

Contents lists available at ScienceDirect

## Asian Pacific Journal of Tropical Biomedicine

journal homepage:www.elsevier.com/locate/apjtb



Document heading doi:10.1016/S2221-1691(11)60099-3 © 2011 by the Asian Pacific Journal of Tropical Biomedicine. All rights reserved.

# Antibacterial efficacy of the seed extracts of *Melia azedarach* against some hospital isolated human pathogenic bacterial strains

Abdul Viqar Khan<sup>1\*</sup>, Qamar Uddin Ahmed<sup>2</sup>, M Ramzan Mir<sup>3</sup>, Indu Shukla<sup>3</sup>, Athar Ali Khan<sup>1</sup>

<sup>1</sup>Department of Botany, Faculty of Life Sciences, Aligarh Muslim University, Aligarh–202002, State of Utter Pradesh, India <sup>2</sup>Department of Pharmaceutical Chemistry, Faculty of Pharmacy, International Islamic University Malaysia, 25200–Kuantan, Pahang Darul Makmur, Malaysia <sup>3</sup>Department of Marshielen, Faculty of Life Sciences, Aligarh Muslim, University, Aligarh, 202002, State of Utter Products, India

<sup>3</sup>Department of Microbiology, Faculty of Life Sciences, Aligarh Muslim University, Aligarh–202002, State of Utter Pradesh, India

## ARTICLE INFO

Article history: Received 5 April 2011 Received in revised form 21 April 2011 Accepted 11 May 2011 Available online 20 May 2011

*Keywords: Melia azedarach* Antibacterial efficacy Pathogenic bacteria

#### 1. Introduction

Medicinal plants are part and parcel of human society to combat diseases from the dawn of civilization. In developing countries like India, different plant species are explored by ethnic societies, exploiting them for treatment of various diseases and disorders. Infectious diseases have continued to ravage most developing nations of the world. Approximately half of all deaths caused by infectious diseases each year can be attributed to just three diseases: tuberculosis, malaria, and AIDS. Altogether, these diseases cause over 300 million illnesses and more than 5 million deaths each year<sup>[1]</sup>. Coupled with the scourge of infectious diseases is the recent emergence of drug-resistant microorganisms that have reduced the effectiveness of antimicrobial agents<sup>[2,3]</sup>. According to the World Health Organization (WHO) in 2008, more than 80% of the world's population relied on traditional medicine for their primary healthcare needs. Bioactive compounds with antibacterial effects need to be explored from local natural resources. Being new, such compounds may not have the problem of microbial resistance, and with some structural modification their activity could be diversified. Seeking remedies for

## ABSTRACT

**Objective:** To investigate the antibacterial potential of the polar and non-polar extracts of the seeds of *Melia azedarach (M. azedarach)* L. (Meliaceae) against eighteen hospital isolated human pathogenic bacterial strains. **Methods:** Petrol, benzene, ethyl acetate, methanol, and aqueous extracts at five different concentrations (1, 2, 5, 10 and 15 mg/mL) were evaluated. Disk diffusion method was followed to evaluate the antibacterial efficacy. **Results:** All extracts of the seeds demonstrated significant antibacterial activity against tested pathogens. Among all extracts, ethyl acetate extract revealed the highest inhibition comparatively. The present study also favored the traditional uses reported earlier. **Conclusions:** Results of this study strongly confirm that the seed extracts of *M. azedarach* could be effective antibiotics, both in controlling gram-positive and gram-negative human pathogenic infections.

human ailments from the environment has formed the basis for therapeutics. Various plants are used traditionally by ethnic societies from remote past in the forms of extracts of different plant parts (*e.g.* fruits, leaves, stem–bark and roots) in hot water or alcohol. They can be used in the form of infusions, decoctions or concoctions<sup>[4,5]</sup>.

Melia azedarach (M. azedarach) L. (Meliaceae) (local name: Bakain) is one of the most useful traditional medicinal plant like Azadirachta indica (A. indica) A. juss (local names: Neem, Margose) from the family Meliaceae in India. Each part of M. azedarach has some medicinal properties like A. indica and thus is commercially exploitable. During the last twenty years, apart from the chemistry of this plant, considerable progress has been achieved regarding the biological activity and medicinal applications.

*M. azedarach* L. is native to tropical Asia. It is wide spread and naturalized in most of the tropics and subtropical countries. It was introduced and naturalized in Philippines, United States of America, Brazil, Argentine, many African and Arab countries<sup>[6]</sup>.

Leaves are used in leprosy, scrofula, anthelmintic, antilithic, diuretic, deobstruent, and resolvent. Roots are effectively used as resolvent and deobstruent. Seed oil is the most active medicinal product of the plant and used as an antiseptic for sores and ulcers that show no tendency to heal. It is also used for rheumatism and skin diseases such as ringworm and scabies. Internally, the oil is useful in

<sup>\*</sup>Corresponding author: Dr. Abdul Viqar Khan, Department of Botany, Faculty of Life Sciences, Aligarh Muslim University, Aligarh–202002, State of Utter Pradesh, India. E-mail: viqarvicky@gmail.com

Foundation Project: Supported by SERC Fast Track OYS Schem, Department of Science and Technology (DST), New Delhi, India [No. 304/2004/SR/FT/L-129/2004)].

malaria fever and leprosy. There are many traditional uses in Northern India. Fresh leaf extract is applied externally for burns. Fresh leaf extract is used as mouth wash for gingivitis (Inflamed bleeding gums). Stem bark infusion 30–50 mL is administered orally twice a day for onorrhea. Leaf extract 5 mL is administered orally thrice a day for piles. Leaf extract 5–10 mL is administered orally twice a day for 7 days for pyrexia<sup>[7]</sup>.

Chemical constituents of the seeds including  $\beta$ -sitosterol, vanillin, benzoic acid, vanillic acid, daucosterol,  $\alpha$  –D–glucopyranose, limonoid glycoside *viz* 6,11–diacetoxy–7–oxo–14beta, 15beta–epoxymeliacin (1,5–diene–3–O–beta–D–glucopyranoside) and scopoletin, a hydroxyl coumaramin, melianol meliacin, meliacarpin, meliartenin vanillin, hydroxyl–3–methoxcinnamaldehyde and (+–) pinoresinol have been reported earlier from the seeds of *M. azedarach*[8,9].

Seeds of *M. azedarach* L. have been scientifically reported to exert antimalarial, antifungal<sup>[9,10]</sup>, ovicidal<sup>[11]</sup>, insecticidal<sup>[12]</sup>, antifeedant<sup>[13]</sup>, rodenticidal activities<sup>[14]</sup> in *vitro* and *in vivo* studies.

In view of the ethnobotanical uses and medicinal properties of *M. azedarach*, it is conjectured that this plant might possess antibacterial properties. Therefore, the present study was aimed to evaluate the antibacterial activity of seed extracts of *M. azedarach* against some hospital isolated pathogenic bacterial strains.

#### 2. Materials and methods

#### 2.1. Plant material

Matured seeds of *M. azedarach* L. were collected from different localities of Aligarh district, India. Voucher specimen number [AV24, AV206] of the plant was deposited in the Herbarium of Department of Botany, Faculty of Life Sciences, Aligarh Muslim University, Aligarh–202002, U.P., India.

#### 2.2. Preparation of extracts

Freshly dried and healthy plant material was pulverized into fine powder using an electric grinder. Powder was stored in a desiccator. 500 g plant powder was refluxed with 95% methyl alcohol (MeOH) in a round bottom flask on a hot water bath for 10 h. Mother liquor (crude MeOH extract) was filtered out and residual plant material was again refluxed with 95% MeOH for 10 h. The entire process was repeated four times to obtain maximum yield of MeOH extract. The extract was evaporated to dryness at 35 °C under reduced pressure. Dried methanol extract was refluxed with light petrol [(60−80) °C] for 5 h. After filtration, the residual methanol extract was again refluxed with petrol for 5 h and filtered. This process was repeated five times. Petrol was evaporated under reduced pressure to obtain petrol soluble extract. Petrol insoluble fraction of methanol extract was refluxed with benzene for 5 h. Thereafter, it was filtered and refluxed again with benzene for 5 h and filtered. The

process was repeated five times. Benzene was evaporated under reduced pressure to obtain benzene soluble extract. Benzene insoluble fraction was refluxed with ethyl acetate for 5 h. Thereafter, it was filtered and refluxed again with ethyl acetate for 5 h and filtered. The process was repeated five times. Ethyl acetate was evaporated under reduced pressure to obtain ethyl acetate soluble extract. Ethyl acetate insoluble fraction was refluxed with methyl alcohol (95%) for 5 h, filtered and was repeatedly refluxed for five times with methanol. The methanol soluble fraction was evaporated under reduced pressure to obtain methanol extract, while methanol insoluble residue was discarded.

## 2.3. Preparation of aqueous extract of the seeds

Shade dried plant material (500 g) was ground to a fine powder then it was poured with double distilled water, and left for 72 h at room temperature. The flask was then refluxed over hot water bath for 10 h and the mother liquor was filtered. The distilled water was again refluxed and filtered. This process was repeated for four times. The filtrate was evaporated to complete dryness under reduced pressure on a water bath. Thus the residue was aqueous plant extract.

#### 2.4. Microorganisms

The bacteria were obtained from the bacterial stock, Department of Microbiology, Jawaharlal Nehru Medical College, Aligarh, India. The bacterial cultures were maintained at 4  $^{\circ}$  on nutrient agar.

## 2.5. Antibacterial activity

The agar disk diffusion method was used to determine the susceptibility of plant extracts against chosen bacteria<sup>[15,16]</sup>. Standardized inoculums  $(1-2 \times 10^7 \text{ CFU/mL } 0.5 \text{ McFarland}$  standard) were introduced on the surface of the plates containing Mueller Hinton agar (MHA), which was spread evenly with a glass spreader. The paper disk (6 mm in diameter; Whatman filter paper no 1) containing 1, 2, 5, 10 and 15 mg/mL plant extracts was dried and placed on the agar surface with the help of a sterile forcep. Finally, paper discs were placed with forceps to make complete contact with the surface of the medium. Plates were allowed to stand at room temperature for 30 min (Pre diffusion time) and then incubated aerobically at 37  $^{\circ}$  and examined for the zone of inhibitions after 24 h. The experiments were repeated thrice.

## 3. Results

A result of the *in vitro* antibacterial activity of *M*. *azedarach* seed extracts was described in Table 1. The petrol extract of the seeds was found to be effective against six gram-positive and nine gram-negative pathogenic bacteria. Benzene extract inhibited the growth of three gram-positive and nine gram-negative pathogens. Ethyl acetate, methanol and aqueous extracts displayed inhibitory activity against all the tested pathogens.

## Table 1

Inhibition zone of the seed extracts of *M. azedarach* L. against some hospital isolated human pathogenic bacterial strain (mm).

Extract (mg/mL)	Gram positive bacteria							Gram negative bacteria										
	1	2	3	4	5	6	7	1	2	3	4	5	6	7	8	9	10	11
Petrol extract (1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Petrol extract (2)	8	-	-	10	8	-	10	-	-	-	10	-	-	7	7	-	10	-
Petrol extract (5)	10	7	7	16	11	-	13	7	12	-	14	9	11	11	11	-	14	11
Petrol extract (10)	16	11	12	17	16	-	19	9	17	-	17	11	12	15	15	-	19	14
Petrol extract (15)	21	18	17	22	18	-	23	13	22	-	21	13	17	19	20	-	22	17
Benzene extract (1)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Benzene extract (2)	8	-	-	-	8	-	7	-	-	-	10	-	-	7	7	-	10	11
Benzene extract (5)	10	_	_	_	11	_	10	11	11	_	14	9	11	11	11	-	14	14
Benzene extract (10)	16	_	_	_	16	_	15	14	16	_	17	11	12	15	15	-	19	18
Benzene extract (15)	21	-	-	-	18	_	19	19	21	-	21	13	17	19	20	-	22	21
Ethyl acetate extract (1)	9	-	-	-	-	7	9	-	7	-	7	7	9	-	_	7	9	7
Ethyl acetate extract (2)	12	7	11	10	9	10	12	10	11	11	12	12	12	12	9	10	13	11
Ethyl acetate extract (5)	19	11	17	16	16	15	17	13	17	16	17	17	18	17	16	17	18	19
Ethyl acetate extract (10)	22	16	19	19	20	18	20	17	19	18	22	22	23	19	20	22	22	23
Ethyl acetate extract (15)	26	21	23	23	26	23	26	20	23	23	25	27	26	24	24	27	26	27
Methanol extract (1)	-	-	-	-	-	-	_	-	-	-	-	-	-	-	-	7	-	7
Methanol extract (2)	7	-	-	10	9	9	7	7	-	-	9	10	7	7	8	10	7	11
Methanol extract (5)	12	-	11	16	16	15	12	9	8	9	14	15	10	12	12	16	12	16
Methanol extract (10)	16	10	15	19	20	19	17	13	13	14	18	19	13	16	18	20	18	19
Methanol extract (15)	21	16	20	23	26	21	22	17	17	18	22	23	19	21	21	23	22	21
Aqueous extract (1)	-	-	-	-	-	-	_	-	-	-	-	-	-	-	-	-	-	-
Aqueous extract (2)	7	-	7	10	9	-	7	-	9	-	-	-	-	-	-	-	7	7
Aqueous extract (5)	10	-	11	12	13	9	10	9	11	8	9	11	8	7	11	10	12	11
Aqueous extract (10)	15	8	15	16	18	11	15	11	14	12	14	15	11	12	14	12	15	16
Aqueous extract (15)	18	11	19	19	20	16	19	13	16	13	18	19	15	17	19	16	20	19
Chloramphenicol (30 mg/disk)	18	18	16	-	-	-	16	18	16	-	16	18	-	16	17	19	18	20

Gram positive bacteria: 1. Staphylococcus aureus; 2. Staphylococcus aureus ATCC 25923; 3. Staphylococcus epidermidis; 4. Group–A Streptococcus; 5. Group–B Streptococcus; 6. Enterococcus faecalis; 7. Bacillus subtilis.

Gram negative bacteria: 1. Escherichia coli; 2. Edwardsiella tarda; 3. Klebsiella pneumoniae; 4. Proteus mirabilis; 5. Proteus vulgaris;

6. Pseudomonas aeruginosa; 7. Salmonella typhi; 8. Shigella boydii; 9. Shigella dysenteriae; 10. Shigella flexneri; 11. Plesiomonas shigelloides. -: no inhibition.

## 4. Discussion

In the present era, plant and herb resources are abundant, but these resources are dwindling fast due to the onward march of civilization<sup>[17]</sup>. Although a significant number of studies have been used to obtain purified phytochemicals, a very few screening programmes have been initiated on crude plant materials. It has also been widely observed and accepted that the medicinal value of plants lies in the bioactive phytocomponents present in the plants<sup>[18-36]</sup>. The results of the present study are encouraging as all the tested extracts revealed antibacterial potential, although the inhibitory activity was strain specific and concentration dependent. Petrol fraction showed maximum inhibition against Bacillus subtilis, Proteus mirabilis and Shigella flexneri. Benzene extract inhibited the growth of twelve tested pathogens and the maximum inhibition zone was recorded against Proteus mirabilis and Shigella flexneri. All the tested pathogens showed sensitivity against the ethyl acetate extract and the most affected bacteria were Staphylococcus aureus, Bacillus subtilis (gram positive) and Pseudomonas aeruginosa and Shigella flexneri (gram negative). Methanol extract was also found to be effective against all the strains and maximum inhibition was recorded against Shigella dysenteriae and Plesiomonas shigelloides.

Aqueous extract showed moderate degree of sensitivity against all tested pathogenic bacteria.

From the results it is clear that seeds extracts of *M*. *azedarach* L. are effective in controlling infections caused by both gram positive and gram negative strains. During these investigations it became clear that the most effective crude extract was ethyl acetate, which demonstrated maximum inhibition followed by methanol and aqueous extracts which inhibited the growth of all the tested human pathogens. The petrol and benzene extracts, as compared with the polar extracts, showed antimicrobial action against fifteen and twelve pathogens, respectively. The above results show that seed extracts can be effective antibiotics, both in controlling Gram positive and Gram negative human pathogens.

## **Conflict of interest statement**

We declare that we have no conflict of interest.

#### Acknowledgements

Thanks are due to Department of Science and Technology SERC Division, New Delhi for financial support to the author

## Dr. Abdul Viqar Khan.

#### References

- World Health Organization (WHO). The World Health Report (Changing history, statistical annex, death by cause, sex and mortality stratum in WHO regions, estimates for 2002). Geneva: WHO; 2004, p. 120–121.
- [2] Goossens H, Ferech M, Vander Stichele R, Elseviers M. Outpatient antibiotic use in Europe and association with resistance: a crossnational database study. *Lancet* 2005; 365: 579–587.
- [3] Mathew AG, Cissell R, Liamthong S. Antibiotic resistance in bacteria associated with food animals: a United States perspective of livestock production. *Foodborne Pathog Dis* 2007; 4: 115–133.
- [4] Meena M. Enhancement of bioactive compound production, antimicrobial activity and evaluation in animal models. J Med Plant Res 2009; 3(7): 495-497.
- [5] Cox PA. The ethnobotanical approach to drug discovery: strengths and limitations. England: John Wiley & Sons; 1994, p. 25–36.
- [6] Khan AV, Khan AA, Shukla I. In vitro antibacterial potential of Melia azedarach crude leaf extracts against some human pathogenic bacterial strains. Ethnobot Leafl 2008; 12: 39–45.
- [7] Khan AV. Ethnobotanical studies on plants with medicinal and anti-bacterial properties. (PhD Thesis). Aligarh Muslim University, Aligarh; 2002, p. 1–293.
- [8] Chong XT, Tian GZ, Cheng ZL, Yao QQ. Study on chemical constituents of the seed of *Melia azedarach L. Food and Drug* 2009; 11: 30–31.
- [9] Carpinella MC, Ferrayolic CG, Palacios SM. Antifungal synergistic effect of scopoletin, a hydroxycoumarin isolated from *Melia* azedarach L. fruits. J Agric Food Chem 2005; 53: 2922–2927.
- [10] Nathan SS, Savitha G, George DK, Marmadha A, Suganya L, Chung PG. Efficacy of *Melia azedarach* L. extract on the malarial vector *Anopheles stepheni* Listone (Dipetra:Culicidae). *Bioresour Technol* 2006; 97: 1316–1323.
- [11] Maciel MV, Morais SM, Bevilaqua CML, Camurca–Vasconcelos ALF, Costa CTC, Castro CMS. Ovicidal and larvicidal acivity of *Melia azedarach* extracts on *Haemonchus contortus*. *Vet Parasitol* 2006; 140: 98–104.
- [12] Akhtar Y, Young YR, Isman MB. Comparative bioactivity of selected extracts from Meliaceae and some commercial botanical insecticides against two noctuid caterpillars, *Trichoplusia ni* and *Pseudoletia unipunctat*. *Phytochem Rev* 2008; **7**: 77–88.
- [13] Charleston DS, Kfir R, Vet LE, Dicke M. Behavior responses of diamond back moth *Plutella xylostella* (Lepidoptera: Plutellidae) to extracts derived from *Melia azedarach* and *Azadirachta indica*. *Bull Entomol Res* 2005; **95**: 457–465.
- [14] Roop JK, Dhaliwal PK, Guraya SS. Extracts of Azadirachta indica and Melia azedarach seeds inhibit folliculogenesis in albino rats. Braz J Med Biol Res 2005; 38: 943–947.
- [15] National Committee for Clinical Laboratory Standards (NCCLS). Performance standards for antimicrobial disk susceptibility testing. Fourteenth informational supplement. Wayne, Pennsylvania: NCCLS; 2004.
- [16] National Committee for Clinical Laboratory Standards (NCCLS). Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard-sixth edition. Wayne, Pennsylvania: NCCLS; 2003.
- [17] Vogel HG. Similarities between various systems of traditional medicine. Considerations for the future of ethnopharmacology. J Ethnopharmacol 1991; 35: 179–190.
- [18] Veeramuthu D, Muniappan A, Savarimuthu I. Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from

Tamil Nadu, India. BMC Complement Altern Med 2006; 6: 35.

- [19] Abubakar S, Ahmed QU, Othman AS, Omar MN. Bacteriostatic and bactericidal activity of the polar and non-polar extracts of Andrographis paniculata against skin disease causing pathogenic bacteria. J Med Plant Res 2011; 5: 7–14.
- [20] Khan AV, Ahmad R, Khan AA, Shukla I. Antibacterial activity of Oxystelma esculentum leaf extracts against some hospital isolated human pathogenic bacterial strains. J Herbal Med Toxicol 2008; 2: 67–70.
- [21] Khan AV, Ahmad QU, Shukla I, Khan AA. Antibacterial efficacy of Bacopa Monnieri leaf extracts against pathogenic bacteria. *Asian Biomed* 2010; 4: 651–655.
- [22] Khan AV, Khan AA. Ethnobotany of Eclipta prostrate. Indian J Tradit Knowl 2008; 2: 316–320.
- [23] Madhumitha G, Saral AM. Preliminary phytochemical analysis, antibacterial, antifungal and anticandidal activities of successive extracts of *Crossandra infundibuliformis*. Asian Pac J Trop Med 2011; 4(3): 192–195.
- [24] Johnson M, Wesely EG, Kavitha MS, Uma V. Antibacterial activity of leaves and inter-nodal callus extracts of *Mentha arvensis* L. *Asian Pac J Trop Med* 2011; 4(3): 196–200.
- [25] Peixoto JRO, Silva GC, Costa RA, Fontenelle JLS, Vieira GHF, Filho AAF, et al. *In vitro* antibacterial effect of aqueous and ethanolic Moringa leaf extracts. *Asian Pac J Trop Med* 2011; 4(3): 201–204.
- [26] Chatterjee SK, Bhattacharjee I, Chandra G. Isolation and identification of bioactive antibacterial components in leaf extracts of *Vangueria spinosa* (Rubiaceae). Asian Pac J Trop Med 2011; 4(1): 35-40.
- [27] Mandal S, DebMandal M, Pal NK, Saha K. Antibacterial activity of honey against clinical isolates of *Escherichia coli*, *Pseudomonas* aeruginosa and Salmonella enterica serovar Typhi. Asian Pac J Trop Med 2010; 3(12): 961–964.
- [28] Kannan RRR, Arumugam R, Anantharaman P. Antibacterial potential of three seagrasses against human pathogens. Asian Pac J Trop Med 2010; 3(12): 890–893.
- [29] Johnson M, Wesely EG, Zahir Hussain MI, Selvan N. In vivo and in vitro phytochemical and antibacterial efficacy of Baliospermum montanum(Willd.) Muell. Arg. Asian Pac J Trop Med 2010; 3(12): 894–897.
- [30] Merin DD, Prakash S, Bhimba BV. Antibacterial screening of silver nanoparticles synthesized by marine micro algae. Asian Pac J Trop Med 2010; 3(10): 797–799.
- [31] Kaur J, Rathinam X, Kasi M, Leng KM, Ayyalu R, Kathiresan S, et al. Preliminary investigation on the antibacterial activity of mango (*Mangifera indica* L: Anacardiaceae) seed kernel. Asian Pac J Trop Med 2010; 3(9): 707–710.
- [32] Naik MI, Fomda BA, Jaykumar E, Bhat JA. Antibacterial activity of lemongrass (*Cymbopogon citratus*) oil against some selected pathogenic bacterias. *Asian Pac J Trop Med* 2010; 3(9): 535-538.
- [33] Bhimba BV, Meenupriya J, Joel EL, Naveena DE, Kumar S, Thangaraj M. Antibacterial activity and characterization of secondary metabolites isolated from mangrove plant Avicennia officinalis. Asian Pac J Trop Med 2010; 3(7): 544-546.
- [34] Bhattacharjee I, Chatterjee SK, Chandra G. Isolation and identification of antibacterial components in seed extracts of Argemone mexicana L. (Papaveraceae). Asian Pac J Trop Med 2010; 3(7): 547-551.
- [35] Ghosh A, Das BK, Roy A, Mandal B, Chandra G. Antibacterial activity of some medicinal plants. J Nat Med 2008; 62: 259-262.
- [36] Helmy WA, Amer H, El-Shayeb NMA. Biological and antimicrobial activities of aqueous extracts from neem tree (Azadirachta indica A. Juss., Meliaceae). J Appl Sci Res 2007; 3(10): 1050-1055.