



Contents lists available at ScienceDirect

Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.elsevier.com/locate/apjtb



Document heading doi:10.1016/S2221-1691(12)60220-2 © 2012 by the Asian Pacific Journal of Tropical Biomedicine. All rights reserved.

Antibacterial activity of the essential oils from the leaves of *Eucalyptus globulus* against *Escherichia coli* and *Staphylococcus aureus*

Bachir Raho G^{1*}, Benali M²¹Science Faculty Mustapha Stambouli University of Mascara – Algeria²Science Faculty, Djillali Liabès University of Sidi Bel Abbès – Algeria

ARTICLE INFO

Article history:

Received 22 February 2012

Received in revised form 23 March 2012

Accepted 28 April 2012

Available online 28 September 2012

Keywords:

Antimicrobial activity

Essential oil

*Eucalyptus globulus**Escherichia coli**Staphylococcus aureus*

ABSTRACT

Objective: To examine the *in vitro* antimicrobial activities of essential oil of the leaves of *Eucalyptus globulus* (*E. globulus*). **Methods:** The essential oils of this plant were obtained by the hydrodistillation method. The inhibitory effects of this essential oil were tested against *Escherichia coli* (*E. coli*) and *Staphylococcus aureus* (*S. aureus*) by using agar disc diffusion and dilution broth methods. **Results:** The results obtained showed that essential oil of the leaves of *E. globulus* has antimicrobial activity against gram negative bacteria (*E. coli*) as well as gram positive bacteria (*S. aureus*). **Conclusion:** The encouraging results indicate the essential oil of *E. globulus* leaves might be exploited as natural antibiotic for the treatment of several infectious diseases caused by these two germs, and could be useful in understanding the relations between traditional cures and current medicines.

1. Introduction

The Myrtaceae family includes 140 genera and about 3800 species distributed in tropical and subtropical regions of the world[1]. *Eucalyptus* is one of the world's important and most widely planted genera[2]. It is a tall, evergreen tree, native to Australia and Tasmania, successfully introduced worldwide, now extensively planted in many other countries[3]. It was introduced in Algeria in 1854 by Ramel[4]. *Eucalyptus* species are well known as medicinal plants because of their biological and pharmacological properties. In the international pharmacopeia, the most important and represented species, however, is *Eucalyptus globulus* (*E. globulus*) which is the main furnisher of essential oils[5]. These essential oils are in great demand in the market[5], since they find applications as anesthetic, anodyne, antiseptic, astringent, deodorant, diaphoretic, disinfectant, expectorant, febrifuge, fumigant, hemostat, inhalant, insect repellent, preventive, rubefacient, sedative yet stimulant, vermifuge, for a folk remedy for abscess, arthritis, asthma, boils, bronchitis, burns, cancer, diabetes,

diarrhea, diphtheria, dysentery, encephalitis, enteritis, erysipelas, fever, flu, inflammation, laryngalgia, laryngitis, leprosy, malaria, mastitis, miasma, pharyngitis, phthisis, rhinitis, sores, sore throat, spasms, trachalgia, worms, and wounds[6]. Sometimes their demand is also high in the soap and cosmetic industries[5].

The spread of drug resistant microbial pathogens is one of the most serious threats to successful treatment of infectious diseases[7].

Escherichia coli (*E. coli*) and *Staphylococcus aureus* (*S. aureus*) are two opportunistic pathogens that cause severe and life-threatening infections in immunocompromised patients[8]. The Gram-positive bacterium *S. aureus* is mainly responsible for post operative wound infection, toxic shock syndrome and food poisoning. The gram-negative bacterium *E. coli* is present in human intestine and causes lower urinary tract infection, coleocystis or septicemia[9]. Several studies have documented increasing resistance rates in *S. aureus* and *E. coli* to antibiotics[3,10–13].

The main objective of this study is to examine the antimicrobial activities of the Water-distilled extracts of *E. globulus* leaves on two clinically significant microorganisms, *E. coli* and *S. aureus* by means of the agar diffusion test and dilution broth method.

*Corresponding author: Science Faculty Mustapha Stambouli University of Mascara – Algeria
Tel:00213551774584.
E-mail:bachir_rahog@yahoo.fr.

2. Materials and methods

2.1. Plant material and essential oil extraction

Fresh plant leaves of *E. globulus* were collected during the flowering stage from the Djillali liabes University campus in Sidi Bel Abbes City, North West of Algeria. The plant leaves were identified by environmental sciences department, then were taken to the Biotoxicology Laboratory of Djillali liabes University for the extraction.

The fresh leaves were subjected to steam distillation using a Clevenger-type apparatus. Briefly, the plant leaves were completely immersed in water and heated to boiling, after which the essential oil was evaporated together with water vapour and finally collected after decantation. The distillate was isolated and dried in a Rota-vapor to giving greenish-yellow oil. The oil was stored at 4 °C until the antimicrobial screening^[14,15]. The extraction yield of this essential oil was 1.2 % (w/w).

2.2. Bacterial strains

The essential oil was tested against two clinical isolated strains provided by the Laboratory of Medical Analysis, located in Dr. Hassani Abdelkader Hospital University Center (CHU) of Sidi Bel Abbes City, situated in the North West of Algeria for patients suffering from certain infectious diseases. The identity of the microorganisms used in this study (*E. coli* and *S. aureus*) was confirmed by standard biochemical tests and morphological studies^[16,17].

2.3. Antimicrobial screening

Three methods were used to determine the antibacterial activity; agar disc diffusion method and dilution broth method.

The agar disc diffusion method was employed to determine the antimicrobial activities of the essential oil. Disc-assay was found to be a simple, cheap and reproducible practical method^[18]. A suspension of each sample tested micro-organism diluted prior to 10⁻¹, 10⁻² and 10⁻³ (1 mL of 10⁸ cells /mL) was spread on a solid agar medium in Petri dishes (Mueller-Hinton agar). Filter paper discs (6 mm in diameter) were soaked in 13 µL of the essential oil and placed on the inoculated plates and allowed to dry for 15 min, then incubated at 37 °C for 24 h. The diameters of the inhibition zones were measured in millimeters^[19].

Dilution broth susceptibility assay^[20] was used for the antimicrobial evaluation. Stock solutions of the essential oils were prepared in ethanol by mixing 1 ml of the extracts with 9 ml of alcohol in test tubes to obtain the mother solution, followed by successive dilutions at 10⁻² and 10⁻³. The control was prepared by mixing 1 mL of distilled water with 9 mL of alcohol solution. 1 mL of each dilution and 0.5 mL of tested culture strains are added to 8 mL of a nutrient broth, maintained after in a Marie bath at 37 °C under stirring for 24 h, then seeded by streaking the surface of the agar medium and incubated at 37 °C for 24 h.

3. Results

Figure 1 and 2 summarizes the microbial growth inhibition by the essential oil of *E. globulus*, which showed good antibacterial activities against the two tested organisms.

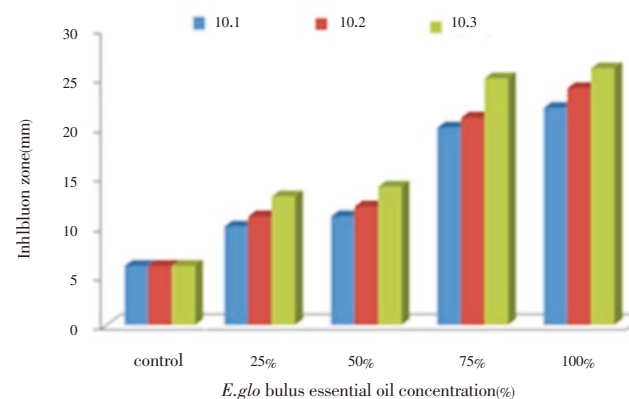


Figure 1. Antimicrobial activity evaluation of the essential oil *E. globulus* leaves against *E. coli*, using the agar disc diffusion method.

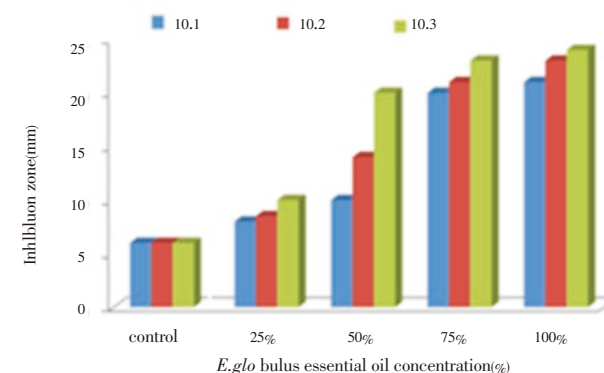


Figure 2. Antimicrobial activity evaluation of the essential oil *E. globulus* leaves against *S. aureus*, using the agar disc diffusion method.

Table 1

Antimicrobial activity evaluation of the essential oil of *E. globulus* leaves using the dilution broth method against the two bacterial strains.

Microbial strains	Essential oil dilution				Control
	10 ⁻³ , 10 ⁻⁴	10 ⁻⁵	10 ⁻¹	10 ⁻²	
<i>E. coli</i>	+	+	++	+++	+++
<i>S. aureus</i>	+	++	++	+++	+++

++: Comparable growth with that Control +; slow growth

The results revealed that the essential oil showed antibacterial activity with varying magnitudes, depending on the size of inoculums and the concentration of essential oil. Diameter of inhibition zone of essential oil of *E. globulus* leaves varied from 8 to 26 mm. The largest zone of inhibition was obtained for *E. coli* (10⁻³ dilution) with 100 % concentration of essential oil of *E. globulus* and the lowest for *S. aureus* (10⁻¹ dilution) with 25 % concentration of essential oil leaves.

A more significant inhibition was seen with a higher essential oil concentration. At low concentrations, a very

limited inhibitory effect was observed on the growth of microorganisms in comparison with those controls.

With increasing essential oil leaves concentration, an obvious inhibitory effect on the growth of and *S. aureus*, was significantly increased.

Like previous test, the application of the dilution broth method confirms by its results shown in Table 1 the important antibacterial activity of the essential oil of *E. globulus* leaves on these two microbial strains.

In most cases, the essential oil of *E. globulus* leaves has the same inhibitory effect on both germs except for the dilution 10^{-3} where we see that the gram (–) bacterium *E. coli* was found to be more sensitive to the oil than the gram (+) bacterium *S. aureus*.

4. Discussion

The addition of essential oil leaves in broth culture inoculated with *S. aureus* and *E. coli* inhibited the growth of these organisms. The rate of inhibition was greater, on gram negative bacteria (*E. coli*) than that observed on gram positive bacterium (*S. aureus*). In most cases the size of inoculum and the concentration of essential oil leaves affected the growth/survival of the organisms.

These results are almost similar to those shown by other works on the antimicrobial activity of essential oil of *E. globulus* leaves as well as those of similar species^[14,21–29], and confirms its traditional uses^[26,30,31].

The growths of tested bacteria in high concentrations of essential oil leaves were highly inhibited, where it was considered that these organisms were sensitive to the oil.

Some authors have reported that gram–negative microorganisms are slightly more sensitive to essential oils when compared to gram–positive^[32–42]. The gram–positive and gram–negative microorganisms differ in several aspects other than with respect to the structure of their cellular walls, mainly with regard to the presence of lipoproteins and lipopolysaccharides in gram–negative bacteria that form a barrier to hydrophobic compounds^[43,44].

Some researchers reported that there is a relationship between the chemical structures of the most abundant in the tested essential oil and the antimicrobial activity.

The antibacterial activity of Eucalyptus extracts has been due to the components such as 1,8–cineole, citronellal, citronellol, citronellyl acetate, p–cymene, eucamalol, limonene, linalool, β – pinene, γ –terpinene, α – terpinol, alloocimene and aromadendrene^[45].

The essential oils from the leaf of *E. globulus* showed varying degrees of antibacterial activity against two clinical isolates. From the above experiment it can be inferred that extract suggest significant growth inhibiting effects on Gram–positive (*E. coli*) and Gram–negative bacteria (*S. aureus*). The efficacy of leaf oil of *E. globulus* against these microorganisms may provide a scientific ground for the application of the herb in the prevention and treatment of bacterial infections caused by various pathogenic bacteria such as Staphylococcus aureus and Escherichia coli, which have developed resistance to antibiotics. The incorporation of this oil into the drug formulations is also recommended.

The results of this study present the herb as a good candidate to explore new alternative antibacterial agents to combat pathogenic microorganisms.

Conflict of interest statement

We declare that we have no conflict of interest.

References

- [1] Ali N, Ahmed G, Ali Shah S, Shah I, Ghias M, Khan I. Acute toxicity, brine shrimp cytotoxicity and relaxant activity of fruits of callistemon citrinus Curt. *BMC Complement Altern Med* 2011; **11**:99.
- [2] Akin M, Aktumsek A, Nostro A. Antibacterial activity and composition of the essential oils of *Eucalyptus camaldulensis* Dehn and *Myrtus communis* L. growing in Northern Cyprus. *Afr J Biotechnol* 2010; **9** :531–535.
- [3] Mubita C, Syakalima M, Chisenga C, Munyeme M, Bwalya M, Chifumpa G, et al. Antibigrams of faecal *Escherichia coli* and *Enterococci* species isolated from pastoralist cattle in the interface areas of the Kafue basin in Zambia. *Veterinarski Arhiv* 2008; **78**(2): 179–185.
- [4] Boulekbache–Makhlouf L, Meudec E, Chibane M, Mazauric JP, Slimani S, Henry M, et al. Analysis by high–performance liquid chromatography diode array detection mass spectrometry of phenolic compounds in fruit of *Eucalyptus globulus* cultivated in Algeria. *J Agric Food Chem* 2010; **58**(24):12615–12624.
- [5] Bajaj YPS. *Medicinal and aromatic plants*. Volume 8, Biotechnology in agriculture and forestry. Berlin, Heidelberg, New York: Springer Edition; 1995, P. 194–196.
- [6] Elliot WR, Jones D. The Encyclopaedia of Australian plants, Vol.4. Melbourne: Lothian Publishing Company Pty Ltd; 1986.
- [7] Owlia P, Sadari H, Rasooli I, Sefidkon F. Antimicrobial characteristics of some herbal Oils on Pseudomonas aeruginosa with special reference to their chemical compositions. *Indian J Pharmacol Res* 2009; **8** (2): 107–114.
- [8] Lestari ES. Antimicrobial resistance among Staphylococcus aureus and Escherichia coli isolates in the Indonesian population inside and outside hospitals. 14th European Congress of Clinical Microbiology and Infectious Diseases. Prague / Czech Republic, May 1–4; 2004.
- [9] Jose B, Jogi Reddy L. Evaluation of antibacterial activity of the leaf and flower essential oils of *Gliricidia sepium* from south India. *Intl J App Pharm*. 2010; **2**: 20–22.
- [10] Chambers HF, Deleo FR. Waves of resistance: Staphylococcus aureus in the antibiotic era. *Nat Rev Microbiol* 2009; **7**(9):629–641.
- [11] Karou SD, Nadembega MCW, Zeba B, Ilboudo DP, Ouermi D, Pignatelli S, et al. Évolution de la résistance de Staphylococcus aureus aux antibiotiques au Centre Médical Saint Camille de Ouagadougou. *Médecine Tropicale* 2010; **70**(3) : 241–244.
- [12] Hossain MT, Siddique MP, Hossain FMA, Zinnah MA, Hossain MM, Alam MK, et al. Isolation, identification, toxin profile and antibiogram of Escherichia coli isolated from broilers and layers in Mymensingh district of Bangladesh. *Bangl J Vet Med* 2008; **6** (1): 1–5.
- [13] Arredondo–García JL, Amábile–Cuevas CF. High resistance prevalence towards ampicillin, co–trimoxazole and ciprofloxacin,

- among uropathogenic *Escherichia coli* isolates in Mexico City. *J Infect Developing Countries* 2008; **2**(5): 350–353.
- [14] Ait-Ouazzou A, Lorán S, Bakkali M, Laglaoui A, Rota C, Herrera A, et al. Chemical composition and antimicrobial activity of essential oils of *Thymus algeriensis*, *Eucalyptus globulus* and *Rosmarinus officinalis* from Morocco. *J Sci Food Agric* 2011; **91**(14):2643–2651.
- [15] Song A, Wang Y, Liu Y. Study on the chemical constituents of the essential oil of the leaves of *Eucalyptus globulus* Labill from China. *Asian J Traditional Med* 2009; **4**: 134– 140.
- [16] Von Graevenitz A. Practical substitution for the indol, methyl red, Voges–Proskauer, citrate system. *Applied Microbiol* 1971; **21**:1107–1109.
- [17] Ben Hassen S, Messadi L, Ben Hassen A. Identification et caractérisation des espèces de *Staphylococcus* isolées de lait de vaches atteintes ou non de mammites. *Ann Méd Vét* 2003; **147** : 41–47.
- [18] Maidment C, Dyson A, Beard J. A study into measuring the antibacterial activity of lysozyme-containing foods. *Nutr Food Sci.* 2009; **39** (1):29 – 35.
- [19] Tepe B, Donmez E, Unlu M, Candan F, Daferera D, Vardar-Unlu G, et al. Antimicrobial and antioxidative activities of the essential oils and methanol extracts of *Salvia cryptantha* (Montbret et Aucher ex Benth.) and *Salvia multicaulis* (Vahl). *Food Chem* 2004; **84**: 519–525.
- [20] Bouhadjera K, Bendahou ZM, Tabti B. Anti-microbial Activity of Extracts from Algerian *Aristida pungens* L. *Pak J Biol Sci* 2005; **8**: 206 –210.
- [21] Elaissi A, Hadj Salah K, Mabrouk S, Mohamed Larbi K, Chemli R, Skhiri FH. Antibacterial activity and chemical composition of 20 *Eucalyptus* species' essential oils. *Food Chem* 2011; **129**(4):1427–1434.
- [22] Bachheti RK, Joshi A, Singh A. Oil content variation and Antimicrobial activity of *Eucalyptus* leaves oils of three different Species of Dehradun, Uttarakhand, India. *Int J Chem Tech Res* 2011; **3**(2): 625–628.
- [23] Inouye S, Takizawa T, Yamaguchi H. Antibacterial activity of essential oils and their major constituents against respiratory tract pathogens by gaseous contact. *J Antimicrob Chemother* 2001; **47**:565–573.
- [24] Salari MH, Amine G, Shirazi MH, Hafezi R, Mohammadypour M. Antibacterial effects of *Eucalyptus globulus* leaf extract on pathogenic bacteria isolated from specimens of patients with respiratory tract disorders. *Clin Microbiol Infect* 2006; **12**:194–196.
- [25] Chung KH, Yang KS, Kim J, Kim JC, Lee KY. Antibacterial activity of essential oils on the growth of *Staphylococcus aureus* and measurement of their binding interaction using optical biosensor. *J Microbiol Biotechnol* 2007; **17**:1848–1855.
- [26] Cermelli C, Fabio A, Fabio G, Quaglio P. Effect of *Eucalyptus* essential oil on respiratory bacteria and viruses. *Curr Microbiol* 2008; **56**:89–92.
- [27] Chhetri HP, Yogol NS, Sherchan J, Anupa KC, Mansoor S, Thapa P. Phytochemical and antimicrobial evaluations of some Medicinal plants of Nepal. *Kathmandu Univ J Sci Engineer Technol.* 2008; **1**(V): 49–54.
- [28] Armando CC, Rahma HY. Evaluation of the yield and the antimicrobial activity of the essential oils from: *Eucalyptus globulus*, *Cymbopogon citratus* and *Rosmarinus officinalis* in Mbarara district (Uganda). *Rev Colombiana Cienc Anim* 2009; **1**:240–249.
- [29] Fit IN, Rapuntean G, Rapuntean S, Chirila F, Nadas GC. Antibacterial effect of essential vegetal extracts on *Staphylococcus aureus* compared to Antibiotics. *Not Bot Hort Agrobot Cluj* 2009; **37**: 117–123.
- [30] Gil L, Tadesse W, Tolosana E, López R. *Eucalyptus* species management, history, status and trends in Ethiopia. Proceedings from the Congress held in Addis Ababa. September 15th–17th, 2010.
- [31] Tyagi AK, Malik A. Antimicrobial potential and chemical composition of *Eucalyptus globulus* oil in liquid and vapour phase against food spoilage microorganisms. *Food Chem* 2011; **126**(1): 228–235.
- [32] Chaudhry NMA, Tariq P. *In vitro* antibacterial activities of Kalonji, Cumin and Poppy seed. *Pakistan J Bot* 2008; **40**: 461–467.
- [33] Farah H, Abdelhafid B. Huiles essentielles, utilisations et activités biologiques. Application à deux plantes aromatiques. *Les technologies de laboratoire* 2008; **8**:20–27.
- [34] Pirbalouti AG, Malekpoor F, Enteshari S, Yousefi M, Momtaz H, Hamed B. Antibacterial activity of some folklore medicinal plants used by Bakhtiari tribal in Southwest Iran. *Intl J Biol* 2010; **2**: 55– 63.
- [35] Oyedemi SO, Afolayan AJ. Antibacterial and antioxidant activities of hydroalcoholic stem bark extract of *Schotia latifolia* Jacq. *Asian Pac J Med* 2011; **4**(2): 952–958.
- [36] Madhumitha G, Saral AM. Preliminary phytochemical analysis, antibacterial, antifungal and anticandidal activities of successive extracts of *Crossandra infundibuliformis*. *Asian Pac J Med* 2011; **4**(3): 192–195.
- [37] Johnson M, Wesely EG, Kavitha MS, Uma V. Antibacterial activity of leaves and inter-nodal callus extracts of *Mentha arvensis* L. *Asian Pac J Med* 2011; **4**(3): 196–200.
- [38] Peixoto JRO, Silva GC, Costa RA, José res Lira de Sousa Fontenelle, Vieira GH, Filho AAF, et al. *In vitro* antibacterial effect of aqueous and ethanolic *Moringa* leaf extracts. *Asian Pac J Med* 2011; **4**(3): 201–204.
- [39] Chatterjee SK, Bhattacharjee I, Chandra G. Isolation and identification of bioactive antibacterial components in leaf extracts of *Vangueria spinosa* (Rubiaceae). *Asian Pac J Med* 2011; **4**(1): 35–40.
- [40] Johnson M, Wesely EG, Zahir Hussain MI, Selvan N. *In vivo* and *in vitro* phytochemical and antibacterial efficacy of *Baliospermum montanum* (Willd.) Muell. Arg. *Asian Pac J Med* 2011; **3**(11): 894–897.
- [41] Ahmed Moussa, Djebli Nouredine, Meslem Abdelmelek, Aissat Saad. Antibacterial activity of various honey types of Algeria against Pathogenic Gram–Negative Bacilli: *Escherichia coli* and *Pseudomonas aeruginosa*. *Asian Pac J Dis* 2012; **2**(3): 211–214.
- [42] Ravikumar S, Gokulakrishnan R, Boomi P. *In vitro* antibacterial activity of the metal oxide nanoparticles against urinary tract infectious bacterial pathogens. *Asian Pac J Dis* 2012; **2**(2): 85–89.
- [43] Wang J, Chen C. Biosorbents for heavy metals removal and their future. *Biotechnol Adv* 2009; **27**(2):195–226.
- [44] Mazutti M, Mossi AJ, Cansian RL, Corazza ML, Dariva CJ, Oliveira V. Chemical profile and antimicrobial activity of Boldo (*Peumus boldus* Molina) extracts obtained by compressed carbon dioxide extraction. *Braz J Chem Eng* 2008; **25**: 427– 434.
- [45] Nezhad FM, Zeigham H, Mota A, Sattari M, Yadegar A. Antibacterial activity of eucalyptus extracts on methicillin resistance *Staphylococcus aureus*. *Res J Biol Sci* 2009; **4**(8): 905–908.