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# Inhibition of advanced glycation endproducts formation by Korean thistle, *Cirsium maackii*

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#### ABSTRACT

Objective: To evaluate inhibitory potential of seven Korean thistles against the advanced glycation endproducts (AGE) formation as well as to identify responsible compounds from the most active species. Methods: We used an in vitro AGE inhibition assay to evaluate the antidiabetic complication potential of the methanol extracts of the selected Korean thistles. Results: Among the seven Korean thistles, the leaves of Cirsium maackii (C. maackii) exhibited the most significant inhibitory activity against AGE formation. By means of bioassay-directed fractionation, a lignan, chlorogenic acid and 14 flavonoids were isolated from the active ethyl acetate soluble fraction of a methanol extract from C. maackii leaves. Luteolin and its 5-O-glucoside have been previously isolated; however, a lignan and 13 known compounds were isolated for the first time from C. maackii leaves in this study. Most of the isolated compounds exhibited inhibitory activities against potential AGE formation. Among them, cernuoside was shown to be the most potent AGE inhibitor with an IC<sub>50</sub> value of 21.21  $\mu$  mol/L. Most importantly, two major flavonoids, luteolin and its 5–O-glucoside, also significantly inhibited AGE formation, with IC<sub>50</sub> values of 36.33 and 37.47  $\mu$  mol/L, respectively. Structure activity relationship revealed that the presence of free 3' and 4' dihydroxyl group in flavonoids skeleton played an important role in AGE inhibition. Conclusions: These results indicate that C. maackii and C. maackii-derived flavonoids might be explored further to develop therapeutic agents for the prevention of diabetic complications due to their significant inhibitory activity against AGE formation.

# 1. Introduction

Persistent hyperglycemia induces the formation of reactive oxygen species (ROS) and advanced glycation endproducts (AGE), elevates the activity of aldose reductase, and activates protein kinase C isoforms, all of which play a key role in the pathogenesis of many long-term diabetic complications such as retinopathy, cataractogenesis, nephropathy and neuropathy<sup>[1]</sup>. In particular, increased formation of AGE due to prolonged hyperglycemia and carbonyl stress in diabetes and their subsequent accumulation in various tissues has been implicated in the pathogenesis of many diseases, including atherosclerosis, cardiac dysfunction, and vascular inflammation<sup>[2,3]</sup>. AGE are a chemically heterogeneous group of compounds formed in the Maillard reaction or browning reaction, when reducing sugars react non-enzymatically with amine residues, predominantly lysine and arginine, on proteins, lipids, and nucleic acids, ultimately leading to their chemical modification<sup>[4]</sup>. Recently, AGE formation has drawn considerable attention due to its involvement in the development of vascular complications during atherogenesis. During AGE generation, highly reactive oxygen/nitrogen species and dicarbonyl compounds are produced as intermediates that may attack numerous biomolecules. AGE also contribute to tissue injury by altering extracellular matrix structures through the formation of protein cross-links and modifying shortlived intracellular proteins such as metabolic enzymes and mitochondrial protein complexes<sup>[5]</sup>. These observations led to the development of numerous AGE inhibitors with diverse

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chemical structures for use as possible therapeutic agents. Recently, there has been a growing interest in alternative therapies, especially in the therapeutic use of plant-derived natural products for the management of diabetes due to lower toxicity and fewer side effects than synthetic drugs.

Thistles, all members of the Compositae family, are perennial plants, 0.5–2.0 m in height, possessing lance– shaped, spiny–toothed leaves and white to purple flowers. The thistles grow abundantly in Korea, Japan, and China<sup>[6,7]</sup>. Thus far, 250 species of thistles have been identified worldwide, and about 10 of these species have been found in Korea<sup>[8]</sup>. Because thistle has been used in traditional folk medicine as a diuretic, antiphlogistic, hemostatic, and detoxifying agent<sup>[9,10]</sup>, most thistles have been surveyed by phytochemists and pharmacologists. Especially, milk thistle is one of the most popular herbal remedies used by patients with liver disease<sup>[11]</sup>. A great deal of research has been conducted regarding the flavonoids harbored by thistles, and a variety of activities are exhibited by these species<sup>[12–15]</sup>.

Previously, we reported that there is a correlation between antioxidant activity and the HPLC profiles of several Korean thistles<sup>[16]</sup>, and the rat lens aldose reductase inhibitory activity of *C. maackii* and its major active constituents<sup>[17]</sup>. As a part of our ongoing search for anti-diabetic agents derived from natural sources, we investigated the AGE formation inhibitory activities of seven Korean thistles including *Cirsium chanroenicum (C. chanroenicum), Cirsium lineare (C. lineare), Cirsium maackii (C. maackii), Cirsium nipponicum (C. nipponicum), Cirsium pendulum (C. pendulum), Cirsium setidens (C. setidens), and Carduus crispus (C. crispus) and identified compounds responsible for AGE inhibitory activities from the most active species.* 

# 2. Materials and methods

# 2.1. General experimental procedures

The <sup>1</sup>H– and <sup>13</sup>C–NMR spectra were measured using a JEOL JNM ECP–400 spectrometer (JEOL, Japan) at 400 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C. Column chromatography was performed using silica (Si) gel 60 (70–230 mesh, Merck, Darmstadt, Germany) and Sephadex LH–20 (20–100  $\mu$  m, Sigma, St. Louis, MO, USA). All TLC analyses were conducted on precoated Merck Kieselgel 60 F254 plates (20 cm×20 cm, 0.25 mm, Merck) using 50% H<sub>2</sub>SO<sub>4</sub> as a spray reagent.

#### 2.2. Plant extracts

Methanolic extracts of *C. lineare* collected in September 2002, *C. chanroenicum* in September 2003, *C. setidens* in September 2001, *C. nipponicum* in August 2001, *C. pendulum* in July 2003, and *C. crispus* in May 2005, were purchased from the Korean Plant Extract Bank under the Korea Research Institute of Bioscience and Biotechnology, Korea in February 2006. Whole *C. maackii* plants were collected in Ulsan, Republic of Korea, in August 2007. The plants were authenticated by a specialist in Cirsium taxonomy, Dr. Y.

Kadota at the Department of Botany, National Museum of Nature and Science in Tsukuba, Japan. Each whole plant was separated into flowers, leaves, roots, and stem and dried in the shade for a week. A whole plant was registered and deposited as a voucher specimen at the herbarium of the National Museum of Nature and Science in Tsukuba, Japan as well as at the Department of Food Science and Nutrition, Pukyong National University (Professor J.S. Choi).

# 2.3. Chemicals and reagents

Bovine serum albumin (BSA), aminoguanidine hydrochloride, D-(-)-fructose, D-(+)-glucose, and dimethyl sulfoxide (DMSO) were purchased from Sigma Chemical Co. (St. Louis, MO, USA). Sodium azide was purchased from Junsei Chemical Co. (Tokyo, Japan). All solvents used were purchased from Merck, Duksan Pure Chemical Co., unless stated otherwise.

# 2.4. Extraction, fractionation and isolation

The powder from leaves of C. maackii (1.28 kg) was refluxed with methanol (MeOH) for 3 h (4 L×5). The total filtrate was then concentrated and dried in vacuo at 40  $^\circ C$  in order to render the MeOH extract (316.3 g). This extract was suspended in distilled water and successively partitioned with methylene chloride (CH<sub>2</sub>Cl<sub>2</sub>), ethyl acetate (EtOAc), and *n*-butanol (*n*-BuOH) to yield the CH<sub>2</sub>Cl<sub>2</sub> (68.8 g), EtOAc (80.6 g), and n-BuOH (30.7 g) fractions, respectively, as well as an aqueous residue (125.8 g). The EtOAc fraction (80.0 g) was first chromatographed over a silica gel column (8 cm×80 cm) using a mixed solvent of CH<sub>2</sub>Cl<sub>2</sub> and MeOH (CH<sub>2</sub>Cl<sub>2</sub>:MeOH = 20:1 to 1:1, v:v, gradient) to afford 16 subfractions (F01-F16). Compounds apigenin (22 mg), luteolin (10.5 g) and quercetin (28 mg) were separately purified from EF01 (0.8 g), EF03 (13.9 g), and EF04 (0.5 g), respectively, with a solvent mixture of *n*-hexane and EtOAc (*n*-hexane:EtOAc = 8:1. v:v). Silica gel column chromatography of combined EF05 (2.38 g) and EF06 (2.43 g) using identical solvent conditions led to the isolation of apigenin 7–O-glucuronide methyl ester (48 mg), cernusoside (56 mg) and apigenin 7-O-glucuronide (27 mg). Compounds luteolin 5–O–glucoside (18.0 g), luteolin 7–O– glucoside (35 mg), tracheloside (16 mg), diosmin (230 mg), naringenin 5-0-glucoside (25 mg), eriodictyol 5-0glucoside (15 mg), and luteolin 4'-O-glucoside (23 mg) were purified from EF08 (28.2 g), EF09 (0.5 g), and combined EF10 (1.5 g), EF11 (1.3 g) and EF12 (1.2 g) on Sephadex LH-20 columns (100% MeOH), respectively. Repeated chromatography of EF14 (1.1 g) over a silica gel column with CH<sub>2</sub>Cl<sub>2</sub>:MeOH:H<sub>2</sub>O (26:14:4, v:v) afforded quercetin 3-Oglucoside (130 mg). Fraction EF13 (3.0 g) was recrystallized from 100% MeOH, yielding chlorogenic acid (350 mg). The combined EF14 and EF15 (7.4 g) was then chromatographed over a silica gel column with CH<sub>2</sub>Cl<sub>2</sub>:MeOH:H<sub>2</sub>O (26:14:4, v:v) to give rutin (450 mg). The isolated compounds were identified and characterized by different spectroscopic methods, including <sup>1</sup>H- and <sup>13</sup>C-NMR as well as by comparisons with published spectral data and TLC analyses.

#### 2.5. Inhibition of AGE formation

Inhibition of AGE formation was determined according to the modified method of Vinson and Howard<sup>[18]</sup>. To prepare the AGE reaction solution, 10 mg/mL BSA in 50 mmol/L sodium phosphate buffer (pH 7.4), with 0.2 g/L sodium azide to prevent bacterial growth, was added to 0.2 mol/L fructose and 0.2 mol/L glucose. The reaction mixture (950  $\mu$  L) was then mixed with various concentrations of the samples (50  $\mu$  L, final concentration: 0.20–200.00  $\mu$  g/mL for the extracts and compounds; and 0.10–100.00  $\mu$  g/mL for the fractions) dissolved in 10% (v/v) DMSO. After incubation at 37 °C for 7 d, the fluorescence intensity of the reaction products was determined using a spectrofluorometric detector (FLx800 microplate fluorescence reader, Bio-Tek Instruments, Inc., Winooski, VT, USA), with respective excitation and emission wavelengths at 350 and 450 nm. The percent inhibition of AGE formation was determined from a graphical plot of the data and is expressed as the mean±SEM (triplicate experiments). Aminoguanidine, a nucleophilic hydrazine compound, was used as a reference compound in the AGE assay.

# 2.6. Statistical analysis

Statistical significance was analyzed using one–way ANOVA and Student's *t*–test (Systat In., Evanston, IL., USA), with P<0.01 considered to be statistically significant. All results are expressed as mean±SEM from three experiments.

## 3. Results

# 3.1. AGE formation inhibitory activities of the selected Korean thistles

The inhibitory activities of the MeOH extracts of the tested Korean thistles on AGE formation were investigated. The MeOH extracts from the roots, whole plants, stem, and leaves of the same C. maackii exerted good inhibitory activities against AGE formation, with IC<sub>50</sub> values of  $(129.31\pm5.29)$ ,  $(114.23\pm7.04)$ ,  $(62.74\pm1.41)$ , and  $(28.20\pm0.84)$  $\mu$  g/mL, respectively, which are comparable to the IC<sub>50</sub> value of (149.04 $\pm$ 11.27)  $\mu$  g/mL for the positive control, aminoguanidine (128.08  $\mu$  g/mL). The inhibitory activity increased in the order of leaves > stem > whole plants > roots of the same C. maackii plant. In contrast, the MeOH extracts from the C. maackii flower, from the whole C. crispus, C. nipponicum, C. pendulum, C. lineare, and C. setidens plants, and both the aerial and underground parts of C. chanroenicum did not show any inhibitory activity at concentrations up to 200.00  $\mu$  g/mL.

# 3.2. AGE formation inhibitory activities of different solvent soluble fractions of C. maackii

Because the MeOH extract of *C. maackii* had marked AGE inhibitory activity, we partitioned the extract with several solvents and further assessed the AGE inhibitory activity of

the fractions at concentrations ranging from 1.00 to 100.00  $\mu$  g/mL. The EtOAc and *n*-BuOH fractions among the several solvent-soluble fractions obtained from the MeOH extract showed significant AGE inhibitory activities with IC<sub>50</sub> values of (2.78±0.04) and (87.84±4.91)  $\mu$  g/mL, respectively, compared to aminoguanidine with the IC<sub>50</sub> value of (128.08±6.89)  $\mu$  g/mL. However, the CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O fractions did not show any activity at the test concentrations.

# 3.3. AGE formation inhibitory activities of the isolated compounds from C. maackii

Since the EtOAc fraction was found as the most active fraction, it was selected for chromatography in order to isolate active compounds. Repeated chromatography of the EtOAc fraction yielded 16 compounds, including a lignan, chlorogenic acid, and 14 flavonoids. The AGE formation inhibitory activities of the compounds isolated from C. maackii are summarized in the Table 1. As shown in the Table 1, most of the isolated compounds exhibited promising AGE formation inhibitory activities. Among them, cernuoside was shown to be the most potent AGE inhibitor followed by luteolin 7-O-glucoside and quercetin with their respective  $IC_{50}$  values of (21.21±0.25), (24.22±0.29) and  $(24.35\pm0.95)$   $\mu$  mol/L, compared to the positive control aminoguanidine. In addition, chlorogenic acid, luteolin and its glycosides, quercetin 3-O-glucoside, and rutin also showed strong inhibitory activities against AGE formation. Interestingly, although apigenin showed weak inhibitory activity, its two glycosides, apigenin 7-0-glucuronide methyl ester and apigenin 7-O-glucuronide, did not show inhibitory activity at the concentration tested.

#### Table 1

Inhibition of AGE formation by compounds isolated from EtOAc fraction of *C. maackii* leaves.

Compounds	$\rm IC_{50}$ ( $\mu$ g/mL) $^{\rm a}$	$IC_{50}(\mu M)^{b}$
Apigenin (1)	152.52±6.95	$564.89 \pm 25.72$
Luteolin (2)	10.40±0.03	36.33±0.10
Quercetin (3)	7.35±0.29	24.35±0.95
Apigenin7-O-glucuronide methyl	>200	-
ester (4)		
Cernuoside (5)	9.50±0.11	21.21±0.25
Apigenin 7–0–glucuronide (6)	>200	-
Luteolin 5–0–glucoside (7)	16.80±0.13	37.47±0.29
Luteolin 7–0–glucoside (8)	10.85±0.13	24.22±0.29
Tracheloside (9)	>200	-
Diosmin (10)	41.20±0.56	137.21±1.87
Naringenin 5– <i>O</i> –glucoside (11)	45.25±0.99	104.26±2.28
Eriodictyol 5-0-glucoside (12)	28.77±0.66	63.93±1.47
Luteolin 4´-O-glucoside (13)	15.24±0.07	33.87±0.16
Quercetin 3-0-glucoside (14)	18.80±0.43	47.00±1.13
Chlorogenic acid (15)	10.36±0.01	29.28±0.04
Rutin (16)	31.99±0.74	52.44±1.22
Aminoguanidine	129.36±1.60	1 170.15±14.47

<sup>a</sup>Test concentrations of samples were 0.20–200.00  $\mu$  g/mL dissolved in DMSO. IC<sub>50</sub> is expressed as the mean±SEM from triplicate experiments.

<sup>b</sup>Aminoguanidine was used as a positive control.

# 4. Discussion

Persistent hyperglycemia induces increased formation of AGE, which has been implicated in the onset of diabetic complications. In this point of view, we investigated the AGE formation inhibitory activities of the selected Korean thistles using an in vitro AGE formation inhibitory assay. Interestingly, only different parts of C. maackii, which was previously found to be a good inhibitor of rat lens aldose reductase, exhibited potent inhibitory activities against AGE formation. The significant variability in AGE inhibitory activities between Cirsium species or even between parts of the same plant might result from differences in the active components between species and different parts of the same plant, leading to variation in the inhibitory effect on diabetic complications. Therefore, the present study was designed to identify the active compounds responsible for potent AGE inhibitory activity of C. maackii in addition to the relationship between their structure and function.

Solvent partitioning of the MeOH extract of C. maackii vielded four subfractions, among which EtOAc and *n*-BuOH fractions showed significant AGE inhibitory activities. However, the CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O fractions did not show any activity at the test concentrations. It is interesting to note that both EtOAc and n-BuOH fractions had marked AGE inhibitory activity, whereas the other fractions showed weak or no activities at the tested concentrations. Plants possess a range of nonpolar to polar phytochemicals. The comparative inhibitory activities of several fractions derived from the MeOH extract of C. maackii leaves can be explained by their compositional differences. Flavonoids were expected to be present in large amounts in the polar EtOAc and n-BuOH fractions. From a health point of view, this may be meaningful to consumers because most of the components in the EtOAc and *n*-BuOH fractions are easily dissolved in water. In the present study, from the most active EtOAc fraction of the MeOH extract of the C. maackii leaves, a lignan, chlorogenic acid, and 14 known flavonoids were isolated through repeated column chromatography and were subsequently characterized by <sup>1</sup>H- and <sup>13</sup>C-NMR, DEPT, HMQC, and HMBC, as well as comparisons with published spectral data. luteolin and its 5-0-glucoside had been previously isolated, however, a lignan, along with 13 known compounds were isolated from C. maackii leaves for the first time in this study[19-30].

Besides apigenin, luteolin, quercetin, apigenin 7–glucuronide and its methyl ester, chlorogenic acid and rutin were previously described as AGE inhibitors, cernuoside, luteolin 4', 5 or 7–O–glucoside, diosmin, naringenin 5–glucoside, eriodictyol 5–glucoside, quercetin 3–glucoside tested in our study showed concentration–dependent inhibitory activity against AGE formation for the first time. Although apigenin and diosmin had weak activities at the tested concentrations, the order of AGE inhibitory activity of most active flavonoids was similar, with IC<sub>50</sub> values ranging from 21.21 to 52.44  $\mu$  mol/L. Structural comparisons of these flavonoids and their inhibitory activities clearly show that the ortho-dihydroxy function of the B ring in the flavonoid skeleton, as evidenced by luteolin, quercetin, and their glycosides including rutin, quercetin 3-glucoside, and luteolin 5 or 7-O-glucoside, plays a pivotal role in the observed inhibitory activity. When such an O-dihydroxyl group was absent (i.e., apigenin and its glycosides), weak inhibitory activity was observed. The mechanism by which flavonoids inhibit AGE formation has yet to be established. The basic structure of flavonoids and other structural factors is important in the inhibitory mechanism. Muthenna et al. suggested the following two possible mechanisms for flavonoid-induced inhibition of AGE formation: (i) it scavenges free radicals directly or (ii) it chelates the metal ions by forming complexes with them and thereby partly inhibits post-Amadori formation<sup>[31]</sup>. It was also reported that protein glycation was accompanied by oxidative reactions<sup>[32]</sup>. Yim *et al.* has also suggested that glycation of albumin leads to the increased production of ROS<sup>[33]</sup>. Jung *et al.* demonstrated that the presence of an ortho-hydroxyl group on the B-ring plays an important role in the antioxidant activity in the total ROS system[17]. Similar trends were observed in flavonoid-induced inhibitory activity against AGE formation in the present study. Considering present and previous results on ROS inhibition of flavonoids<sup>[17,34]</sup>, it appears that the inhibitory mechanism of flavonoids against glycation is at least partly due to antioxidant activity. Flavonoids are of current interest in research due to the important biological and pharmacological properties attributed to their antioxidant properties<sup>[35]</sup>. Our results indicate several major flavonoids exhibit significant AGE inhibitory potential. According to previous research, two key flavonoids, luteolin and luteolin 5-O-glucoside inhibit RLAR activity and the generation of ROS[34]. Our previous comparative HPLC analysis of different kinds of thistles devoid of AGE inhibitory activity revealed the absence of these compounds<sup>[16]</sup>, indicating luteolin derivatives may play a role in the ability of C. maackii to inhibit diabetic complications.

Since a study comparing the ability of thistles to inhibit diabetic complications has not yet been reported, we studied the ability of some Korean thistles to inhibit AGE formation and isolated the active components from *C. maackii*, the most promising AGE inhibitory thistle found in the study. In conclusion, these previous data, together with our present results from AGE assays, indicate that *C. maackii* and *C. maackii* –derived flavonoids could be promising therapeutic agents for preventing diabetic complications. Based on the *in vitro* results, further experiments are needed to clarify the mechanisms of *C. maackii* and its flavonoids *in vivo*.

# Conflict of interest statement

We declare that we have no conflict of interest.

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