a commonly

available plant which plays an important role in management of dengue fever. It is best consumed in the form of a tea<sup>[85]</sup>.

2) *Boesenbergia rotunda* (Temu kunci): A paste made with the roots is often used for treatment. The use of the herb reduces muscular pain and nausea which causes great distress in people who suffer from this condition<sup>[86]</sup>.

3) *Kaempferia parviflora*: Leaves and stem are used as herbal remedy against virus. Studies have shown that DEN 2 particles are directly inactivated by some bioactive compound in Kaempferia parviflora<sup>[87]</sup>.

4) *Carica papaya*: Juice of the leaves is utilized for increasing the platelet count<sup>[88]</sup>.

5) Solanum villosum: Berry extract is reported to possess larvicidal activity against Stegomyia aegepti<sup>[89]</sup>.

6) *Combretum collinum*: Extract of shoot bark possess larvicidal against *A. aegypti*[90].

7) Azadirecta indica and Pongamia glabra: Herbal formulation named PON-NEEM imparts larvicidal, ovicidal, oviposition deterrant activity against A. aegypti and A. albopictus<sup>[91]</sup>.

8) Nyctanthes arbortistis, Catharanthus roseus, Eupatorium odoratum: Leaf extract exert larvicidal activity against A. aegypti<sup>[92]</sup>.

9) *Citrus limetta*: Extract from peels exert larvicidal activity against *A. aegypti*[93].

10) Acalypha alnifolia: Leaf extract exert larvicidal activity against A. aegypti, Aedes stephensi (A. stephensi)[94].

11) *Delonix elata*: Leaf and seed extracts are reported to exert larvicidal and ovicidal activities against *A. aegypti* and *A. stephensi*[95].

# 4.4. General prevention

The only way to prevent dengue virus acquisition is to avoid being bitten by a vector mosquito. Although this can be accomplished by avoiding travel to areas where dengue is endemic, that is not an ideal strategy because it would require a person to avoid most tropical and subtropical regions of the world, many of which are popular travel and work destinations<sup>[96]</sup>. Other measures are as follows:

a) Apply N,N-diethyl-3-methylbenzamide-containing mosquito repellant;

b) Wear protective clothing, preferably impregnated with permethrin insecticide;

c) Remain in well-screened or air-conditioned places;

d) The use of mosquito netting is of limited benefit, as *Aedes* are day-biting mosquitoes;

e) Eliminate the mosquito vector using indoor sprays[97,98].

#### 5. Conclusion

The authors aimed at presenting an overview on dengue, a common pathogenic disease. An exhaustive, cumulative and in-depth literature review has been used to present the details of the etiology, transmission and treatment models with special emphasis on drugs of natural origin. The pathophysiological changes on several major body organs have been dealt in great detail. The myths regarding the disease need to be avoided and the safe bet as always remains: prevention is better than cure!

# **Conflict of interest statement**

We declare that we have no conflict of interest.

# References

- Henchal EA, Putnak RJ. The dengue viruses. *Clin Microbiol Rev* 1990; **3**: 376–396.
- [2] Pinheiro FP, Corber SJ. Global situation of dengue and dengue haemorrhagic fever and its emergence in the Americas. World Health Stat Q 1997; 50: 161–168.
- [3] Dengue fever reports. Ministry of Health Singapore. 2004.
- [4] Amarasinghe A, Kuritsk JN, Letson GW, Margolis HS. Dengue virus infection in Africa. *Emerg Infectious Dis* 2011; 17: 1349– 1354.
- [5] [Online]Available from: http://www.indiatimes.com/india/actualdengue-cases-in-india-37-million-46458.html [Accessed on Jan, 2013].
- [6] Chambers TJ, Hahn CS, Galler R, Rice CM. Flavivirus genome organization, expression and replication. *Annu Rev Microbiol* 1990; 44: 649–688.
- [7] Lai CY, Tsai WY, Lin SR, Kao CL, Hu HP, King CC, et al. Antibodies to envelope glycoprotein of dengue virus during the natural course of infection are predominantly cross-reactive and recognize epitopes containing highly conserved residues at the fusion loop of domain []. J Virol 2008; 82: 6631–6643.
- [8] Simmons CP, Farrar JJ, Nguyen VV, Wills B. Dengue. N Engl J Med 2010; 366(15): 1423–1432.
- [9] Chen LH, Wilson ME. Dengue and chikungunya infections in travelers. *Curr Opin Infect Dis* 2010; 23: 438–444.
- [10][Online]Available from: http://en.wikipedia.org/wiki/Dengue\_ fever#refWHO2009.
- [11]Wolff K, Johnson RA. Viral infections of skin and mucosa. *Fitzpatrick's color atlas and synopsis of clinical dermatology*. 6th ed. New York: McGraw-Hill Medical; 2009.
- [12]Knoop KJ, Stack LB, Storrow A, Thurman RJ. Tropical medicine. Atlas of emergency medicine. 3rd ed. New York: McGraw-Hill Professional; 2010.
- [13]Gould EA, Solomon T. Pathogenic flaviviruses. *Lancet* 2008; **371**: 500–509.
- [14]Rodenhuis–Zybert IA, Wilschut J, Smit JM. Dengue virus life cycle: viral and host factors modulating infectivity. *Cell Mol Life Sci* 2010; **67**: 2773–2786.
- [15]Pongsumpun P, Tang IM. Effect of the seasonal variation in the extrinsic incubation period on the long term behavior of the dengue hemorrhagic fever. *Epidemic International J Biological*

Life Sci 2007; 3: 208-214.

- [16]Guzman MG, Halstead SB, Artsob H, Buchy P, Farrar J, Gubler DJ, et al. Dengue: a continuing global threat. *Nat Rev Microbiol* 2010; 8(12 Suppl): S7–S16.
- [17]Vector-borne viral infections. World Health Organization. [Online]Available from: http://www.who.int/vaccine\_research/ documents/Vector\_Borne\_Viral\_Infections.[Accessed on Jan 2013].
- [18]Center for Disease Control and Prevention. Chapter 5: dengue fever (DF) and dengue hemorrhagic fever (DHF). 2010 Yellow Book.
- [19]Service MW. Importance of ecology in Aedes aegypti control. Southeast Asian J Trop Med Public Health 1992; 23: 681–690.
- [20]Reiter P. Yellow fever and dengue: a threat to Europe. Euro Surveill 2010; 15: 19509.
- [21]Gubler DJ. The arboviruses: epidemiology and ecology. In: Monath TP. (Ed.). *Boca Raton*. CRC Press; 1988, p. 378–379.
- [22]Singh R, Kissoon N. Dengue hemorrhagic fever and shock syndromes. *Pediatr Crit Care Med* 2010; 12: 90–100.
- [23]Varatharaj A. Encephalitis in the clinical spectrum of dengue infection. *Neurol India* 2010; 58: 585–591.
- [24]Chen LH, Wilson ME. Dengue and chikungunya infections in travelers. *Curr Opin Infect Dis* 2010; 23: 438–444.
- [25]WHO dengue guidelines for diagnosis, treatment, prevention, and control. Geneva: WorldHealthOrganization. [Online] Available from: whqlibdoc.who.int/publications/2009/9789241547871\_eng. pdf. [Accessed on Jan 2013].
- [26]Rodenhuis–Zybert IA, Wilschut J, Smit JM. Dengue virus life cycle: viral and host factors modulating infectivity. *Cell Mol Life Sci* 2010; 67(16): 2773–2786.
- [27]Schwartz E, Mileguir F, Grossman Z, Mendelson E. Evaluation of ELISA based serodiagnosis of Dengue fever in travellers. J Clin Virol 2000; 19: 169–173.
- [28]Jelinek T, Wasthuber J, Proll S, Schtterkirchner M, Loscher T. Influence of rheumatoid factor on the specificity of a rapid immunochromatographic test for diagnosing Dengue infection. *Euro J Clin Microbiol Infect Dis* 2000; **19**: 555–556.
- [29]Vaugn DW, Nisalak A, Solomon T, Kalayanarooj S, Minh Dung N, Kneen R, et al. Rapid serologic diagnosis of dengue virus infection using a commercial capture ELISA that distinguishes primary and secondary infections. *Am J Trop Med Hyg* 1999; **60**: 693–698.
- [30]Allwinn R, Schieferstein C, Glauke S, Doerr HW. Rapid diagnosis of primary dengue fever by the immunochromatographic test and by electron microscopy–A case report. *Infection* 1999; 27: 365– 367.
- [31]Palmer CJ, King SD, Cuadrado RR, Perez E, Baum M, Ager AL. Evaluation of the MRL diagnostics dengue fever virus IgM capture ELISA and the PanBio Rapid Immunochromatographic Test for diagnosis of dengue fever in Jamaica. *J Clin Microbiol* 1999; 37: 1600–1601.
- [32]Kuno G, Cropp CB, Wong–Lee J, Gubler DJ. Evaluation of an IgM immunoblot kit for dengue diagnosis. *Am J Trop Med Hyg* 1998; 59: 757–762.
- [33]Cuzzubbo AJ, Vaughn DW, Nisalak A, Solomon T, Kalayanarooj S, Aaskov J, et al. Comparison of PanBio dengue Duo IgM and IgG

capture ELISA and venture technologies dengue IgM and IgG Dot Blot. *J Clin Virol* 2000; **16**: 135–144.

- [34]Wu SJL, Paxton H, Hanson B, Kung CG, Chen TB, Rossi C, et al. Comparison of two rapid diagnostic assays for detection detection of immunoglobulin M antibodies to dengue virus. *Clin Diagn Lab Immunol* 2000; 7: 106–110.
- [35]Cuzzubbo AJ, Endy TP, Nisalak A, Kalayanarooj S, Vaughn DW, Ogata SA, et al. Use of recombinant envelope proteins for serological diagnosis of dengue virus infection in a immunochromatographic assay. *Clin Diagn Lab Immunol* 2001; 8: 1150-1155.
- [36]Laferte J, Pelegrino JL, Guzman MG, Gonzalez G, Vazquez S, Hermida C. Rapid diagnosis of dengue virus infection using a novel 10 μ L IgM antibody capture ultramicro ELISA assay (MAC UMELISA Dengue). Adv Mod Biotech 1992; 1: 19.4.
- [37]Ilkal MA, Dhanda V, Rodrigues JJ, Mohan Rao CVR, Mourya DT. Xenodiagnosis of laboratory acquired infection by mosquito inoculation and immunofluorescence. *Indian J Med Res* 1984; 79: 587–590.
- [38]Yull TM, Sukhavachana P, Nisalak A, Russell PK. Denguevirus recovery by direct and delayed plaques in LLC–MK2 cells. Am J Trop Med Hyg 1968; 17: 441.
- [39]Race MW, Williams MC, Agostini CFM. Dengue in the Caribbean: virus isolation in a mosquito (Aedes pseudoscutellaris) cell line. *Trans R Soc Trop Med Hyg* 1979; **73**: 18–22.
- [40]Philip Samuel P, Tyagi BK. Diagnostic methods for detection & isolation of dengue viruses from vector mosquitoes. *Indian J Med Res* 2006; **123**: 615–628.
- [41]Tesh RB. A method for the isolation and identification of dengue viruses using mosquito cell cultures. Am J Trop Med Hyg 1979; 28: 1053–1059.
- [42]Maneekarn N, Morita K, Tanaka M, Ugarashi A, Usawattanakul W, Srisanthana V, et al. Application of polymerase chain reaction for identification of dengue virus isolated from patient sera. *Microbiol Immunol* 1993; **37**: 41–47.
- [43]Saiki RK, Gelfand DH, Stoffel S, Scharf SJ, Higuchi R, Horn GT, et al. Primer directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science* 1988; 239: 487–491.
- [44]Henchal EA, Polo SL, Yaemsiri VVC, Hoke CH. Sensitivity and specificity of a universal primer set for the rapid diagnosis of dengue virus infections by polymerase chain reaction and nucleic acid hybridization. *Am J Trop Med Hyg* 1991; **45**: 418–428.
- [45]Morita K, Tanka M, Igarashi A. Rapid identification of dengue virus serotype by polymerase chain reaction. J Clin Microbiol 1991; 29: 2107–2110.
- [46]Rohani A, Ismail A, Saat Z, Lee HL. Detection of dengue virus from field Aedes aegypti and Aedes albopictus adults and larvae. *Southeast Asian J Trop Med Public Health* 1997; 28: 138–144.
- [47]Pinheir VC, Tadei WP, Barros PM, Vascocelos PF, Cruz AC. Detection of dengue virus serotype 3 by reverse transcription– polymerase chain reaction in Aedes aegyptis Diptera–Culicidae captured in Manaus, Amazonas. *Mem Inst Oswaldo Cruz* 2005; 100: 833–839.
- [48]Guzmán MG, Kourí G. Advances in dengue diagnosis. Clin Diagnostic Immunol 1996; 3: 621-627.

- [49]Jagadishkumar K, Jain P, Manjunath VG, Umesh L. Hepatic involvement in dengue fever in children. *Iran J Pediatr* 2012: 22: 231–236.
- [50]Butt N, Abbassi A, Munir SM, Ahmad SM, Sheikh QH. Haematological and biochemical indicators for the early diagnosis of dengue viral infection. *J Coll Physicians Surg Pak* 2008; 18: 282–285.
- [51]Kuo CH, Tai DI, Chang-Chien CS, Lan CK, Chiou SS, Liaw YF. Liver biochemical tests and dengue fever. Am J Trop Med Hyg 1992; 47: 265–270.
- [52]Yacoub S, Griffiths A, Chau TT, Simmons CP, Wills B, Hien TT, et al. Cardiac function in Vietnamese patients with different dengue severity grades. *Crit Care Med* 2012; **40**: 477–483.
- [53]Marques N, Gan VC, Leo Y–S. Dengue myocarditis in Singapore: two case reports. *Infection* 2013. doi: 10.1007/s15010-012-0392-9.
- [54]Chuah SK. Transient ventricular arrhythmia as a cardiac manifestation in dengue haemorrhagic fever: a case report. *Singapore Med J* 1987; 28: 569–572.
- [55]Wali JP, Biswas A, Chandra S, Malhotra A, Aggarwal P, Handa R, Wig N, Bahl VK. Cardiac involvement in Dengue Haemorrhagic Fever. Int J Cardiol 1998; 13; 64(1): 31–36.
- [56]Kabra JK, Juneja R, Madhulika J, Jain Y, Singhal T, Dar L, et al. Myocardial dysfunction in children with dengue haemorrhagic fever. *Natl Med J India* 1998; 11: 59–61.
- [57]Goonasekera CDA, Thenuwara BG, Kumarasiri RPV. Peritoneal dialysis in dengue shock syndrome may be detrimental. J Trop Med 2012; 1–5. doi:10.1155/2012/917947.
- [58]Lum LCS, Thong MK, Cheah YK, Lam SK. Dengue associated adult respiratory distress syndrome. Ann Trop Paediatrics 1995; 15: 335–339.
- [59]Haritoglou C, Scholz F, Bialasiewicz A, Klauss V. Ocular manifestation in dengue fever. *Ophthalmologie* 2000; 97: 433– 436.
- [60]Cherng–Hui Yip V, Sanjay S, Koh YT. Ophthalmic complications of dengue fever: a systematic review. *Ophthalmol Ther* 2012; 1: 2. DOI 10.1007/s40123–012–0002–z.
- [61]Mairuhu ATA, Mac Gillavry MR, Setiati TE, Soemantri A, ten Cate H, Brandjes DPM, et al. Coagulation and fibrinolysis in dengue fever. *Lancet Infect Dis* 2003; **3**: 33–41.
- [62]Bhamarapravati N. Hemostatic defects in dengue haemorrhagic fever. J Infect Dis 1989; (suppl 4): S826–S829.
- [63]La Russa VF, Innis BL. Mechanisms of dengue virus-induced bone marrow suppression. *Baillières Clin Haematol* 1995; 8: 249– 270.
- [64]Wang S, He R, Patarapotikul J, Innis BL, Anderson R. Antibody– enhanced binding of dengue–2 virus to human platelets. *Virology* 1995; **213**: 254–257.
- [65]Lin CF, Lei HY, Liu CC, Liu HS, Yeh TM, Wang ST, et al. Generation of IgM anti-platelet autoantibody in dengue patients. J Med Virol 2001; 63: 143–149.
- [66]Gubler DJ, Kuno G. Dengue and dengue hemorrhagic fever: its history and resurgence as a global health problem. In: *Dengue and dengue hemorrhagic fever*. Willingford, UK: CAB International; 1997, p. 1–22.

- [67]Araújo FM, Araújo MS, Nogueira RM, Brilhante RS, Oliveira DN, Rocha MF, et al. Central nervous system involvement in dengue: a study in fatal cases from a dengue endemic area. *Neurology* 2012; 78: 736–742.
- [68]Miagostovich MP, Ramos RG, Nicol AF, Nogueira RMR, Cuzzi-Maya T, Oliveira AV, et al. Retrospective study on dengue fatal cases. *Clinical Neuropathol* 1997; 16: 204–208.
- [69]Simadibrata M, Acute pancreatitis in dengue hemorrhagic fever. Acta Med Indones 2012; 44: 57–61.
- [70]Karoli R, Fatima J, Singh G, Maini S. Acute pancreatitis: An unusual complication of Dengue Fever. JAPI 2012; 60: 64–65.
- [71]Dalrymple NA, Mackow ER. Roles for endothelial cells in Dengue virus infection. Advances Virol 2012. doi:10.1155/2012/840654.
- [72]Jessie K, Fong MY, Devi S, Lam SK, Wong KT. Localization of dengue virus in naturally infected human tissues, by immunohistochemistry and in situ hybridization. *J Infectious Dis* 2004; 189: 1411-1418.
- [73]Salgado DM, Eltit JM, Mansfield K, Panqueba C, Castro D, Vega MR, et al. Heart and skeletal muscle are targets of dengue virus infection. *Pediatric Infectious Dis J* 2010; 29: 238–242.
- [74]Innis BL, Nisalak A, Nimmannitya S, Kusalerdchariya S, Chongswasdi V, SuntayakornS, et al. An enzymelinked immunosorbent assay to characterize dengue infections where dengue and Japanese encephalitis cocirculate. Am J Trop Med Hyg 1989; 40: 418-427.
- [75]Chitra TV, Panicker S. Maternal and fetal outcome of dengue fever in pregnancy. J Vector Borne Dis 2011; 48: 210–213.
- [76]Perret C, Chanthavanich P, Pengsaa K, Limkittikul K, Hutajaroen P, Bunn JE, et al. Dengue infection during pregnancy and transplacental antibody transfer in Thai mothers. *J Infect* 2005; 51: 287–293.
- [77]Gibbons RV, Vaughn DW. Dengue: an escalating problem. *BMJ* 2002; **324**: 1563–1566.
- [78]Putnak JR, Coller BA, Voss G, Vaughn DW, Clements D, Peters I, et al. An evaluation of dengue type-2 inactivated, recombinant subunit, and live-attenuated vaccine candidates in the rhesus macaque model. *Vaccine* 2005; 23: 4442–4452.
- [79]Maves RC, Ore RM, Porter KR, Kochel TJ. Immunogenicity and protective efficacy of a psoralen-inactivated dengue-1 virus vaccine candidate in Aotus nancymaae monkeys. *Vaccine* 2011; 29: 2691–2696.
- [80]Tassniyom S, Vasanawathana S, Chirawatkul A, Rojanasuphot S. Failure of high-dose methylprednisolone in established dengue shock syndrome: a placebo-controlled, double-blind study. *Pediatrics* 1993; 92: 111-115.
- [81]WHO. Dengue. [Online] Available from: http://www.who.int/ topics/dengue/en/. [Accessed on October, 2011].
- [82]Wichmann O, Stark K, Shu PY, Niedrig M, Frank C, Huang JH. Clinical features and pitfalls in the laboratory diagnosis of dengue in travellers. *BMC Infect Dis* 2006; 6: 120.
- [83]Wills BA, Nguyen MD, Ha TL, Dong TH, Tran TN, Le TT, et al. Comparison of three fluid solutions for resuscitation in dengue shock syndrome. N Engl J Med 2005; 353: 877–889.
- [84]Yadav SP, Sachdeva A, Gupta D, Sharma SD, Kharya G. Control of massive bleeding in dengue hemorrhagic fever with severe

thrombocytopenia by use of intravenous anti–D globulin. *Pediatr* Blood Cancer 2008; **51**(6): 812–813.

- [85]Innvista.com. Boneset. [Online] Available from: http://www. innvista.com/health/herbs/boneset.htm [Accessed on Jan 2013].
- [86]Tan SK, Pippen R, Yusof R, Ibrahim H, Khalid N, Rahman NA. Inhibitory activity of cyclohexenyl chalcone derivatives and flavonoids of fingerroot, *Boesenbergia rotunda* (L.), towards dengue-2 virus NS3 protease. *Bioorg Med Chem Lett* 2006; 16: 3337–3340.
- [87]Moon HI, Cho SB, Lee JH, Paik HD, Kim SK. Immunotoxicity activity of sesquiterpenoids from black galingale (Kaempferia parviflora Wall. Ex. Baker) against Aedes aegypti L. Immunopharmacol Immunotoxicol 2011; 33: 380-383.
- [88]Ahmad N, Fazal H, Ayaz M, Abbasi BH, Mohammad I, Fazal L. Dengue fever treatment with *Carica papaya* leaves extracts. *Asian Pacific J Trop Biomed* 2011; 330–333.
- [89]Chowdhury N, Ghosh A, Chandra G. Mosquito larvicidal activities of Solanum villosum berry extract against the dengue vector Stegomyia aegypti. BMC Complement Altern Med 2008. doi: 10.1186/1472-6882-8-10.
- [90]Odda J, Kristensen S, Kabasa J, Waako P. Larvicidal activity of Combretum collinum Fresen against Aedes aegypti. J Vector Borne Dis 2008; 45: 321–324.
- [91]Maheswaran R, Ignacimuthu S. A novel herbal formulation against dengue vector mosquitoes *Aedes aegypti* and *Aedes albopictus*. *Parasitol Res* 2012; **110**: 1801–1813.
- [92]Alam MF, Safhi MM, Chopra AK, Dua VK. Toxicological properties of several medicinal plants from the Himalayas (India) against vectors of malaria, filariasis and dengue. *Trop Biomed* 2011; 28: 343–350.
- [93]Kumar S, Warikoo R, Mishra M, Seth A, Wahab N. Larvicidal efficacy of the *Citrus limetta* peel extracts against Indian strains of *Anopheles stephensi* Liston and *Aedes aegypti* L. *Parasitol Res* 2012; 111: 173–178.
- [94]Kovendan K, Murugan, K, Vincent, S. Evaluation of larvicidal activity of Acalypha alnifolia Klein ex Willd. (Euphorbiaceae) leaf extract against the malarial vector, Anopheles stephensi, dengue vector, Aedes aegypti and Bancroftian filariasis vector, Culex quinquefasciatus (Diptera: Culicidae). Parasitol Res 2012; 110: 571-581.
- [95]Marimuthu G, Rajamohan S, Mohan R, Krishnamoorthy Y. Larvicidal and ovicidal properties of leaf and seed extracts of *Delonix elata* (L.) Gamble (family: Fabaceae) against malaria (*Anopheles stephensi* Liston) and dengue (*Aedes aegypti* Linn.) (Diptera: Culicidae) vector mosquitoes. *Parasitol Res* 2012; **111**: 65–77.
- [96]Billingsley PF, Foy B, Rasgon JL. Mosquitocidal vaccines: a neglected addition to malaria and dengue control strategies. *Trends Parasitol* 2008; 24: 396–400.
- [97]Erlanger TE, Keiser J, Utzinger J. Effect of dengue vector control interventions on entomological parameters in developing countries: a systematic review and meta–analysis. *Med Vet Entomol* 2008; 22: 203–221.
- [98]Kay B, Vu SN. New strategy against Aedes aegypti in Vietnam. Lancet 2005; 365: 613–617.