

Contents lists available at ScienceDirect

# Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.elsevier.com/locate/apjtb



Document heading

doi: 10.1016/S2221-1691(15)30159-3

©2015 by the Asian Pacific Journal of Tropical Biomedicine. All rights reserved.

# Medicinal plants with hepatoprotective activity in Iranian folk medicine

Majid Asadi-Samani<sup>1</sup>, Najme Kafash-Farkhad<sup>2</sup>, Nafiseh Azimi<sup>3</sup>, Ali Fasihi<sup>4</sup>, Ebrahim Alinia-Ahandani<sup>2</sup>, Mahmoud Rafieian-Kopaei<sup>1\*</sup>

<sup>1</sup>Medical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran

### PEER REVIEW

#### Peer reviewer

Prof. Si-Yuan Pan, Department of Pharmacology, School of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing 100102, China.

Tel: +86 010 9473 8627 E-mail: siyuan-pan@163.com

#### Comments

This is a valuable review. It introduced 15 plants which are used as hepatoprotectives in Iranian folk medicine. This paper will promote the utilization of natural and traditional resources for contemporary health care. Herbal medicines have an extremely valuable, rich, lengthy, and extensive practical history.

Details on Page 153

### ABSTRACT

There are a number of medicinal combinations in the Iranian traditional medicine which are commonly used as tonic for liver. In this review, we have introduced some medicinal plants that are used mainly for the treatment of liver disorders in Iranian folk medicine, with focus on their hepatoprotective effects particularly against CC1<sub>4</sub> agent. In this study, online databases including Web of Science, PubMed, Scopus, and Science Direct were searched for papers published from January 1970 to December 2013. Search terms consisted of medicinal plants, traditional medicine, folk medicine, hepatoprotective, Iran, liver, therapeutic uses, compounds, antioxidant, CC1<sub>4</sub>, anti-inflammatory, and antihepatotoxic, hepatitis, alone or in combination. Allium hirtifolium Boiss., Apium graveolens L., Cynara scolymus, Berberis vulgaris L., Calendula officinalis, Nigella sativa L., Taraxacum officinale, Tragopogon porrifolius, Prangos ferulacea L., Allium sativum, Marrubium vulgare, Ammi majus L., Citrullus lanatus Thunb, Agrimonia eupatoria L. and Prunus armeniaca L. are some of the medicinal plants that have been used for the treatment of liver disorders in Iranian folk medicine. Out of several leads obtained from plants containing potential hepatoprotective agents, silymarin, β-sitosterol, betalain, neoandrographolide, phyllanthin, andrographolide, curcumin, picroside, hypophyllanthin, kutkoside, and glycyrrhizin have been demonstrated to have potent hepatoprotective properties. Despite encouraging data on possibility of new discoveries in the near future, the evidence on treating viral hepatitis or other chronic liver diseases by herbal medications is not adequate.

### **KEYWORDS**

Medicinal plants, Iran, Compounds, Liver, Therapeutic uses, CC14

### 1. Introduction

Liver diseases which are still a global health problem may be classified as acute or chronic hepatitis (inflammatory liver diseases), hepatosis (non inflammatory diseases) and cirrhosis (degenerative disorder resulting in liver fibrosis). Unfortunately, treatments of choice for liver diseases are controversial because conventional or synthetic drugs for the treatment of these diseases are insufficient and sometimes cause serious side effects[1].

Since ancient times, mankind has made use of plants in the treatment of various ailments because their toxicity factors appear to have lower side effects[2]. Many of the currently available drugs

\*Corresponding author: Mahmoud Rafieian-Kopaei, Medical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

Tel: +98 (381)3349509 Fax: +98 (381)3349509

E-mail: biology\_2011@yahoo.com

Foundation Project: Supported by Deputy of Research and Technology of Shahrekord University of Medical Sciences (Grant No. 2132-75).

Article history:
Received 25 Sep 2014
Received in revised form 28 Oct, 2nd revised form 30 Oct 2014
Accepted 25 Nov 2014
Available online 8 Dec 2014

<sup>&</sup>lt;sup>2</sup>Department of Biology, Faculty of Science, Urmia University, Urmia, Iran

<sup>&</sup>lt;sup>3</sup>Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran

<sup>&</sup>lt;sup>4</sup>Department of Genetics, Faculty of Biology Science, Tarbit Modares University, Tehran, Iran

were derived either directly or indirectly from medicinal plants. Recent interest in natural therapies and alternative medicines has made researchers pay attention to traditional herbal medicine. In the past decade, attention has been centered on scientific evaluation of traditional drugs with plant origin for the treatment of various diseases. Due to their effectiveness, with presumably minimal side effects in terms of treatment as well as relatively low costs, herbal drugs are widely prescribed, even when their biologically active constituents are not fully identified[3].

The utility of natural therapies for liver diseases has a long history. Despite the fact that most recommendations are not based on documented evidence, some of these combinations do have active constituents with confirmed antioxidant, anti-inflammatory, anticarcinogenic, antifibrotic, or antiviral properties. Although a large number of these plants and formulations have been investigated, the studies were mostly unsatisfactory. For instance, the therapeutic values, in most of these studies, were assessed against a few chemicals-induced subclinical levels of liver damages in rodents. The reasons that make us arrive at such a conclusion are lack of standardization of the herbal drugs, limited number of randomized placebo controlled clinical trials, and paucity of traditional toxicologic evaluations[4].

Hundreds of plants have been so far examined to be taken for a wide spectrum of liver diseases[5.6]. Natural products, including herbal extracts, could significantly contribute to recovery processes of the intoxicated liver. According to reliable scientific information obtained from the research on medicinal plants, plants such as *Silybum marianum*, *Glycyrrhiza glabra*, *Phyllanthus* species (amarus, niruri, emblica), and *Picrorhiza kurroa* have been widely and most of the time fruitfully applied for the treatment of liver disorders, exerting their effects via antioxidant-related properties[7-10].

Iranians have been using herbal medicine for the treatment of some common diseases; as a result, a large number of studies have been conducted to suggest new wild medicinal plants in different parts of Iran. Iranian traditional medicine is mostly relied on the consumption of plant materials. One of the important and well-documented utilities of plant products is their use as hepatoprotective agents. There are a number of medicinal combinations in the Iranian traditional medicine which are commonly used as tonic for liver. Allium hirtifolium Boiss. (A. hirtifolium), Apium graveolens L. (A. graveolens), Cynara scolymus (C. scolymus), Berberis vulgaris L. (B. vulgaris), Calendula officinalis (C. officinalis), Nigella sativa L. (N. sativa), Taraxacum officinale (T. officinale), Tragopogon porrifolius (T. porrifolius), Prangos ferulacea L. (P. ferulacea), Allium sativum L. (A. sativum), Marrubium vulgare L. (M. vulgare), Ammi majus L. (A. majus), Citrullus lanatus (C. lanatus), Agrimonia eupatoria L. (A. eupatoria) and Prunus armeniaca L. (P. armeniaca) are some of medicinal plants that have been used mainly for the treatment of liver disorders in Iranian folk medicine.

### 2. Medicinal plants

### 2.1. A. hirtifolium

A. hirtifolium from Alliaceae family, commonly known as Persian shallot (moosir in Persian) is endemic to Iran[11]. Based on available pharmaceutical investigations, antioxidant and hepatoprotective effects of A. hirtifolium have been also demonstrated. In addition, A. hirtifolium extracts had antioxidant properties comparable to or slightly higher than garlic extracts[12].

The commonly known phytochemical compounds identified in *A. hirtifolium* are saponins, sapogenins, sulphur containing compounds (*e.g.* thiosulfinates) and flavonoids including shallomin, quercetin and kaempferol<sup>[12]</sup>. Alliin, alliinase, allicin, s-allyl-cysteine, diallyl disulphide, diallyltrisulphide, and methylallyltrisulphide are the most important biological secondary metabolites of *A. hirtifolium*[13]. Disulphide and trisulfide compounds are among the most important compounds existing in *A. hirtifolium*[14]. Researches have shown that both the corn and the flower of shallot contain a high density of glycosidic flavonols. Linolenic, linoleic, palmitic, palmitoleic, stearic, and oleic acids have been identified in *A. hirtifolium* oil, as well<sup>[15]</sup>.

Treating rats with hydroalcoholic extract of A. hirtifolium could protect liver cells against oxidant effects of alloxan, and consequently caused a significant reduction in serum concentration of alkaline phosphatase (ALP), alanine transaminase (ALT), and aspartate transaminase (AST). Biochemical results have confirmed the usefulness of A. hirtifolium extract in decreasing the destructive effects of alloxan on liver tissue, and consequently decreasing the enzymes' leakage into cytosol, which is possibly achieved by herbal antioxidant compounds including flavonoids[12]. It was also reported that consumption of A. hirtifolium caused a reduction in AST level compared to the group with a hypercholesterolemic diet[16]. A research on the effect of hydroalcoholic A. hirtifolium extract on the level of liver enzymes in streptozotocin-induced diabetic rats indicated that hydroalcoholic extract of A. hirtifolium could significantly decrease serum levels of liver enzymes [AST, ALT, ALP and (lactate dehydrogenase) LDH] in a dose-dependent manner. Antioxidant micronutrients in the extract of A. hirtifolium may also restore liver damages. Shallomin and other active constituents of A. hirtifolium did not produce any adverse effect on the organs such as liver and kidney[17].

### 2.2. A. graveolens

A. graveolens, commonly known as celery, is an edible plant of the Umbelliferae family that grows mostly in the Mediterranean areas. It has been considered as a medicinal plant for a long time[18,19]. Data obtained from literature reveal that A. graveolens has many pharmacological properties such as antifungal, antihypertensive, antihyperlipidemic, diuretic, and anticancer[20-23]. This plant has also been shown to have some other medicinal features including

hyperlipidemic effects as well as antioxidative and hepatoprotective activities[20].

The active constituents are isoimperatorin, isoquercitrin, linoleic acid, coumarins (seselin, osthenol, apigravin, and celerin), furanocoumarins (including bergapten), flavonoids (apigenin, apiin), phenolic compounds, choline, and unidentified alkaloids[21]. *A. graveolens* is full of betacarotene, folic acid, vitamin C, sodium, magnesium, silica, potassium, chlorophyll, and fiber. The essential oil contains deltalimonene and various sesquiterpene[21,22].

Seeds of A. graveolens are used in Iranian medicine for liver ailments and disorders, have effects on liver, and exhibit hepatoprotective activities. Examining the antihepatotoxic effect of A. graveolens seeds' methanolic extracts on rats' liver showed a significant hepatoprotective activity[21]. The roots open obstruction of the liver and spleen, and help in dropsy and jaundice treatment[23]. Due to apigenin-related anti-inflammatory and antioxidant properties, A. graveolens seeds could counteract the pro-oxidant effect of 2-acetylaminofluorine through scavenging superoxide radicals, consequently declining hepatic glutathione-S-transferase (GST) and decreasing release of γ-glutamyl transpeptidase in serum; as a result, A. graveolens could be assumed as a potent plant against experimentally induced hepatocarcinogenesis in rats[24]. In addition, different extracts of the plant were examined for their hepatoprotective activity against CC14-induced hepatotoxicity in albino rats. The methanolic extracts, comparable to silymarin as a conventional drug, exhibited a higher hepatoprotective activity[25]. Another study indicated that the extracts of A. graveolens root significantly decreased CC1<sub>4</sub>-induced acute hepatic injury, increasing the activities of AST and ALT and preventing CC1<sub>4</sub>-induced acute liver injury[26]. Crude ethanol extract of the whole plant was indicative of anti-inflammatory effects in rats. Furthermore, topical anti-inflammatory effects of A. graveolens leaves' extract have been demonstrated by Mencherini et al[27]. Significant anti-inflammatory effect of the aqueous and hexane extracts of A. graveolens was shown at all doses (100-500 mg/kg body weight). Both extracts presented remarkable anti-inflammatory effect, which confirmed the traditional use of A. graveolens in inflammation-associated diseases[28].

## 2.3. C. scolymus

C. scolymus (artichoke) from Apiaceae family, a species of perennial thistle and with a Mediterranean origin, is traditionally used for the treatment of digestive disorders, moderate hyperlipidemia, and liver and bile diseases. The leaf extract of C. scolymus has been used for its hepatoprotective effects[29]. Also, C. scolymus extract could yield nutritional supplements with antioxidant and antimicrobial effects[30]. In C. scolymus leaf extract, there are compounds such as cynarin, luteolin, chlorogenic acid, and caffeic acid, other flavonoids, and polyphenol compounds, some of which have antioxidant properties. C. scolymus leaf extract also positively

affected the changes in rat serum liver enzyme induced by CC14 and histopathological damage to liver tissue[31]. In rats pretreated with artichoke extract, plasma transaminase activities significantly decreased and histopathological changes in the liver ameliorated[32]. C. scolymus can be conducive to the reduction in phosphatidate phosphohydrolase activity and liver triglyceride. C. scolymus has benefits for controlling of hyperlipidemia, oxidative stress in hyperlipidemic regimes, and abnormalities in lipid profiles[33]. In the rabbits intoxicated with CC14, C. scolymus leaf extract counteracted the toxic effect of CC14, blood sugar, cholesterol, triglycerides, leukocytes, and a number of erythrocytes[34]. In other studies, C. scolymus was significant in keeping the normal liver function parameters, maintained the hepatic redox status as it is manifested by significant increase in antioxidant enzyme activities and reduction in glutathione accompanied by inhibition of lipid peroxidation (LPO) and protein oxidation, decreased nitric oxide and tumor necrosis factor alpha, and stabilized membrane in the untreated paracetamolintoxicated rats[35].

### 2.4. B. vulgaris

B. vulgaris (barberry), a well known medicinal plant in Iran and also a food, belongs to Berberidaceae family. As a shrub with 1 to 3 meters in height, B. vulgaris grows in many regions of the world, including Iran (especially Khorasan)[36]. Fruit, leaves, and stem have medical usages including hepatoprotection. B. vulgaris fruit extract contains various flavonoids that act as antioxidant[37]. Berberine, oxyacanthine, and other alkaloids such as berbamine, palmatine, columbamine, malic acid, jatrorrhizine, and berberrubine comprise some other compounds[38]. Stigmasterol, terpenoids lupeol, oleanolic acid, stigmasterol glucoside and polyphenols were also identified in this plant[39]. Berberine, an isoquinoline alkaloid with a long medicinal history, exists in roots, rhizomes, and stem bark of the plant. Berberine inhibits potassium and calcium currents in isolated rat hepatocytes. It has hepatoprotective effects, both preventive and curative, on CC14-induced liver injury through scavenging the peroxidative products. CC14 significantly increased the serum alanine aminotransferase, aspartate aminotransferase, and alkaline phosphatase levels in rats. Treatment with the methanolic extract of B. vulgaris fruit significantly helped these changes reach to an almost normal level. In addition, the extract could prevent CC14induced liver oxidative damage in rats[40]. Domitrović's study was indicative of berberine's effect on protecting the liver from CC1<sub>4</sub>induced injury. The hepatoprotective mechanisms of berberine could be attributed to the free radical scavenging, decline in oxidative/ nitrosative stress, and the inhibition of inflammatory response in the liver[41]. In addition, B. vulgaris extract/β-cyclodextrin exhibited better hepatoprotective effects than free extract on oral administration possibly due to greater bioavailability. Formulated extract could be used as an economical phytotherapeutical supplement that is helpful for chronic or acute conditions or a support for routine therapies of serious hepatic disorders. In Hermenean's study, pre-treatment with formulated or non formulated extract prevented the increase in ALT, AST, and malondialdehyde (MDA) levels, and helped the level of antioxidant enzymes return to normal values. According to histopathological and electron-microscopic examination, in both pre-treated groups, more moderate damage in liver was observed with a more pronounced protective effect after administration of the formulated extract[42].

### 2.5. C. officinalis

C. officinalis (marigold), from Asteraceae family, is a medicinal plant and cosmetic herb popularly known in Europe and the USA. The dried flower heads or the dried ligulate flowers of this plant are used for pharmaceutical and/or cosmetic purposes[43]. Antibacterial, anti-inflammatory, antiviral, and antioxidant activities have been already noted for C. officinalis[44]. It has been taken in order to treat fevers and jaundice and to promote menstruation. Extracts, tinctures, balms, and salves of C. officinalis have been applied directly to heal wounds and soothe inflamed and injured skin. C. officinalis compounds, which are potentially active chemical constituents, are monoterpenes, such as α-thujene and T-muurolol, sesquiterpene and flavonol glycosides, triterpene alcohols, triterpenoid saponins, flavonoids, carotenoides, xanthophylls, phenolic acids, mucilage, bitters, phytosterols, tocophrrols, calendulin, resin, and volatile oil[45,46]. The anti-inflammatory features of C. officinalis flowers, according to in vivo pharmacological tests, have been associated with the triterpenoid fatty acid esters[43]. In Singh's study, 80% effect of methanolic extract of leaves (500 mg/kg orally, four doses at 12 hours interval) of C. officinalis was investigated against acetaminophen-induced hepatic damage in albino rats. The potential hepatoprotective effects of C. officinalis extracts against CC1<sub>4</sub>induced oxidative stress and cytotoxicity in isolated primary rat hepatocytes were detected[46], confirmed by significant improvement in cell viability and enzymes leakages (ALT, AST, and LDH). Also, the reduction of hepatocytolysis and steatosis, and return to normal values of various enzymes activity could be attributed to hepatoprotective effects<sup>[47]</sup>. C. officinalis plant extracts significantly improved cell survival, contributing greatly to preserving the cellular membranes integrity against CC14. Moreover, plant extracts of C. officinalis protect the intracellular antioxidant defense system, indicated by preserving GST and inhibiting LPO[48]. Protective role of the flower extract of C. officinalis against CC14- induced acute hepatotoxicity and cisplatin-induced nephrotoxicity has been shown[49]. Possible mechanism of action of the flower extract may be due to its antioxidant activity and reduction of oxygen radicals[50].

# 2.6. N. sativa

N. sativa is an aromatic plant of Ranunculaceae family, traditionally used by the Middle East nations for asthma, cough,

bronchitis, headache, rheumatism, fever, influenza, and eczema. Several biological activities, including antioxidant activity and resolution of hepato-renal toxicity have been reported for N. sativa seeds[51]. N. sativa contains more than 30 fixed oils. The volatile oil has been proved to contain thymoquinone and many monoterpenes such as p-cymene and α-pinene. The CC1<sub>4</sub> treatment increased the LPO and liver enzymes, and decreased the antioxidant enzyme levels. N. sativa treatment helped the elevated LPO and liver enzyme levels decrease and the reduced antioxidant enzyme levels increase[52]. The levels of liver enzymes and total oxidative status, oxidative stress index, and myeloperoxidase in treated mice were significantly lower, and total antioxidant capacity in liver tissue was significantly higher compared to the controls[53]. N. sativa is useful in the treatment of rheumatism and related inflammatory diseases and the antiinflammatory effect was confirmed in rats[54]. Also, the aqueous extract of N. sativa has an anti-inflammatory effect demonstrated by its inhibitory effects on carrageenan-induced paw edema[51]. Pretreatment of mice with 12.5 mg/kg thymoquinone (an N. sativa derived-compound) significantly reduced the elevated levels of serum enzymes as well as hepatic MDA content and significantly increased hepatic nonprotein sulfhydryl(-SH)[55]. N. sativa contributes to inhibition of enzymes present in the neoglucogenesis pathway in the liver[56].

### 2.7. T. officinale

T. officinale (from Asteraceae family), commonly known as dandelion, grows almost everywhere in the world[57]. With a long history of traditional use in the treatment of hepatobiliary problems, its root has been shown to have sesquiterpene lactones, triterpenes, carbohydrates, fatty acids (myristic), carotenoids (lutein), flavanoids (apigenin and luteolin), minerals, taraxalisin, coumarins, and cichoriin. Aesculin has been reported from the leaf. Germacraneand guaiane-type sesquiterpene lactones including taraxinic acid derivatives were obtained from the roots of this plant[58]. Also, several flavonoids, e.g. caffeic acid, chlorogenic acid, luteolin, and luteolin 7-glucoside, have been isolated from the dandelion[59]. Ethanolic extract of T. officinale was effective on decrease in serum ALT levels[58]. Hydroalcoholic acid extract of the root enhanced levels of superoxide dismutase (SOD), catalase (CAT), GST, and LPO[60]. Oral administration of extracts of the T. officinale roots has proved to increase bile flow[1]. Another study distinguished that treatment with root extract of T. officinale was effective on reduction of serum ALT and ALP levels in rats[61]. Root extract reduces serum AST, ALT, ALP, and LDH activities and increases hepatic antioxidant activities such as CAT, GST, glutathione reductase, glutathione peroxidase, and glutathione. Thus, aqueous extract of T. officinale root protects against alcohol-induced toxicity in the liver by elevating antioxidativity and decreasing LPO. Sesquiterpene lactones in the plant have a protective effect against acute hepatotoxicity induced by the administration of CC14 in mice, which was indicated by reduced levels of hepatic enzyme markers, such as serum transaminase (ALT, AST), ALP, and total bilirubin[62].

### 2.8. T. porrifolius

T. porrifolius, belonging to Asteraceae family and known as purple salsify, is grown up for its edible root and shoot[63]. It has bioactive compounds which prevent cancer or other free radicalassociated illnesses. The nutritional value of this plant is derived from monounsaturated and essential fatty acids, polyphenols, vitamins, and fructo-oligosaccharides, having probiotic effects on the intestinal microflora. The most abundant compounds of this plant include 4-vinyl guaiacol (19.0%), hexadecanoic acid (17.9%), hexahydrofarnesylacetone (15.8%), and hentriacontane (10.7%)[64,65]. T. porrifolius has apparently yielded the hepatogenic/hepatoprotective effects against liver diseases or hepatotoxicity induced by a variety of hepatotoxic agents such as chemicals, drugs, pollutants, and infection with parasites, bacteria, or viruses (hepatitis A, B, and C). These beneficial effects of plants are related to the polyphenolic compounds. The study of antioxidant activity of the methanolic extract of the aerial part of T. porrifolius as well as its protection against CC14-induced hepatotoxicity in rats showed a dose-dependent increase in the activity of liver antioxidant enzymes. About 250 mg/kg body weight dose increased the activity of CAT, SOD, and GST. Also, substantial hepatoprotective capacity against CC14-induced hepatic injury has been shown, attributable to restoring the activity of AST, ALT, and LDH to normal levels[63,66]. Investigation of the effects of water extract of *T. porrifolius* shoot on lipemia, glycemia, inflammation, oxidative stress, hepatotoxicity, and gastric ulcer using a rat model showed that after one month of T. porrifolius water extract intake, a significant decrease in the levels of serum cholesterol, triglyceride, glucose, and liver enzyme (ALP, ALT, and LDH) was observed. Pretreating rats with T. porrifolius extract demonstrated considerable anti-inflammatory effects in both acute and chronic inflammation caused by carrageenan and formalin. In addition, T. porrifolius revealed effective antioxidant capability owing to its remarkable scavenging activity[67].

### 2.9. P. ferulacea

P. ferulacea from Apiaceae family grows in Southern Iran and is used in Iranian herbal medicine mainly for gastrointestinal disorders. The genus of Prangos with the common Persian name of Jashir includes 15 species, occurring widely in many regions of the country. In addition to Iran, other species of this genus are distributed in East Europe to Turkey, Caucasia, and Central Asia. P. ferulacea has been used in folk medicine as carminative, emollient, and tonic for gastrointestinal disorders, antiflatulent, sedative, anti-inflamatory, anti-viral, antihelminthic, antifungal, and antibacterial. Monoterpenes, sesquiterpenes, coumarines, flavonoids, alkaloids, tannins, saponins, and terpenoids are some important compounds identified in this plant. P. ferulacea was shown that the oils (both

fruit and leaf essential oils) were rich in monoterpenes, specially α-pinene, and β-pinene. Some of these components have an antioxidant effect against oxidative stress. In a study, the protective and antioxidant effects of P. ferulacea are reported to be higher compared to  $\alpha$ -tocopherol (vitamin E) and the effect of GST has been demonstrated. The study of effects of P. ferulacea hydroalcoholic extract on changes in rats' liver structure and serum activities of ALT and AST after alloxan injection indicated that in diabetic rats, the serum ALT and AST significantly increased. Moreover, necrosis of hepatocytes, cytoplasmic vacuolations, and lymphocytic inflammation were observed. Diabetic rats treated by root extract of P. ferulacea in contrast to the diabetic group exhibited a significant decrease in these enzymes. Also, root hydroalcoholic extract of P. ferulacea was shown to affect changes in aminotransferases and to prevent the histopathological changes of liver related to alloxaninduced diabetes in rats[68-71].

#### 2.10. A. sativum

A. sativum (garlic) is one of the world's most known medicines that have been used for flavouring and as a medical herb mainly due to its prophylactic and therapeutic capacities. Garlic from Alliaceae family has known nutritional properties, particularly for its bioactive components, and is used as antidiabetic, antiinflammatory, antihypertension, antimicrobial, antiatherosclerotic, and hepatoprotective in different diet-oriented therapeutical regimes to heal various lifestyle-associated disorders[72]. Garlic and its supplements are taken in many cultures for their hypolipidemic, antiplatelet, and procirculatory effects. In Iran, it is known for being useful for gastrointestinal disorders. In addition, some garlic combinations may be immune-enhancing and chemopreventive. Some combinations could be antioxidative while others may stimulate oxidation. Sapogenins, saponins, sulphuric compounds, and flavonoids have been detected in different species of Allium genus[12]. Additional biological effects attributed to garlic extract may be due to S-allylcysteine, S-allylmercaptocysteine, and N (alpha)-fructosyl arginine that are formed throughout the extraction[72]. Most garlic's beneficial effects are due to organosulphate molecule allicin[73]. The hepatoprotective effect of garlic extracts on Cd-induced oxidative damage in rats has been reported. A. sativum extract decreased hepatic activities of ALT, AST, and alkaline phosphatase and simultaneously increased the plasma activities of ALT and AST. Cd-induced oxidative damage in rat liver is predisposed to decreasing by moderate dose of A. sativum extracts probably through reduced LPO and improved antioxidant defense system that could not prevent and protect Cd-induced hepatotoxicity[74]. A. sativum chemical compounds have curative effects on iron liver excess[75]. In another study, the hepatoprotective effects by A. sativum, ginger (Zingiber officinale), and vitamin E against CC14-induced liver damage were examined in male Wistar albino rats. Serum alanine amino transferase, aspartate amino transferase, and alkaline phosphatase levels decreased significantly

24 h after CC1<sub>4</sub> administration in rats pretreated with garlic, ginger, vitamin E, and various mixtures of garlic and ginger compared to the rats treated with only CC14. LPO expressed by serum MDA was assayed to assess the severity of liver damage by CC14, including the extent of hepatoprotection by garlic, ginger, and vitamin E. MDA concentration was significantly decreased in rats pretreated with garlic, ginger, vitamin E, and various mixtures of garlic and ginger compared to the rats administered by CC14 alone. Histological examination of the liver was indicative of severe infiltration of inflammatory cells in rats treated with CC14 alone although the change in the normal architecture of the hepatic cells decreased considerably in pre-treated rats[76]. The hepatoprotective activity of A. sativum extract at a dose of 300 mg/kg body weight, administered intraperitoneally for 14 d before the induction of D-galactosamine and lipopolysaccharide (DGalN/LPS) was investigated against DGalN/LPS-induced hepatitis in rats. Pretreatment with aqueous A. sativum extract helped the altered parameters (ALT, AST, ALP, LDH, gamma glutamyl transferase, bilirubin, LPO, tumor necrosis factor, and myeloperoxidase activity level, total cholesterol, triglycerides, free fatty acids, and antioxidant enzyme activities) reach to nearly normal control values. Aqueous A. sativum extract could afford a significant protection in the DGalN/LPS-induced hepatic damage easing[77]. An investigation of chemopreventive effects of A. sativum extract and silymarin on N-nitrosodiethylamine and CC1<sub>4</sub>-induced hepatotoxicity in male albino rats indicated synergistic effect of silymarin and A. sativum, and their hepatoprotective features against hepatotoxicity[78].

The main part of therapeutic effect of *A. sativum* is attributed to antioxidant compounds, probably associated with its phenolic compounds and flavonoid substances. *A. sativum* (irrespective of the processes such as baking) has phenolic, flavonoid, and flavenol compounds as well as allicin[79]. The effect of fresh *A. sativum* on inhibiting the oxidation was higher compared to three-month *A. sativum*. Phenolic compounds of the fresh *A. sativum* were higher than the three-month *A. sativum*. The amount of allicin was 15 µg/mL and 8 µg/mL in fresh and three-month dated aqueous *A. sativum* extract, respectively.

A. sativum could play a crucial role in prevention and control of some cancers. The phenolic compounds and antioxidant activity of A. sativum aerial parts are less in comparison to A. sativum itself; however, higher doses might have anticancer features. Also, A. sativum had anticancer activity against WEHI-164 tumor cells, and some processes such as heating reduced the effect noticeably. The anticancer activities of different kinds of A. sativum may be related to the level of allicin, flavonoids, and phenolic components. Therefore, fresh A. sativum has bioactive components and hence anticancer efficacy[80].

### 2.11. M. vulgare

M. vulgare, from Lamiaceae family, commonly known as "horehound", is a commonplace Mediterranean plant

traditionally used to treat various diseases. The plant possesses antihypertensive, analgesic, anti-inflammatory, and hypoglycemic effect, antidyslipidemic activity, and antioxidant properties[2,81]. Antimicrobial activity against Gram positive bacteria, analgesic properties, and antihypertensive, antidiabetic, and antioxidant properties were noted, particularly related to diterpenes, sterols, phenylpropanoids, and flavonoids. Totally 46 compounds, comprising 96.3% of the oil, were detected. The main components of the oil were (E)-caryophyllene, germacrene D, and bicyclogermacrene. Evaluation of the antihepatotoxic and antioxidant properties of the extract against CC14-induced hepatic damage in rats showed a significant antihepatotoxic effect via significantly reducing AST, ALT, and LDH[82]. In another study, aqueous extract of the whole plant was examined for antihepatotoxic activity against CC1<sub>4</sub>-induced hepatic damage in male Wistar rats. The extract at 500 mg/kg body weight dose for 7 d was compared with the standard drug silymarin at 10 mg/kg body weight dose. The aqueous extract had significant antihepatotoxic activity and reduced the elevated levels of serum enzymes such as serum glutamic-oxaloacetic transaminase, serum glutamic pyruvic transaminase, and ALP and increasing total protein[83]. Some studies also reported the antioxidant effect of free radical scavengers in the extract thanks to flavonoid content[2,82].

### 2.12. A. majus

A. majus from Apiaceae family was originated from Asia and is a 0.30 to 0.60 meter high, annual plant with ascending branches and leaves that are finely dissected into filiform segments. The inflorescence is umbellate with small white flowers. The involucels are of different linear bracts and the involucral bracts are pinnately parted. The main toxins are the furocoumarins. All parts of the plants, in particular the seeds, could be phototoxic to cattle, sheep, fowl, and humans after ingestion or skin contact and subsequent exposure to sunlight. A. majus is a local medicinal plant with fruits that are contraindicated in nursing, pregnancy, tuberculosis, liver and kidney diseases, human immunodeficiency virus, and other autoimmune diseases. It is commonly used for skin disorders such as psoriasis and vitiligo. A. majus is contraindicated in diseases associated with photosensitivity, cataract, invasive squamous-cell cancer, known sensitivity to xanthotoxin, and in children under the age of 12[84]. A. majus concomitantly accumulates various 7-Oprenylated umbelliferones as the predominant coumarines[85]. It is considered as a source of 6-hydroxy-7-methoxy coumarine which is known as the major coumarin. A. majus with confirmed antioxidant effect could be used in diabetic nephropathy and myocardial injury thanks to different active compounds such as quercetine, kaempferol, and marmesinin that inhibit cytochrome P450 such as xanthotoxin bergapten, imperatorin, and isoimpinellin. Treatment of rats with different doses of A. majus seeds' extract could cause hepatoprotective effects against CC14-induced liver damage, in a dose-dependent fashion[86].

### 2.13. C. lanatus

C. lanatus (from Cucurbitaceae family) is used in traditional herbal medicine. Its fruits are eaten as a febrifuge when fully ripe or almost putrid. The fruit is also diuretic and helpful for the treatment of dropsy and renal stones. The rind of the fruit is prescribed for alcoholic poisoning and diabetes. C. lanatus contains bioactive compounds including cucurbitacin, triterpenes, anthraquinones, sterols, alkaloids, flavanoids, saponins, tannins, flavones aglycone, and simple phenols[87]. The aqueous extract of *C. lanatus* is believed to be a good source of glucose, fibre, vitamin C, lycopene, and beta carotene. Epidemiologically, antioxidant may reduce or inhibit the effect of oxidative stress in tissues. Watermelon juice at 120 g/70 kg body weight of rats decreased SOD activity and low density lipoprotein-cholesterol, and increased CAT and high density lipoprotein-cholesterol, which could indicate its antioxidant effects[88]. The majority of cucurbitacin has cytoprotective activity on HepG<sub>2</sub> cells. Cucurbitacin was demonstrated to have high potential as liver anti-fibrosis agent[89]. Studies have been done to investigate the effect of C. lanatus juice on LPO in rat's liver, kidney, and brain. In vivo administration of CC14 once a week for 28 d caused a significant increase in serum markers of liver damage, AST, ALT, and total bilirubin, and decline in albumin compared to the control group. However, administration of CC14 with watermelon juice or ursodeoxycolic acid attenuated these changes significantly. LPO level increased in the liver, kidney, and brain tissues after CC1<sub>4</sub> administration. However, watermelon juice and ursodeoxycolic acid treatment prevented increase in LPO. According to the results, watermelon juice protects the liver, kidney, and brain tissues from in vitro CC14 toxicity in rats probably thanks to the antioxidant activity and inhibition of lipid peroxide formation. Together, biological evidence supports watermelon juice usefulness in the treatment of chemical-induced hepatotoxicity[90].

# 2.14. A. eupatoria

A. eupatoria (from Rosaceae family) is 35-120 cm high, semiorbicular, and rather thick, with densely hirsute stems, herbaceous stipules, coarsely acutely serrated or lobed margins, and pilose and pubescent petiole. The leaf blade is interrupted imparipinnate with 3-5 pairs of leaflets[91]. A. eupatoria grows on clay soils. In popular medicine, it is employed for the treatment of several disorders, e.g. inflammations. The aqueous extract of A. eupatoria is full of several phenolic compounds and its ethyl acetate fraction has exhibited antioxidant activity and lower toxicity[92]. A. eupatoria is rich in coumarins, flavonoids, tannins, terpenoids, and phenolic compounds including protocatechuic acid, coumaric acid, chlorogenic acid, quercitrin, and gallic acid[93]. A. eupatoria, a medicinal herb, caused effects on the liver cells in a preliminary study. However, the active components and the biologic effect of A. eupatoria on liver tumor remain to be fully elucidated. The hepatoprotective effects of A. eupatoria on hepatocarcinogenesis induced by diethylnitrosamine and CC14 were studied in the in vivo models. There is evidence on the biologic actions of A. eupatoria and its benefits for liver tumor therapy. The hepatoprotective effects of *A. eupatoria* water extract against ethanol-induced liver injury have been already shown. Animals were treated orally with *A. eupatoria* extract at 10, 30, 100, and 300 mg/kg/day doses. After ethanol chronic consumption, serum aminotransferase activities and proinflammatory cytokines pronouncedly increased, although attenuated by *A. eupatoria* extract. The cytochrome P450 activity and LPO also increased after ethanol consumption while glutathione concentration decreased. *A. eupatoria* extract ameliorates chronic ethanol-induced liver injury, and protection likely relates to the suppression of oxidative stress and toll-like receptor-mediated inflammatory signaling[94]. Hepatoprotective effects of aqueous extract of *A. eupatoria* were investigated in experimental liver-damaged models. Hepatoprotective effects of the plant were monitored by reducing serum AST and ALT levels[95].

### 2.15. P. armeniaca

The P. armeniaca (commonly called apricot) belongs to the Rosaceae family, with many medicinal properties. It has organic acids, salicylic acid, tannins and potassium salts, pcoumaric acid, and protocatechuic, ferulic, and diferulic acids. The plant is used as antitussive and antiasthmatic, with hepatoprotective effects[96]. The plant has two new flavonoid glycosides, 4', 5, 7-trihydroxy flavone-7-O-[ $\beta$ -D-mannopyranosyl (1"" $\rightarrow$ 2")]- $\beta$ -D-allopyranoside and 3, 4', 5, 7-tetrahydroxy-3, 5'-di-methoxy flavone 3-O- $[\alpha$ -Lrhamnopyranosyl (1''' $\rightarrow$ 6'')]- $\beta$ -D-galactopyranoside[97]. Fruit of the plant is rich in carotenoids, flavonoids, and phenols. Hepatoprotective effect of ground apricot kernel (GAK) (0.5,1 and 1.5 mg/kg/body weight/rat) was examined in rats injected with 10 mg/kg dimethylnitrosamine, demonstrating that the GAKsupplemented diet resulted in improving liver function, liver CAT, SOD, and GST and hence reducing AST, ALT, and MDA, which was confirmed by liver histology. Hierarchically high levels of GAK (1.5 mg/kg/body weight/rat) yielded the best results compared to other tested levels[98]. Animal studies have shown that P. armeniaca administration to rats with chronic ethanol feeding decreases the levels of ALT and AST in the serum, which reduces oxidative stress and LPO in the liver by increasing the levels of antioxidant enzymes. Studies showed that supplementation of β-carotene prevented ethanol-induced increase in the serum aminotransferases and inhibited the depletion of the antioxidant molecule GST in the liver. Additionally, in vitro studies on the hepatocytes isolated from the ethanol-fed rats indicated that β-carotene enhanced the cell viability, and increased CAT activity and level of GST. In mechanistic studies performed on hepatocytes isolated from the rats fed with ethanol, β-carotene ameliorated the oxidative stress, enhanced antioxidant, and decreased the expression of CYP2E1 and apoptosis. Lutein and meso-zeaxanthin present in P. armeniaca in small quantities are effective on treatment of alcohol-induced damage. Administering lutein and meso-zeaxanthin, compared to alcohol, reduced the serum levels of aminotransferases, alkaline phosphatase, and bilirubin to decrease the levels of LPO, conjugated diene, and hydroperoxides in rat liver. Based on histopathological studies, administering

ethanol-treated rats with lutein and meso-zeaxanthin reversed the histopathological abnormalities and reduced hydroxyproline, which is an indicator of fibrosis[99]. P. armeniaca fed to Wistar rats decreased oxidative stress and enhanced histological damage; its dietary intake can reduce the risk of liver steatosis caused by free radicals[100]. Examining hepatoprotective effect and antioxidant role of sun, sulphited-dried P. armeniaca, and its kernel against ethanol-induced oxidative stress indicated that administration of sun, sulphited-dried apricot, but not its kernel, supplementation restored the ethanol-induced imbalance between MDA and antioxidant system towards nearly normal values particularly in tissues. Altogether, apricot has a hepatoprotective effect in rats fed with ethanol, probably through the antioxidative defense systems[101]. P. armeniaca feeding had beneficial effects on CC14-induced liver steatosis and damage probably due to its antioxidant nutrient (beta-carotene and vitamin) contents and high radical-scavenging activity[102].

#### 3. Conclusion

The protective effect of these plants' extract against CC14 may be related to polyphenolic compounds, terpenoids, alkaloids, coumarines, phytosterols, etc. Polyphenolic compounds such as flavonoids can protect the cells against emptying reduced glutathione via increasing the capability of antioxidant enzymes (such as CAT, SOD and glutathione peroxidase). Flavonoids, which act as antioxidant, free radical scavenging and antilipoperoxidant agents, are helpful for hepatoprotection. Furthermore, these compounds with antioxidant properties can counteract free radicals in the environment and therefore avoid their destructive effects. Terpenoids such as carotenoids with antihepatotoxic activity are also known as antioxidants. Ursolic acid is a triterpene, with potential hepatoprotective effects. Also, out of several leads obtained from plants containing potential hepatoprotective agents, silymarin, β-sitosterol, betalain, neoandrographolide, phyllanthin, andrographolide, curcumin, picroside, hypophyllanthin, kutkoside, and glycyrrhizin have been demonstrated to have potent hepatoprotective properties. Silymarin and glycyrrhizin were significantly effective on treatment of hepatitis, alcohol-associated liver disease, and cirrhosis. β-sitosterol showed anti-inflammatory, antioxidant, angiogenic, and proliferative effects. Betalain-containing species are used as common medicines for various diseases (e.g. hepatic disorders, malaria, jaundice and scanty urine). Betalains can act as antioxidant, anticancer, antiviral, and antiparasitosis.

Despite encouraging data on possibility of new discoveries in the near future, evidence on treatment of viral hepatitis or other chronic liver diseases by herbal medications is not adequate. Therefore, herbal medications should be recommended within the setting of more finely-conducted clinical trials. Better training of both patients and physicians about herbal preparations seems necessary.

### **Conflict of interest statement**

The authors declare no conflict of interest.

### Acknowledgements

This work was funded by Deputy of Research and Technology of Shahrekord University of Medical Sciences (Grant No. 2132-75). We also thank all who contribute, in some way, to conducting this research.

### **Comments**

# Background

The human liver is one of the most important organs in the body. It has a wide range of functions, including detoxification, protein synthesis, and production of biochemicals necessary for digestion. Because of wrong lifestyle and dietary habits, food/drinking contamination, and chemical drug abuse, the incidence of liver diseases and/or liver function abnormalities is increasing in the world. Therefore, there is need of new hepatoprotective remedies, including herbal/folk medicines.

### Research frontiers

The present manuscript reviewed the plants which have hepatoprotection or antihepatotoxicity, liver tonic, in Iranian folk medicine.

### Related reports

In this study, online databases including Web of Science, PubMed, Scopus, and Science Direct were searched for papers published from January 1970 to December 2013.

### Innovations and breakthroughs

It is well known that there are many plants/herbs that have hepatoprotective action in the world. In the present study, authors have reviewed the hepatoprotective activity of folk medicines in Iran.

### **Applications**

From the literature survey, it has been found that Iranian folk medicine may be used as an adjuvant for the treatment and prevention of liver injury.

#### Peer review

This is a valuable review. It introduced 15 plants which are used as hepatoprotectives in Iranian folk medicine. This paper will promote the utilization of natural and traditional resources for contemporary health care. Herbal medicines have an extremely valuable, rich, lengthy, and extensive practical history.

# References

- Kumar CH, Ramesh A, Kumar JNS, Ishaq BM. A review on hepatoprotective activity of medicinal plants. *Int J Pharm Sci Res* 2011; 2: 501-515.
- [2] Elberry AA, Harraz FM, Ghareib SA, Gabr SA, Nagy AA, Abdel-Sattar

- E. Methanolic extract of *Marrubium vulgare* ameliorates hyperglycemia and dyslipidemia in streptozotocin-induced diabetic rats. *Int J Diabetes Mellit* 2011; **11**: 1877-1878.
- [3] Levy C, Seeff LD, Lindor KD. Use of herbal supplements for chronic liver disease. *Clin Gastroenterol Hepatol* 2004; **2**: 947-956.
- [4] Thyagarajan SP, Jayaram S, Gopalakrishnan V, Hari R, Jeyakumar P, Sripathi MS. Herbal medicines for liver diseases in India. *J Gastroenterol Hepatol* 2002; 17: S370-S376.
- [5] Asadi-Samani M, Rafieian-Kopaei M, Azimi N. Gundelia: a systematic review of medicinal and molecular perspective. *Pak J Biol Sci* 2013; 16: 1238-1247.
- [6] Asadi-Samani M, Bahmani M, Rafieian-Kopaei M. The chemical composition, botanical characteristic and biological activities of *Borago* officinalis: a review. Asian Pac J Trop Med 2014; 7: S22-S28.
- [7] McBride A, Augustin KM, Nobbe J, Westervelt P. Silybum marianum (milk thistle) in the management and prevention of hepatotoxicity in a patient undergoing reinduction therapy for acute myelogenous leukemia. J Oncol Pharm Pract 2012; 18: 360-365.
- [8] Tatiya AU, Surana SJ, Sutar MP, Gamit NH. Hepatoprotective effect of poly herbal formulation against various hepatotoxic agents in rats. *Pharmacognosy Res* 2012; 4: 50-56.
- [9] Shukla B, Visen P, Patnaik GK, Dhawan BN. Choleretic effect of picroliv, the hepatoprotective principle of *Picrorhiza kurroa*. *Planta Med* 1991; 57: 29-33.
- [10] Hu XP, Shin JW, Wang JH, Cho JH, Son JY, Cho CK, et al. Antioxidative and hepatoprotective effect of CGX, an herbal medicine, against toxic acute injury in mice. *J Ethnopharmacol* 2008; 120: 51-55.
- [11] Rechinger KH. Flora Iranica, Alliaceae. Graz: Akademische Druck u. Verlagsanstalt; 1984.
- [12] Kazemi S, Asgary S, Moshtaghian J, Rafieian M, Adelnia A, Shamsi F. Liver-protective effects of hydroalcoholic extract of *Allium hirtifolium* Boiss. In rats with alloxan-induced di-abetes mellitus. *ARYA Atheroscler* 2010; 6: 11-15.
- [13] Azadi HG, Riazi GH, Ghaffari SM, Ahmadian S, Khalife TJ. Effects of Allium hirtifolium (Iranian shallot) and its allicin on microtubule and cancer cell lines. Afr J Biotechnol 2009; 8: 5030-5037.
- [14] Rose P, Whiteman M, Moore PK, Zhu YZ. Bioactive S-alk(en)yl cysteine sulfoxide metabolites in the genus *Allium*: the chemistry of potential therapeutic agents. *Nat Prod Rep* 2005; **22**: 351-368.
- [15] Fattorusso E, Iorizzi M, Lanzotti V, Taglialatela-Scafati O. Chemical

- composition of shallot (*Allium ascalonicum* Hort.). *J Agric Food Chem* 2002; **50**: 5686-5690.
- [16] Asgari S, Setorki M, Rafieian-Kopaei M, Heidarian E, Shahinfard N, Ansari R, et al. Postprandial hypolipidemic and hypoglycemic effects of *Allium hertifolium* and *Sesamum indicum* on hypercholesterolemic rabbits. *Afr J Pharm Pharmacol* 2012; 6: 1131-1135.
- [17] Amin M, Pipelzadeh MH, Mehdinejad M, Rashidi I. An in vivo toxicological study upon shallomin, the active antimicrobial constitute of Persian shallot (Allium hirtifolium, Boiss) extract. Jundushapur J Nat Pharm Prod 2012; 7: 17-21.
- [18] Kooti W, Ghasemiboroon M, Asadi-Samani M, Ahangarpoor A, Abadi A, Noori M, et al. The effects of hydro-alcoholic extract of celery on lipid profile of rats fed a high fat diet. *Adv Environ Biol* 2014; 8(9): 325-330.
- [19] Rechinger KH, Lemond JM, Hedge IC. Flora Iranica (Umbelliferae).
  Graz: Akademische Druck u. Verlagsanstalt; 1994, p. 269-297.
- [20] Mansi K, Abushoffa AM, Disi A, Aburjai T. Hypolipidemic effects of seed extract of celery (*Apium graveolens*) in rats. *Pharmacogn Mag* 2009; 5: 301-305.
- [21] Asif HM, Akram M, Usmanghani K, Akhtar N, Shah PA, Uzair M, et al. Monograph of *Apium graveolens* Linn. *J Med Plants Res* 2011; 5: 1494-1496.
- [22] Nagella P, Ahmad A, Kim SJ, Chung IM. Chemical composition, antioxidant activity and larvicidal effects of essential oil from leaves of *Apium graveolens. Immunopharmacol Immunotoxicol* 2012; 34: 205-209.
- [23] Fazal SS, Singla RK. Review on the pharmacognostical and pharmacological characterization of *Apium graveolens* Linn. *Indo Glob J Pharm Sci* 2012; **2**: 36-42.
- [24] Sultana S, Ahmed S, Jahangir T, Sharma S. Inhibitory effect of celery seeds extract on chemically induced hepatocarcinogenesis: modulation of cell proliferation, metabolism and altered hepatic foci development. *Cancer Lett* 2005; 221: 11-20.
- [25] Ahmed B, Alam T, Varshney M, Khan SA. Hepatoprotective activity of two plants belonging to the Apiaceae and the Euphorbiaceae family. J Ethnopharmacol 2002; 79: 313-316.
- [26] Wu L, Chen Y. Protective effect of celery root against acute liver injury by CCl<sub>4</sub>. West China J Pharm Sci 2008; doi: 10.3969/j.issn.1006-0103.2008.04.013.
- [27] Mencherini T, Cau A, Bianco G, Della Loggia R, Aquino RP, Autore G. An extract of *Apium graveolens* var. dulce leaves: structure of the

- major constituent, apiin, and its anti-inflammatory properties. *J Pharm Pharmacol* 2007; **59**: 891-897.
- [28] Ramezani M, Nasri S, Yassa N. Antinociceptive and anti-inflammatory effects of isolated fractions from *Apium graveolens* seeds in mice. *Pharm Biol* 2009; 47: 740-743.
- [29] Gebhardt R. Antioxidative and protective properties of extracts from leaves of the artichoke (*Cynara scolymus* L.) against hydroperoxideinduced oxidative stress in cultured rat hepatocytes. *Toxicol Appl Pharmacol* 1997; 144: 279-286.
- [30] Vamanu E, Vamanu A, Nita S, Colceriu S. Antioxidant and antimicrobial activities of ethanol extracts of *Cynara scolymus* (*Cynarae folium*, Asteraceae family). *Trop J Pharm Res* 2011; **10**: 777-783.
- [31] Fallah Huseini H, Zareei Mahmoudabady A, Ziai SA, Mehrazma M, Alavian SM, Mehdizadeh M, et al. The effects of *Cynara scolymus* L. leaf and *Cichorium intybus* L. root extracts on carbon tetrachloride induced liver toxicity in rats. *J Med Plants* 2011; 10: 33-40.
- [32] Mehmetçik G, Ozdemirler G, Kocak-Toker N, Çevikbaş U, Uysal M. Effect of pretreatment with artichoke extract on carbon tetrachlorideinduced liver injury and oxidative stress. *Exp Toxicol Pathol* 2008; 60: 475-480.
- [33] Heidarian E, Jafari-Dehkordi E, Seidkhani-Nahal A. Lipid-lowering effect of artichoke on liver phosphatidate phosphohydrolase and plasma lipids in hyperlipidemic rats. *J Med Plants Res* 2011; 5: 4918-4924.
- [34] Păunescu A, Ponepal CM, Drăghici O, Marinescu AG. The CCl<sub>4</sub> action upon physiological indices in *Lepus timidus* and the protective role of some substances. *An UO Fasc Biol* 2009; 16: 104-107.
- [35] Ali ZY, Atia HA, Ibrahim NH. Possible hepatoprotective potential of Cynara scolymus, Cupressus sempervirens and Eugenia jambolana against paracetamol-induced liver injury: in-vitro and in-vivo evidence. Nat Sci 2012; 10: 75-86.
- [36] Parsaee H, Shafel MN, Boskabady MH. Effects of hydro-ethanolic extract of *Berberis vulgaris* fruit on rabbit isolated heart. *DARU J Pharm* Sci 2006; 14: 208-213.
- [37] Hadaruga DI, Hadaruga NG, Bandur GN, Rivis A, Costescu C, Ordodi VL, et al. *Berberis vulgaris* extract/β cyclodextrin nanoparticles synthesis and characterization. *Rev Chim (Bucharest)* 2010; **61**: 669-675.
- [38] Fatehi M, Saleh TM, Fatehi-Hassanabad Z, Farrokhfal K, Jafarzadeh M, Davodi S. A pharmacological study on *Berberis vulgaris* fruit extract. *J Ethnopharmacol* 2005; 102: 46-52.
- [39] Imanshahidi M, Hosseinzadeh H. Pharmacological and therapeutic effects of *Berberis vulgaris* and its active constituent, berberine.

- Phytother Res 2008; 22: 999-1012.
- [40] Feng Y, Siu KY, Ye X, Wang N, Yuen MF, Leung CH, et al. Hepatoprotective effects of berberine on carbon tetrachloride-induced acute hepatotoxicity in rats. *Chin Med* 2010; **5**: 33.
- [41] Domitrović R, Jakovac H, Blagojević G. Hepatoprotective activity of berberine is mediated by inhibition of TNF-α, COX-2, and iNOS expression in CCl<sub>4</sub>-intoxicated mice. *Toxicology* 2011; **280**: 33-43.
- [42] Hermenean A, Popescu C, Ardelean A, Stan M, Hadaruga N, Mihali CV, et al. Hepatoprotective effects of *Berberis vulgaris* L. extract/β cyclodextrin on carbon tetrachloride–induced acute toxicity in mice. *Int J Mol Sci* 2012; 13: 9014-9034.
- [43] Hamburger M, Adler S, Baumann D, Förg A, Weinreich B. Preparative purification of the major anti-inflammatory triterpenoid esters from Marigold (*Calendula officinalis*). *Fitoterapia* 2003; 74: 328-338.
- [44] Preethi KC, Kuttan G, Kuttan R. Antioxidant potential of an extract of Calendula officinalis flowers in vitro and in vivo. Pharm Biol 2006; 44: 691-697.
- [45] Okoh OO, Sadimenko AP, Asekun OT, Afolayan AJ. The effects of drying on the chemical components of essential oils of *Calendula officinalis* L. *Afr J Biotechnol* 2008; 7(10): 1500-1502.
- [46] Singh MK, Sahu P, Nagori K, Dewangan D, Kumar T, Alexander A, et al. Organoleptic properties in-vitro and in-vivo pharmacological activities of Calendula officinalis Linn: an over review. J Chem Pharm Res 2011; 3: 655-663.
- [47] Khan MU, Rohilla A, Bhatt D, Afrin S, Rohilla S, Ansari SH. Diverse belongings of *Calendula officinalis*: an overview. *Int J Pharm Sci Drug Res* 2011; 3: 173-177.
- [48] Preethi KC, Kuttan R. Hepato and reno protective action of *Calendula officinalis* L. flower extract. *Indian J Exp Biol* 2009; **47**: 163-168.
- [49] Mishra AK, Mishra A, Chattopadhyay P. Calendula officinalis: an important herb with valuable theraputic dimensions-an overview. J Glob Pharm Technol 2010; 2: 14-23.
- [50] Braga PC, Dal Sasso M, Culici M, Spallino A, Falchi M, Bertelli A, et al. Antioxidant activity of *Calendula officinalis* extract: inhibitory effects on chemiluminescence of human neutrophil bursts and electron paramagnetic resonance spectroscopy. *Pharmacology* 2009; 83: 348-355.
- [51] Al-Ghamdi MS. The anti-inflammatory, analgesic and antipyretic activity of *Nigella sativa*. *J Ethnopharmacol* 2001; **76**: 45-48.
- [52] Kanter M, Coskun O, Budancamanak M. Hepatoprotective effects of

- Nigella sativa L and Urtica dioica L on lipid peroxidation, antioxidant enzyme systems and liver enzymes in carbon tetrachloride-treated rats. World J Gastroenterol 2005; 11: 6684-6688.
- [53] Yildiz F, Coban S, Terzi A, Ates M, Aksoy N, Cakir H, et al. *Nigella sativa* relieves the deleterious effects of ischemia reperfusion injury on liver. *World J Gastroenterol* 2008; 14: 5204-5209.
- [54] Alemi M, Sabouni F, Sanjarian F, Haghbeen K, Ansari S. Antiinflammatory effect of seeds and callus of *Nigella sativa* L. extracts on mix glial cells with regard to their thymoquinone content. *AAPS PharmSciTech* 2013; 14: 160-167.
- [55] Paarakh PM. Nigella sativa Linn.-A comprehensive review. Indian J Nat Prod Resour 2010; 1: 409-429.
- [56] Houcher Z, Boudiaf K, Benboubetra M, Houcher B. Effects of methanolic extract and commercial oil of *Nigella sativa* L. on blood glucose and antioxidant capacity in alloxan-induced diabetic rats. *Pteridines* 2007; 18: 8-18.
- [57] Molina-Montenegro MA, Atala C, Gianoli E. Phenotypic plasticity and performance of *Taraxacum officinale* (dandelion) in habitats of contrasting environmental heterogeneity. *Biol Invasions* 2010; 12: 2277-2284.
- [58] Tabassum N, Shah MY, Qazi MA, Shah A. Prophylactic activity of extract of *Taraxacum officiale* Weber. against hepatocellular injury induced in mice. *Pharmacologyonline* 2010; 2: 344-352.
- [59] Choi UK, Lee OH, Yim JH, Cho CW, Rhee YK, Lim SI, et al. Hypolipidemic and antioxidant effects of dandelion (*Taraxacum officinale*) root and leaf on cholesterol-fed rabbits. *Int J Mol Sci* 2010; 11: 67-78.
- [60] Das SK, Mukherjee S. Biochemical and immunological basis of silymarin effect, a milk thistle (*Silybum marianum*) against ethanolinduced oxidative damage. *Toxicol Mech Methods* 2012; 22(5): 409-413.
- [61] Fallah H, Zarrei M, Ziai M, Mehrazma M, Alavian SM, Kianbakht S, et al. The effects of *Taraxacum officinale* L. and *Berberis vulgaris* L. root extracts on carbon tetrachloride induced liver toxicity in rats. *J Med Plants* 2010; 9: 45-52.
- [62] Mahesh A, Jeyachandran R, Cindrella L, Thangadurai D, Veerapur V, Muralidhara Rao D. Hepatocurative potential of sesquiterpene lactones of *Taraxacum officinale* on carbon tetrachloride induced liver toxicity in mice. *Acta Biol Hung* 2010; 61: 175-190.
- [63] Mroueh M, Daher C, El Sibai M, Tenkerian C. Antioxidant and hepatoprotective activity of *Tragopogon porrifolius* methanolic extract. *Planta Med* 2011; doi: 10.1055/s-0031-1282460.

- [64] Formisano C, Rigano D, Senatore F, Bruno M, Rosselli S. Volatile constituents of the aerial parts of white salsify (*Tragopogon porrifolius* L., Asteraceae). *Nat Prod Res* 2010; 24: 663-668.
- [65] Konopiński M. Influence of intercrop plants and varied tillage on yields and nutritional value of salsify (*Tragopogon porrifolius L.*) roots. *Acta Sci Pol Hortorum Cultus* 2009; 8: 27-36.
- [66] Govind P. Medicinal plants against liver diseases. *Int Res J Pharm* 2011; **2**: 115-121.
- [67] Zeeni N, Daher CF, Saab L, Mroueh M. Tragopogon porrifolius improves serum lipid profile and increases short-term satiety in rats. Appetite 2014; 72: 1-7.
- [68] Massumi MA, Fazeli MR, Alavi SHR, Ajani Y. Chemical constituents and antibacterial activity of essential oil of *Prangos ferulacea* (L.) Lindl. fruits. *Iran J Pharm Sci* 2007; 3: 171-176.
- [69] Akhgar MR, Pahlavanzadeh-Iran S, Lotfi-Anari P, Faghihi-Zarandi A. Composition of essential oils of fruits and leaves of *Prangos ferulacea* (L.) Lindl. growing wild in Iran. *Trends Mod Chem* 2011; 1: 1-4.
- [70] Kafash-Farkhad N, Asadi-Samani M, Rafieian-Kopaei M. A review on phytochemistry and pharmacological effects of *Prangos ferulacea* (L.) Lindl. *Life Sci J* 2013; 10: 360-367.
- [71] Kafash Farkhad N, Farokhi F, Tukmacki A, Soltani Band K. Hydroalcoholic extract of the root of *Prangos ferulacea* (L.) Lindl can improve serum glucose and lipids in alloxan-induced diabetic rats. *Avicenna J Phytomed* 2012; 2: 179-187.
- [72] Amagase H, Petesch BL, Matsuura H, Kasuga S, Itakura Y. Intake of garlic and its bioactive components. *J Nutr* 2001; **131**: 955S-962S.
- [73] Touloupakis E, Ghanotakis DF. Nutraceutical use of garlic sulfurcontaining compounds. Adv Exp Med Biol 2010; 698: 110-121.
- [74] Obioha UE, Suru SM, Ola-Mudathir KF, Faremi TY. Hepatoprotective potentials of onion and garlic extracts on cadmium-induced oxidative damage in rats. *Biol Trace Elem Res* 2009; 129: 143-156.
- [75] Ghorbel H, Feki I, Friha I, Khabir AM, Boudawara T, Boudawara M, et al. Biochemical and histological liver changes occurred after iron supplementation and possible remediation by garlic consumption. *Endocrine* 2011; 40: 462-471.
- [76] Patrick-Iwuanyanwu KC, Wegwu MO, Ayalogu EO. Prevention of CCl<sub>4</sub>-induced liver damage by ginger, garlic and vitamin E. *Pak J Biol Sci* 2007; 10: 617-621.
- [77] El-Beshbishy HA. Aqueous garlic extract attenuates hepatitis and oxidative stress induced by galactosamine/lipoploysaccharide in rats.

- Phytother Res 2008; 22: 1372-1379.
- [78] Park EY, Ki SH, Ko MS, Kim CW, Lee MH, Lee YS, et al. Garlic oil and DDB, comprised in a pharmaceutical composition for the treatment of patients with viral hepatitis, prevents acute liver injuries potentiated by glutathione deficiency in rats. *Chem Biol Interact* 2005; 155: 82-96.
- [79] Taji F, Shirzad H, Ashrafi K, Parvin N, Kheiri S, Namjoo A, et al. A comparison between the antioxidant strength of the fresh and stale Allium sativum (garlic) extracts. Zahedan J Res Med Sci 2012; 14: 25-29.
- [80] Shirzad H, Taji F, Rafieian-Kopaei M. Correlation between antioxidant activity of garlic extracts and WEHI-164 fibrosarcoma tumor growth in BALB/c mice. *J Med Food* 2011; 14: 969-974.
- [81] Sahpaz S, Garbacki N, Tits M, Bailleul F. Isolation and pharmacological activity of phenylpropanoid esters from *Marrubium vulgare*. *J Ethnopharmacol* 2002; **79**: 389-392.
- [82] Elberry AA, Harraz FM, Ghareib SA, Nagy AA, Gabr SA, Suliaman MI, et al. Antihepatotoxic effect of *Marrubium vulgare* and *Withania somnifera* extracts on carbon tetrachloride-induced hepatotoxicity in rats. *J Basic Clin Pharm* 2010; 1: 247-254.
- [83] Mubashir HM, Bahar A, Suroor AK, Shah MY, Shamshir K. Antihepatotoxic activity of aqueous extract of *Marrubium vulgare* whole plant in CCl<sub>4</sub> induced toxicity. *Indian J Nat Prod* 2009; 25: 3-8.
- [84] Selim YA, Ouf NH. Anti-inflammatory new coumarin from the *Ammi majus* L. *Org Med Chem Lett* 2012; **2**: 1.
- [85] Hübner S, Hehmann M, Schreiner S, Martens S, Lukacin R, Matern U. Functional expression of cinnamate 4-hydroxylase from *Ammi majus* L. *Phytochemistry* 2003; 64: 445-452.
- [86] Mutlag SH, Ismael DK, Al-Shawi NN. Study the possible hepatoprotective effect of different doses of *Ammi majus* seeds'extract against CCl<sub>4</sub> induced liver damage in rats. *Pharm Glob (IJCP)* 2011; 9: 1-5.
- [87] Hassan HA, Yousef MI. Ameliorating effect of chicory (*Cichorium intybus* L.)-supplemented diet against nitrosamine precursors-induced liver injury and oxidative stress in male rats. *Food Chem Toxicol* 2010; 48: 2163-2169.
- [88] Georgina EO, Kingsley O, Esosa US, Helen NK, Frank AO, Anthony OC. Comparative evaluation of antioxidant effects of watermelon and orange, and their effects on some serum lipid profile of Wister albino rats. *Int J Nutr Metab* 2011; 3: 97-102.
- [89] Bartalis J. Hepatoprotective activity of cucurbitacin [dissertation]. Brookings: South Dakota State University; 2005.

- [90] Altas S, Kizil G, Kizil M, Ketani A, Haris PI. Protective effect of DiyarbakIr watermelon juice on carbon tetrachloride-induced toxicity in rats. Food Chem Toxicol 2011; 49: 2433-2438.
- [91] Kline GJ, Sorensen PD. A revision of *Agrimonia* (Rosaceae) in North and Central America. *Brittonia* 2008; **60**: 11-33.
- [92] Correia HS, Batista MT, Dinis TC. The activity of an extract and fraction of *Agrimonia eupatoria* L. against reactive species. *Biofactors* 2007; 29: 91-104.
- [93] Gião MS, Pereira CI, Fonseca SC, Pintado ME, Malcata FX. Effect of particle size upon the extent of extraction of antioxidant power from the plants Agrimonia eupatoria, Salvia sp. and Satureja montana. Food Chem 2009; 117: 412-416.
- [94] Yoon SJ, Koh EJ, Kim CS, Jee OP, Kwak JH, Jeong WJ, et al. Agrimonia eupatoria protects against chronic ethanol-induced liver injury in rats. Food Chem Toxicol 2012; 50: 2335–2341.
- [95] Kang S, Lee C, Koo H, Ahn D, Choi H, Lee J, et al. Hepatoprotective effects of aqueous extract from aerial part of agrimmony. *Korean J Pharmacogn* 2006; 37: 28-32.
- [96] Kshirsagar AD, Mohite R, Aggrawal AS, Suralkar UR. Hepatoprotective medicinal plants of ayurveda— a review. Asian J Pharm Clin Res 2011; 4: 1-8.
- [97] Rashid F, Ahmed R, Mahmood A, Ahmad Z, Bibi N, Kazmi SU. Flavonoid glycosides from *Prunus armeniaca* and the antibacterial activity of a crude extract. *Arch Pharm Res* 2007; **30**: 932-937.
- [98] Abdel-Rahman MK. Can apricot kernels fatty acids delay the atrophied hepatocytes from progression to fibrosis in dimethylnitrosamine (DMN)-induced liver injury in rats? *Lipids Health Dis* 2011; 10: 114.
- [99] Shivashankara AR, Azmidah A, Haniadka R, Rai MP, Arora R, Baliga MS. Dietary agents in the prevention of alcohol-induced hepatotoxicty: preclinical observations. *Food Funct* 2012; 3: 101-109.
- [100]Ahmed T, Sadia H, Khalid A, Batool S, Janjua A. Report: prunes and liver function: a clinical trial. *Pak J Pharma Sci* 2010; **23**: 463-466.
- [101]Yurt B, Celik I. Hepatoprotective effect and antioxidant role of sun, sulphited-dried apricot (*Prunus armeniaca* L.) and its kernel against ethanol-induced oxidative stress in rats. *Food Chem Toxicol* 2011; 49: 508-513.
- [102]Ozturk F, Gul M, Ates B, Ozturk IC, Cetin A, Vardi N, et al. Protective effect of apricot (*Prunus armeniaca* L.) on hepatic steatosis and damage induced by carbon tetrachloride in Wistar rats. *Br J Nutr* 2009; 102: 1767-1775.