



Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Medicine

journal homepage: www.elsevier.com/locate/apjtm

Document heading doi:

Antibacterial activity of selected ethnomedicinal plants from South India

Rajendran Darling Anpin Raja¹, Solomon Jeeva², Juststella Wilfred Prakash¹, Johnson Marimuthu @ Antonisamy^{3*}, Varaprasadham Irudayaraj³¹Department of Botany, Scott Christian College (Autonomous), Nagercoil – 629 003, Tamil Nadu, India²Centre for Biodiversity and Biotechnology, Department of Botany, Nesamony Memorial Christian College, Marthandam – 629 165, Tamil Nadu, India³Department of Plant Biology and Plant Biotechnology, St. Xavier's College (Autonomous), Tirunelveli, Tamil Nadu, India

ARTICLE INFO

Article history:

Received 14 January 2011

Received in revised form 27 February 2011

Accepted 15 March 2011

Available online 20 May 2011

Keywords:

Antibacterial
Folk medicine
Kani tribe

ABSTRACT

Objective: To screen the antimicrobial potential of three ethnomedicinal plants *Chassalia curviflora* Thw. (*C. curviflora*), *Cyclea peltata* Hook. F. & Thomson (*C. peltata*) and *Euphorbia hirta* L. (*E. hirta*) used in folk medicines in Aarukani hills Kani tribe, Tamil Nadu, India against human bacterial pathogens. **Methods:** Antibacterial efficacy was performed by disc diffusion method against the pathogens viz., *Escherichia coli* (*E. coli*) (ATCC 35218), *Staphylococcus aureus* (*S. aureus*) (ATCC 6538), *Salmonella typhi* (*S. typhi*) (MTCC 733), *Proteus vulgaris* (*P. vulgaris*), *Proteus mirabilis* (*P. mirabilis*) and *Streptococcus pyogenes* (*S. pyogenes*) and incubated for 24 h at 37 °C. **Results:** The maximum degree of antibacterial activity was observed in *C. peltata* followed by *C. curviflora*. While *E. hirta* showed comparatively low degree of antibacterial activity. The methanolic extract of *C. peltata* showed the antibacterial activity against three pathogens viz., *S. pyogenes*, *P. vulgaris* and *E. coli* with the inhibition zones 12 mm, 10 mm and 9 mm, respectively. hexane extracts of *C. peltata* also showed the antibacterial activity against two selected pathogens viz., *P. vulgaris* and *P. mirabilis* with 15 mm and 12 mm of inhibition zones. All the three different concentrations (0.25, 0.50 & 0.75 mg/mL) of methanolic extract of *C. peltata* show the inhibitory effect on the three susceptible bacteria *S. pyogenes*, *P. vulgaris* and *E. coli* with the maximum inhibition in the highest concentration (0.75 mg/mL). The methanolic and hexane extracts of *C. curviflora* exhibited the antibacterial activity against only one bacterium each i.e. *P. vulgaris* and *S. typhi* with the maximum zone of inhibition 13 and 11 mm respectively. The methanolic and hexane extracts of *E. hirta* exhibited the antibacterial activity against only one bacterium i.e. *S. pyogenes* with the maximum zone of inhibition 13 and 11 mm respectively. **Conclusions:** The present investigation revealed that the *C. curviflora*, *C. peltata* and *E. hirta* are potentially good source of antibacterial agents and demonstrates the importance of such plants in traditional medicines.

1. Introduction

Since the beginning of civilization, people have used plants. Plants provide people with food, medicines, as well as materials for construction and the manufacture of crafts and tools and many other products like fuel, paints and poisons[1–3]. Nowadays the chemical and pharmacological constituents of medicinal plants used in different traditional

systems around the world are being increasingly explored for human benefit[4–8]. India with about 45 000 plant species and 550 tribal communities belonging to 227 ethnic groups[9] inhabited in varied geographic and climatic zones with diversified plant species and varied culture rich traditional knowledge system. Living close to the nature the tribal communities are custodian of unique traditional knowledge system and wisdom about ambient flora and fauna and rich heritage of ethnomedicine. Since most of these ethnic communities do not have their own scripts and written language, the information about prescriptions, pharmacology, attitude towards diseases and diagnosis of the age old tribal medicine are lying unclaimed. The people of the modern society are totally unaware of this rich traditional medicinal system. However, the studies in tribal

*Corresponding author: Johnson Marimuthu @ Antonisamy, Department of Plant Biology and Plant Biotechnology, St. Xavier's College (Autonomous), Tirunelveli, Tamil Nadu, India.

Tel: +91 97 86 92 43 34

Fax: + 91 46 22 56 17 65

E-mail: ptjohnson@gmail.com

medicine have enabled to identify 1 600 new drug yielding plants. So that the collection of information, documentation, and the scientific analysis of ethnomedicine is became important. Many efforts have been made to discover new antimicrobial compounds from various kinds of sources such as micro-organisms, animals, and plants. One of such resources is folk medicines. Systematic screening of them may result in the discovery of novel effective compounds^[10–13]. The Western Ghats of South India is one of the 29 biological hot spots identified in the world and gifted with large number of plants with exceptional medicinal properties^[14–17]. These plants are mainly utilized by the local ethnic people inhabiting in this region^[18,19]. One of the oldest tribes in the world, the Kani is the major tribal group settled here. They are a traditionally nomadic community, who now lead a primarily settled life in the forest. These people are known for their rich tradition in healing and other cultural activities.

The World Health Organization (WHO) estimated that 80% of the population of developing countries relies on traditional medicines, mostly plant drugs, for their primary health care needs^[20]. Also, modern pharmacopoeia still contains at least 25% drugs derived from plants and many others which are synthetic analogues built on prototype compounds isolated from plants^[21,22]. The primary benefits of using plant derived medicines are that they are relatively safer than synthetic alternatives, offering profound therapeutic benefits and more affordable treatment^[23]. For instance, the plants are generally readily available and their products are biodegradable. Antimicrobial drug resistance is a global problem today as the resistant microorganisms have emerged and spread throughout the world because of their genetic plasticity^[24,25]. Natural products of plants have been considered as the active ingredients of most of the modern medicines. The potential of higher plants as source for new drugs is still largely unexplored. Among the estimated 250 000 – 500 000 plant species, only a small percentage has been investigated phyto-chemically and the fraction submitted to biological or pharmacological screening is even smaller^[26,27]. Several works on plants are pointing out that, the plants utilized by different ethnic communities of the world have the capability to control the growth of various disease causing micro organisms. The increasing failure of chemotherapeutics and antibiotic resistance exhibited by pathogenic microbial infectious agents has led to the screening of several medicinal plants for their potential antimicrobial activity^[28,29]. It is well known that in drug discovery screening of ethnomedicinally important plants is more successful than the random screening. The Kani tribes in Tamil Nadu are using various plants to treat various diseases including pathogenic diseases. The Kani tribes in Aarukani Hills use the root and root bark of *Chassalia curviflora* Thw. (*C. curviflora*) to treat jaundice and wounds; tuber and leaves of *Cyclea peltata* Hook. f. & Thomson (*C. peltata*) to treat Chicken pox, diarrhoea, wounds, scabies and entire plant and latex of *Euphorbia hirta* L. (*E. hirta*) to treat wounds, skin diseases, asthma, and dysentery. Thus all the above three plants are being used by the Kani

tribes to cure mostly the pathogenic diseases. Hence, in the present investigation, the antimicrobial potential of these three plants (*C. curviflora*, *C. peltata* and *E. hirta*) have been evaluated against selected human bacterial pathogens.

2. Materials and methods

Healthy, disease free, plant parts *i.e.*, roots of *C. curviflora* (Wall.) Thw., var. *ophioxylodes* (Wall.) Deb. & B. Krishna tuber of *C. peltata* Hook. f. & Thomson and entire plants of *E. hirta* were collected from wild. The fresh materials were shade dried. Methanolic and hexane extracts were prepared from powdered materials and the extracts were used for antimicrobial studies. Antimicrobial study was carried out by disc diffusion method^[20] against the pathogens *viz.*, *Escherichia coli* (*E. coli*) (ATCC 35218), *Staphylococcus aureus* (*S. aureus*) (ATCC 6538), *Salmonella typhi* (*S. typhi*) (MTCC 733), *Proteus vulgaris* (*P. vulgaris*), *Proteus mirabilis* (*P. mirabilis*) and *Streptococcus pyogenes* (*S. pyogenes*).

3. Results

The methanolic and hexane extracts were tested for antibacterial activity against six human bacterial pathogens by using three differ concentrations *viz.*, 0.25, 0.50 and 0.75 mg/mL. The maximum degree of antibacterial activity was observed in *C. peltata* followed by *C. curviflora*. While, *E. hirta* showed comparatively low degree of antibacterial activity.

The methanolic extract of *C. peltata* showed the antibacterial activity against three pathogens *viz.*, *S. pyogenes*, *P. vulgaris* and *E. coli* with the inhibition zones 12 mm, 10 mm and 9 mm respectively (Table 1). Hexane extracts of *C. peltata* also showed the antibacterial activity against two selected pathogens *viz.*, *P. vulgaris* and *P. mirabilis* with 15 mm and 12 mm of inhibition zones. *E. coli*, *S. aureus*, *S. typhi* and *B. streptococci* are resistant to hexane extracts of *C. peltata* (Table 1). All the three different concentrations (0.25, 0.50, 0.75 mg/mL) of methanolic extract of *C. peltata* show the inhibitory effect on the three susceptible bacteria *S. pyogenes*, *P. vulgaris* and *E. coli* with the maximum inhibition in the highest concentration (0.75 mg/mL). In contrast, the different concentrations of hexane extract shows inhibitory effect on only two bacteria *P. vulgaris* and *P. mirabilis* with the maximum inhibition in the highest concentration (0.75 mg/mL).

The methanolic and hexane extracts of *C. curviflora* exhibited the antibacterial activity against only one bacterium each *i.e.* *P. vulgaris* and *S. typhi* with the maximum zone of inhibition 13 and 11 mm respectively (Table 1). The methanolic and hexane extracts of *E. hirta* exhibited the antibacterial activity against only one bacterium *i.e.* *S. pyogenes* (Table 1) with the maximum zone of inhibition 13 and 11 mm respectively (Table 1). *E. coli*, *S. aureus*, *S. typhi*, *P. vulgaris* and *P. mirabilis* are resistant to the methanolic and hexane extracts of *E. hirta* (Table 1).

Table 1Antibacterial activity of Methanolic and Hexane extracts of *C. peltata*, *C. curviflora* and *E. hirta*.

Solvents	Concentration (mg/mL)	Zone of inhibition (mm)																	
		<i>C. peltata</i>						<i>E. hirta</i>						<i>C. curviflora</i>					
		<i>Ec</i>	<i>Sa</i>	<i>St</i>	<i>Pv</i>	<i>Pm</i>	<i>Sp</i>	<i>Ec</i>	<i>Sa</i>	<i>St</i>	<i>Pv</i>	<i>Pm</i>	<i>Sp</i>	<i>Ec</i>	<i>Sa</i>	<i>St</i>	<i>Pv</i>	<i>Pm</i>	<i>Sp</i>
Methanol	0.25	4	–	–	3	–	5	–	–	–	–	–	4	–	–	–	6	–	–
	0.50	9	–	–	8	–	12	–	–	–	–	–	8	–	–	–	13	–	–
	0.75	10	–	–	9	–	12	–	–	–	–	–	13	–	–	–	13	–	–
Hexane	0.25	–	–	–	5	6	–	–	–	–	–	–	3	–	–	4	–	–	–
	0.50	–	–	–	11	14	–	–	–	–	–	–	6	–	–	7	–	–	–
	0.75	–	–	–	12	15	–	–	–	–	–	–	11	–	–	11	–	–	–

Ec – *E. coli*; *Sa* – *S. aureus*; *St* – *S. typhi*; *Pv* – *P. vulgaris*; *Pm* – *P. mirabilis*; *Sp* – *S. pyogenes*.

4. Discussion

In the present investigation, *in vitro* antibacterial efficacy of the crude extracts of three plants was quantitatively assessed on the basis of zone of inhibition. All the plants studied in the present investigation exhibited varying degree of inhibitory effect against the selected bacterial human pathogens. Eloff^[31] reported that methanol was the most effective solvent for plant extraction than hexane and water. In the present study we used methanol and hexane for extraction. The present study confirmed the Eloff observations with maximum activity.

The present study indicates anti bacterial property of the three plants against the selected strains of human pathogenic bacteria varies depends upon the solvent medium used for extraction. *C. peltata* shows highest activity (4/6) against the bacterial pathogens followed by *C. curviflora* (2/6) and *E. hirta* (1/6). Based on the previous literature on these three plants, there is no experimental study on *C. curviflora*. There are some studies on phytochemistry and pharmacology on *C. peltata*^[32, 33], but there is no report on antimicrobial activity. Ngemeny *et al*^[34–43] observed the antibacterial activity of *E. hirta* against *B. cereus*, *K. pneumoniae* and *P. aeruginosa*. Thus, the present study shows the presence of antibacterial activity in *C. curviflora* and *C. peltata* for the first time. In the case of *E. hirta*, in addition to the previous observation, the present study revealed and supplemented the antibacterial activity against the bacterial pathogen *S. pyogenes*.

The presence of antimicrobial activity in a particular part of a particular species may be due to the presence of one or more bioactive compounds such as alkaloids, glycosides, flavonoids, steroids, saponins *etc.*^[44]. Recently, a number of plants have been reported for antimicrobial properties across the world^[22,28,29]. In the present investigation, three ethnomedicinal plants from India have been screened for antimicrobial potential. As mentioned earlier, the roots and root bark of *C. curviflora* are used to treat jaundice and wounds. Among the two susceptible bacteria *P. vulgaris* is known to cause urinary tract infections and wound; *S. typhi* is known to cause fever and food borne illness. In the present study the methanolic extract and hexane extract of *C. curviflora* show the inhibitory activity against the above two bacteria and thus the present study confirms the traditional medical practice of the Kani tribe. Since the above two bacteria are susceptible to different extract (*P. vulgaris* for methanol extract and *S. typhi* for hexane extract), it is clear that the active compound may be of entirely different ones. Base on the present results, it is suggested that the same plant can also be used to treat

urinary infection, typhoid fever and food borne illness.

Present study on *C. peltata* revealed the high degree of antibacterial activity against four different bacteria namely *E. coli*, *P. vulgaris*, *P. mirabilis* and *S. pyogenes*. In general the above four bacteria are known to cause gastroenteritis, food borne illness, urinary tract infections, neonatal meningitis, nosocomial infections, wound, septicemia, pneumonias and from mild superficial skin infections to life-threatening systemic diseases. The Kani tribes, without having any scientific knowledge, for several decades they are successfully using the plant *C. peltata* to treat various pathogenic diseases such as diarrhea, wounds, scabies and chicken pox. The present experimental study confirms the traditional practice and supplement to treat other health problems such as urinary tract infections, neonatal meningitis, nosocomial infections, septicemia and pneumonias. In the meantime the roots of *C. peltata* were also found to contain high concentration of saponins^[45] which may be responsible for the antibacterial activity against various bacteria.

The results of the present study supplement the folkloric usage of the studied plants which possess several known and unknown bioactive compounds with antibacterial properties. By isolating and identifying these bioactive compounds new drugs can be formulated to treat various infectious diseases. Further phytochemical and pharmacological studies on the lesser known plants, *C. curviflora* and *C. peltata*, are necessary to utilize these ethnomedicinally important plants successfully. The presence of intraspecific variation in *C. curviflora* with the presence of three distinct varieties (var. *curviflora*, var. *ophioxylodes* and var. *longifolia*) shows the presence of more diversified chemicals with different bioactivities which are yet to be studied.

Conflict of interest statement

We declare that we have no conflict of interest.

References

- [1] Pugazharasi G, Meenakshi SA, Ramesh Kannan N, Bastin Churchill M, Natarajan E. Screening of antimicrobial activity of *Phyllanthus maderaspatensis* L. *J Basic Appl Bio* 2009; **3**(3&4): 43–49.
- [2] Sadheeshna Kumari S, Huxley AJ, Sasikala. *In vitro* propagation of medicinally important plant *Mimosa invisa*. *J Basic Appl Bio* 2009; **3**(3&4): 27–32.
- [3] Ponni V, Thenmozhi S, Rajan S. Screening of bioactive potentials

- and phytochemical nature of *Solanum trilobatum* extracts. *J Basic Appl Bio* 2009; **3**(3&4): 134–139.
- [4] Suresh Kumar P. Anti-fungal activity of *Leptadenia reticulata* in rat animal model *in vivo*. *J Basic Appl Bio* 2008; **2**(1): 9–13.
- [5] Hasan MF, Khan A, Rahman M, Sikdar B. Determination of antibacterial and antifungal activities of *Polygonum hydropepper* L. root extract. *J Basic Appl Bio* 2009; **3**(1&2): 6–10.
- [6] Jeeshna MV, Manorama S, Paulsamy S. Antimicrobial property of the medicinal shrub, *Glycosmis pentaphylla*. *J Basic Appl Bio* 2009; **3**(1&2): 25–27.
- [7] Suresh SN, Nagarajan N. Preliminary phytochemical and antimicrobial activity analysis of *Begonia malabarica* Lam. *J Basic Appl Bio* 2009; **3**(1&2): 59–61.
- [8] Rajan S, Jeevagangai TJ. Studies on the antibacterial activity of *Aegle marmelos*–fruit pulp and its preliminary phytochemistry. *J Basic Appl Bio* 2009; **3**(1&2): 76–81.
- [9] Ministry of Environment and Forest, Government of India. *State of forest report*. Dehradun: Ministry of Environment and Forest; 2003.
- [10] Tomoko N, Takashi A, Hiromu T, Yuka I, Hiroku M, Munekazu I, et al. Antibacterial activity of extracts prepared from tropical and subtropical plants on methicillin resistant *Staphylococcus aureus*. *J Health Sci* 2002; **48**: 273–276.
- [11] Jeeva S, Kingston C, Kiruba S, Kannan D. Sacred forests–treasure trove of medicinal plants: a case study from south Travancore. In: Trivedi PC. *Indigenous medicinal plants*. Jaipur: Pointer Publishers; 2007, p. 262–274.
- [12] Anpin Raja RD, Prakash JW, Jeeva S. Medicinal plants used by Kani in Aarukani hills of southern Western Ghats. In: Trivedi PC, Editor. *Indigenous medicinal plants*. Jaipur: Pointer Publishers; 2009, p. 1–48.
- [13] Anpin Raja RD, Prakash JW, Jeeva S. Antibacterial activity of some medicinal plants used by Kani tribe, southern Western Ghats, Tamilnadu, India. In: Trivedi PC. *Ethnic tribes and medicinal plants*. Jaipur: Pointer Publishers; 2010, p. 28–45.
- [14] Asbin Anderson A. Soil physicochemical properties of Adakadu Hills in Western Ghats of Kanyakumari district, Tamil Nadu. *J Basic Appl Bio* 2007; **1**(1): 14–17.
- [15] Anpin Raja RD, Prakash JW. Plants used as anti-venom by Kani tribes of Kilamalai reserver forest, Kanyakumari district. *J Basic Appl Bio* 2007; **1**(1): 27–32.
- [16] Meena R, Santhana Kumar G, Asir Selin Kumar R. Ethnomedicinal shrubs of Marunduvalmalai, Western Ghats, Tamilnadu, India. *J Basic Appl Bio* 2009; **3**(1&2): 67–70.
- [17] Mahesh M, Binisha GR, Brinitha BR, Vinaya VG, Jeeva S. Pteridophyte flora of Kanyakumari wildlife sanctuary. In: *National seminar on conservation and management of wetlands in an era of climate change*. Tamilnadu: Department of Botany; 2010.
- [18] Laila Banu NR, Sreeja S, Pinky VR, Prakash JW, Jeenu Jasmine A. Medicinal plants used by the rural people of Kattathurai, Kanyakumari district, Tamilnadu. *J Basic Appl Bio* 2007; **1**(1): 18–22.
- [19] Kingston C. Medicinal plants used in the endemic art of Travancore. *J Basic Appl Bio* 2007; **1**(1): 38–39.
- [20] Heinrich M, Gibbons S. Ethnopharmacology in drug discovery: an analysis of its role and potential contribution. *J Ethnopharm* 2001; **53**: 425–432.
- [21] Higa T, Anka J, Kitamura A, Koyama T, Akshashi M, Uchida T. Bioactive compounds from marine sponges. *Pure Appl Chem* 1994; **66**: 2227–2236.
- [22] Olowosulu AK, Ibrahim YKE. Studies on the antimicrobial screening of aqueous extracts of five plants used in folk medicine in Nigeria. West Africa. *J Biol Sci* 2006; **3**(5): 21–26.
- [23] Bandow JE, Brotz H, Leichert LIO, Labischinski H, Hecker M. Proteomic approach to understanding antibiotic action. *Antimicrob Agents Chemother* 2003; **47**: 948–955.
- [24] Kunin CM. Resistance to antimicrobial drugs – a worldwide calamity. *Ann Int Med* 1993; **118**: 557–561.
- [25] Abdullah WB, Anowa H, Doli G, Amina TS, Kamrun N, Korshed A, et al. Bacteremic typhoid fever in children in an urban slum, Bangladesh. *Emerg Infect Dis* 2005; **11**(2): 326–329.
- [26] Ahmad I, Beg AZ. Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multiple drug resistant human pathogens. *J Ethnopharma* 2001; **74**(2): 113–123.
- [27] Werner F, Paul O, Rainer A. Antibacterial activity of East African medicinal plants. *J Ethnopharma* 1999; **60**: 79–84.
- [28] Scazzocchio F, Cometa MF, Tomassini L, Palmery M. Antibacterial activity of *Hydrastis canadensis* extract and its major isolated alkaloids. *Planta Med* 2001; **67**: 561–564.
- [29] Bansal S, Malwal M, Sarin R. Anti-bacterial efficacy of some plants used in folkloric medicines in arid zone. *J Pharm Res* 2010; **3**(11): 2640–2642.
- [30] Gould JC, JH Bowie. The determination of bacterial sensitivity to antibiotics. *Edinburgh Med J* 1952; **59**: 178–199.
- [31] Eloff JN. Which extractant should be used for the screening and isolation of antimicrobial components from plants? *J Ethnopharmacol* 1998; **60**: 1–8.
- [32] Hullatti KK, Sharada MS. Comparative phytochemical investigation of the sources of Ayurvedic drug Patha: a chromatographic fingerprinting analysis. *Indian J Pharm Sci* 2010; **72**(1): 39–45.
- [33] Shine VJ, Latha PG, Shyamal S, Suja SR, Anuja GI, Sini S, et al. Gastric antisecretory and antiulcer activities of *Cyclea peltata* (Lam.) Hook. f. & Thoms. in rats. *J Ethnopharmacol* 2009; **125**(2): 350–355.
- [34] Ngemenya NM, Mbah AJ, Tane TP, Titanji PKV. Anti bacterial effects of some Cameroonian medicinal plants against common pathogenic bacteria. *Afr J T CAM* 2006; **3**(2): 84–93.
- [35] Manivannan K, Karthikai devi G, Anantharaman P, Balasubramanian T. Antimicrobial potential of selected brown seaweeds from Vedalai coastal waters, Gulf of Mannar. *Asian Pac J Trop Biomed* 2011; **1**(2): 114–120.
- [36] Mandal MD, Mandal S. Honey: its medicinal property and antibacterial activity. *Asian Pac J Trop Biomed* 2011; **1**(2): 154–160.
- [37] Habbal O, Hasson SS, El-Hag AH, Al-Mahrooqi Z, Al-Hashmi N, Al-Bimani Z, et al. Antibacterial activity of *Lawsonia inermis* Linn (Henna) against *Pseudomonas aeruginosa*. *Asian Pac J Trop Biomed* 2011; **1**(3): 173–176.
- [38] Tirupathi RG, Suresh BK, Kumar JU, Sujana P, Rao AV, Sreedhar AS. Anti-microbial principles of selected remedial plants from Southern India. *Asian Pac J Trop Biomed* 2011; **1**(4): 298–305.
- [39] Taye B, Giday M, Anmut A, Seid J. Antibacterial activities of selected medicinal plants in traditional treatment of human wounds in Ethiopia. *Asian Pac J Trop Biomed* 2011; **1**(5): 370–375.
- [40] Elumalai EK, Ramachandran M, Thirumalai T, Vinothkumar P. Antibacterial activity of various leaf extracts of *Merremia emarginata*. *Asian Pac J Trop Biomed* 2011; **1**(5): 406–408.
- [41] Kader G, Nikkon F, Rashid MA, Yeasmin T. Antimicrobial activities of the rhizome extract of *Zingiber zerumbet* Linn. *Asian Pac J Trop Biomed* 2011; **1**(5): 409–412.
- [42] Khan AV, Ahmed QU, Mir MR, Shukla I, Khan AA. Antibacterial efficacy of the seed extracts of *Melia azedarach* against some hospital isolated human pathogenic bacterial strains. *Asian Pac J Trop Biomed* 2011; **1**(6): 452–455.
- [43] Parekh J, Chanda SV. *In vitro* antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turk J Biol* 2007; **31**(1): 53–58.
- [44] Balandrin MJ, Klocke JA. *Medicinal, aromatic and industrial materials from plants*. Berlin: Springer-Verlag; 1988, p. 1–36.
- [45] Kirana H, Srinivasan BP. Effect of *Cyclea peltata* Lam. Roots aqueous extract on glucose levels, lipid profile, insulin, TNF- α and skeletal muscle glycogen in type 2 diabetic rats. *Indian J Exp Biol* 2010; **48**: 499–502.