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The antimicrobial susceptibility profile of ESKAPE pathogens from urinary tract infections in a referral laboratory, Northeast Iran

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ABSTRACT

Objective: To assess the antimicrobial susceptibility pattern of ESKAPE pathogens from Neyshabur, Iran during 2013–2015.

Methods: A total of 345 isolates including 62 *Staphylococcus aureus* (*S. aureus*), 38 *Enterobacter* spp. (including 14 *Enterobacter agglomerans*, 6 *Enterobacter aerogenes* and other 18 *Enterobacter* spp.), 123 *Enterococcus faecium*, 78 *Klebsiella pneumonia*, 10 *Pseudomonas aeruginosa* and 34 *Acinetobacter baumannii* were isolated. The antimicrobial susceptibility pattern of isolates was conducted with Kirby Bauer method. Data were analyzed with SPSS 20.0 software using *F*- and *t*-tests.

Results: Among *S. aureus* isolates, the highest resistance was observed against nalidixic acid (81.35%) and cefixime (74.50%). Thirty-three (53.22%) *S. aureus* isolates were cefoxitin resistant (methicillin-resistant *S. aureus*). The majority of *Enterobacter* species was resistant to amikacin (100.00%) and cephalotin (66.60%). Most *Enterococcus faecium* isolates were resistant to nalidixic acid (89.43%) and amikacin (83.33%), but vancomycin-resistant enterococci isolates were not detected. Moreover, among *Klebsiella pneumonia*, the highest resistance was observed to nalidixic acid (20.98%) and cotrimoxazole (28.39%). Furthermore, all *Pseudomonas aeruginosa* isolates were resistant to cefotaxime (100.00%) and majority to nitroforantoin (88.80%). *Acinetobacter baumannii* isolates showed the highest and the lowest resistance to cefotaxime (100.00%) and cefixime (88.71%), respectively.

Conclusions: The prevalence of ESKAPE pathogens from northeast region was low, but majority of them exhibited high rate of antibiotic resistance to common used antimicrobial agents.

1. Introduction

ESKAPE pathogens, isolated from hospital and community settings, have been recently categorized because of new paradigms in pathogenesis and transmission and overcoming to drugs effectiveness. It has been revealed that ESKAPE pathogens are involved in nearly 41% of infections in patients in intensive care units[1]. Resistance levels are substantial in these pathogens, ranging from resistance that approximately completely excludes an antibiotic from empirical therapeutic insights [e.g. Enterococcus faecium (E. faecium) with resistance to ampicillin or vancomycin] to resistance that shows the potential to change choices of both empirical and definitive antimicrobial therapies [e.g. Acinetobacter baumannii (A. baumannii) or Pseudomonas aeruginosa (P. aeruginosa) with resistance to carbapenems][2,3]. However, the rates of resistance vary in different areas. Staphylococcus aureus (S. aureus) with resistance to methicillin [methicillin-resistant S. aureus (MRSA)] and intermediate level vancomycin (vancomycin-intermediate S. aureus strains) are examples of drug resistant S. aureus from hospital or community settings[4,5]. Emergence of extended spectrum beta-lactamases (ESBL) and carbapenemase enzymes among Gram-negative species have been developed worldwide[6,7]. P. aeruginosa has long been the "holy grail" target for antimicrobial development. The importance of P. aeruginosa in causing deaths of patients with febrile neutropenia and its intrinsic resistance to many early antimicrobial agents led to concerted efforts to find new antibiotics with activity against this species[8,9]. Until now,

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there has been no previous report of ESKAPE prevalence and their antimicrobial susceptibility profile collectively in one study in Iran. The aim of this study was to determine prevalence and antimicrobial susceptibility pattern of ESKAPE isolates from Neyshabur, Northeast Iran during 2013–2015.

2. Materials and methods

2.1. Study sample and design

During the years 2013–2015, a total of 1661 patients (age ranged from 7 months to 74 years old) with urinary tract complications were referred to the referral laboratory of Neyshabur. The midstream urine of them was collected and cultured. The bacterial species were identified by using conventional biochemical tests. Among them, 345 ESKAPE pathogens including 62 *S. aureus*, 18 *Enterobacter* spp., 14 *Enterobacter agglomerans* (*E. aglomerans*), 6 *Enterobacter aerogenes* (*E. aerogenes*), 123 *E. faecium*, 78 *Klebsiella pneumonia* (*K. pneumonia*), 10 *P. aeruginosa* and 34 *A. baumannii* were isolated.

2.2. Antimicrobial susceptibility testing

The antimicrobial susceptibility testing of isolates was conducted with Kirby Bauer method, and following Clinical and Laboratory Standards Institute advice, version 2013[10]. *E. coli* ATCC 25922 and *S. aureus* ATCC 25923 standard strains were used as quality control for the disks. The disks included: nalidixic acid, nitrofurantoin, chloramphenicol, ciprofloxacin, cefoxitin, ceftizoxime, ceftazidime, cotrimoxazol, gentamicin, cefalotin, amikacin, cefotaxime and erythromycin (Padtan Teb). Ceftazidime and cefoxitin resistance was considered as ESBL producers and MRSA strains, respectively.

2.3. Ethical approval

The study protocol was performed according to the Clinical And Laboratory Standards Institute declaration and approved by Academic Center for Education, Culture and Research. Informed written consent was obtained from Academic Center for Education, Culture and Research committee.

2.4. Data analysis

Data were analyzed with SPSS 20.0 software using *F*- and *t*-tests. Any P < 0.05 [95% confidence interval (*CI*)] was considered as significant difference.

3. Results

The prevalence of ESKAPE pathogens was 20.77%, including 3.73% S. aureus, 2.29% Enterobacter spp., 7.40% E. faecium, 4.69% K. pneumonia, 0.60% P. aeruginosa and 2.04% A. baumannii. The age average of patients was (33.10 ± 24.16) years. All these pathogens isolated from female patients were more significantly than those from male patients (Table 1). Other organisms other than ESKAPE cases included: 1180 E. coli, 5 Citrobacter diversus, 5 Citrobacter freundii, 1 Streptococcus pyogenes, 69 Proteus mirabilis, 1 Proteus vulgaris, 1 Serratia marssecence, 23 Staphylococcus epidermidis and 32 Staphylococcus saprophyticus. The number and percentage of ESKAPE isolates are depicted in Table 1. The antibiotic susceptibility profile of ESKAPE pathogens is exhibited in Tables 2 and 3. It was shown that 53.22% of S. aureus isolates were resistant to cefoxitin (indicating MRSA). Moreover, the rate of ceftazidime resistance among Enterobacter spp., E. faecium, K. pneumonia, P. aeruginosa and A. baumannii was 20.00%, 30.00%, 40.00%, and 81.00%, respectively. In addition, 426/1180 (36.10%) E. coli, 1 (20%) Citrobacter diversus,

3 (4.34%) *Proteus mirabilis* were ceftazidime resistant, but all *Serratia* spp. were susceptible to it.

Table 1

The distribution of ESKAPE isolates among female and male patie	ents.
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Genus	Species	Number	Male	Female	P value
			(%)	(%)	(95% CI)
Enterobacter	Enterobacter spp.	18	39.47	50.53	0.0016
	E. aerogenes	6			
	E. agglomerans	14			
Entroccoci	E. faecium	123	16.26	82.74	< 0.0001
Klebsiella	K. pneumonia	78	16.25	82.75	< 0.0001
	Klebsiella ozaenae	2			
Pseudomonas	P. aeruginosa	10	30.00	70.00	0.0008
Staphylococcus	S. aureus	62	9.67	90.23	< 0.0001
Acinetobacter	A. baumannii	34	26.47	72.53	< 0.0001

Table 2

The antibiotic resistance (%) profile of ESKAPE pathogens

Disks/isolates	Enterobacter	Е.	К.	Р.	<i>S</i> .	А.
	spp.	faecium	pneumonia	aeruginosa	aureus	baumannii
Cefixime	30.30	56.38	0.00	85.71	74.50	88.71
Nalidixic acid	42.85	89.43	20.98	60.00	81.35	66.00
Nitrofurantoin	33.00	4.88	13.58	88.80	0.00	89.00
Chloramphenicol	16.66	15.25	19.60	10.00	34.00	10.00
Ciprofloxacin	16.20	12.39	8.64	10.00	9.30	40.00
Ceftizoxim	8.10	1.00	4.94	80.00	1.63	80.00
Cotrimoxazole	40.00	61.98	28.39	60.00	16.07	63.00
Gentamicin	21.20	83.00	12.85	12.50	2.17	16.50
Cefalotin	66.60	100.00	40.00	100.00	0.00	100.00
Amikacin	100.00	83.33	11.00	12.00	3.78	14.00
Cefotaxime	21.43	22.06	28.20	92.00	23.52	100.00
^a Ceftazidime	20.00	-	30.00	40.00	-	81.00
Cefoxitin	80.00	60.00	90.00	100.00	53.22 ^b	93.00
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^a: Supposed possible ESBL producing isolates; ^b: MRSA strains.

Table 3

The sensitivity level (%) of ESKAPE isolates.

Disks/isolates	Enterobacter	Е.	К.	Р.	S.	А.
	spp.	faecium	pneumonia	aeruginosa	aureus	baumannii
Cefixime	75.8	38.0	52.0	0.0	11.0	2.0
Nalidixic acid	79.3	12.0	64.0	2.0	6.0	2.0
Nitrofurantoin	40.0	64.0	65.0	1.0	61.0	0.0
Chloramphenicol	69.0	49.0	41.0	54.0	31.0	42.0
Ciprofloxacin	83.8	40.0	73.0	46.0	54.0	45.0
Ceftizoxim	91.9	93.0	71.0	4.0	45.0	3.0
Cotrimoxazole	60.0	42.0	58.0	20.0	47.0	20.0
Gentamicin	88.0	12.0	59.0	40.0	46.0	32.0
Cefalotin	20.0	0.0	3.0	0.0	90.0	0.0
Amikacin	0.0	10.5	34.0	36.0	38.0	27.0
Cefotaxime	70.0	52.0	28.0	0.0	12.0	0.0
Ceftazidime	71.0	-	50.0	12.0	_	2.0
Cefoxitin	10.0	40.0	4.0	2.0	30.0	0.0

4. Discussion

Knowledge of local antibiotic resistance trends among urinary isolates is crucial not only in guiding clinicians to prescribe appropriate antibiotics but also for observations based recommendations for empirical antibiotic treatment of urinary tract infection[11]. The current study assesses the antimicrobial resistance rates among ESKAPE isolates and possible detection of ESBL production and MRSA among urinary isolates in Neyshabur, Iran. In the current study, the age average of patients was (33.1 ± 24.16) years. Moreover, the age range of patients was 7 months to 74 years old. In this study, the history of patients regarding prior antibiotic consumption, hospitalization, catheter, smoking and contact with hospital settings was not elucidated. Thus we could not determine any relation between most of possible risk factors and presence of ESKAPE isolates.

The prevalence of ESKAPE pathogens from midstream urine was 20.77%, including 3.73% S. aureus, 2.29% Enterobacter spp., 7.40% E. faecium, 4.69% K. pneumonia, 0.60% P. aeruginosa and 2.04% A. baumannii. In this study, E. faecium were the most common isolates from urinary tract infections. Enterococci have been reported from urinary tract infections and aminopenicillins may be used as a resort for vancomycin-resistant enterococci[12]. A study showed that the midstream urine of patients contained 1% P. aeruginosa and K. pneumonia and 18% enterococci, but no other ESKAPE isolates were determined[13]. It was shown that 53.22% of S. aureus isolates were resistant to cefoxitin (indicating MRSA). Sasirekha demonstrated that among 325 clinical isolates from urinary tract infection, the prevalence of MRSA was 27.5% and ESBL positive Gram-negative bacteria were 48.9%[14]. In this study, the rate of ceftazidime resistance among Enterobacter spp., K. pneumonia, P. aeruginosa and A. baumannii was 20.00%, 30.00%, 40.00% and 81.00%, respectively. It was proposed that ceftazidime resistance is a potential for ESBL production.

Antibiotic resistant pathogens have caused rising in morbidity and mortality and increase of economic costs and hospitalization. Vancomycin-resistant enterococci, MRSA and ESBL producer Gramnegative bacteria confer these conditions as reported previously[6,15-17].

Among the oral antibiotics, nitrofurantoin commonly prescribed for urinary tract infections in most of countries showed a high resistance rate against *A. baumannii* (89.00%) and *P. aeruginosa* (88.80%) in our study which highlighted an increased resistance among these organisms in this area, while it was more effective against *K. pneumonia* and *Enterobacter* spp., which are consistent with those of the previous studies[18-22]. A low degree of resistance to gentamicin and amikacin was observed for *K. pneumonia*, *P. aeruginosa*, *S. aureus*, and *A. baumannii*, thus may be helpful in combating severe infections. In our study, carbapenems were not evaluated.

The current study, which examines the prevalence of ESKAPE uropathogens and spread of drug resistance among them, is possibly first of its kind as it is not restricted to a specific pathogen and health care center. On the basis of these findings, it is suggested that urine culture and antimicrobial susceptibility testing for urinary tract infections. Continuous analysis of the antibiotic resistance profile acts as a guide in initiating the empirical treatment of urinary tract infections. Development of local surveillance programs is necessary to provide information which would then enable the development of specific urinary tract infection guidelines.

Conflict of interest statement

We declare that we have no conflict of interest.

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References

- Pendleton JN, Gorman SP, Gilmore BF. Clinical relevance of the ESKAPE pathogens. *Expert Rev Anti Infect Ther* 2013; 11(3): 297-308.
- [2] Rice LB. Progress and challenges in implementing the research on ESKAPE pathogens. *Infect Control Hosp Epidemiol* 2010; **31**(Suppl 1): S7-10.
- [3] Stosor V, Peterson LR, Postelnick M, Noskin GA. *Enterococcus faecium* bacteremia: does vancomycin resistance make a difference? *Arch Intern Med* 1998; **158**(5): 522-7.
- [4] Ghasemian A, Najar Peerayeh S, Bakhshi B, Mirzaee M. Comparison of

biofilm formation between methicillin-resistant and methicillin-susceptible isolates of *Staphylococcus aureus*. *Iran Biomed J* 2016; **20**(3): 175-81.

- [5] Mirzaee M, Najar-Peerayeh S, Behmanesh M, Moghadam MF. Relationship between adhesin genes and biofilm formation in vancomycinintermediate *Staphylococcus aureus* clinical isolates. *Curr Microbiol* 2015; **70**(5): 665-70.
- [6] Davodian E, Sadeghifard N, Ghasemian A, Noorbakhsh S. Molecular detection of *bla*_{VEB-1} beta-lactamase encoding gene among extended spectrum B-lactamase positive wound isolates of *Pseudomonas aeruginosa*. *Arch Pediatr Infect Dis* 2015; **3**(4): e26362.
- [7] Davodian E, Sadeghifard N, Ghasemian A, Noorbakhsh S. Presence of bla_{PER-1} and bla_{VEB-1} beta-lactamase genes among isolates of *Pseudomonas* aeruginosa from south west of Iran. J Epidemiol Glob Health 2016; 6(3): 211-3.
- [8] Taccetti G, Sly PD. Early detection of infection with *Pseudomonas aeruginosa* in cystic fibrosis: the Holy Grail or an achievable goal? *J Cyst Fibros* 2014; 13(5): 491-3.
- [9] Saderi H, Owlia P. Detection of multidrug resistant (MDR) and extremely drug resistant (XDR) *P. aeruginosa* isolated from patients in Tehran, Iran. *Iran J Pathol* 2015; 10(4): 265-71.
- [10] Wolfensberger A, Sax H, Weber R, Zbinden R, Kuster SP, Hombach M. Change of antibiotic susceptibility testing guidelines from CLSI to EUCAST: influence on cumulative hospital antibiograms. *PloS One* 2013; 8(11): e79130.
- [11] Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. *Nat Rev Microbiol* 2015; **13**(5): 269-84.
- [12] Cole KA, Kenney RM, Perri MB, Dumkow LE, Samuel LP, Zervos MJ, et al. Outcomes of aminopenicillin therapy for vancomycin-resistant enterococcal urinary tract infections. *Antimicrob Agents Chemother* 2015; 59(12): 7362-6.
- [13] Hooton TM, Roberts PL, Cox ME, Stapleton AE. Voided midstream urine culture and acute cystitis in premenopausal women. *N Engl J Med* 2013; 369(20): 1883-91.
- [14] Sasirekha B. Prevalence of ESBL, AmpC β-lactamases and MRSA among uropathogens and its antibiogram. EXCLI J 2013; 12: 81-8.
- [15] Engemann JJ, Carmeli Y, Cosgrove SE, Fowler VG, Bronstein MZ, Trivette SL, et al. Adverse clinical and economic outcomes attributable to methicillin resistance among patients with *Staphylococcus aureus* surgical site infection. *Clin Infect Dis* 2003; **36**(5): 592-8.
- [16] Moosavian M, Shoja S, Rostami S, Torabipour M, Farshadzadeh Z. Inducible clindamycin resistance in clinical isolates of *Staphylococcus aureus*. Arch Clin Infect Dis 2014; 9(2): 421-7.
- [17] Ghasemian A, Mirzaee M. Methicillin resistant *Staphylococcus aureus* (MRSA) strains and the staphylococcal cassette chromosome *mec* types in Iran. *Infect Epidemiol Med* 2016; 2(3): 31-4.
- [18] Lei J, Han S, Wu W, Wang X, Xu J, Han L. Extensively drug-resistant Acinetobacter baumannii outbreak cross-transmitted in an intensive care unit and respiratory intensive care unit. Am J Infect Control 2016; doi: 10.1016/j.ajic.2016.03.041.
- [19] Aminzadeh Z, Yadegarynia D, Fatemi A, Armaki SA, Aslanbeygi B. Prevalence and antimicrobial susceptibility pattern of extended spectrum beta lactamase (ESBL) and non-ESBL producing enteric Gram-negative bacteria and activity of nitrofurantoin in the era of ESBL. *Jundishapur J Microbiol* 2013; 6(7): 1-6.
- [20] Giske GG. Contemporary resistance trends and mechanisms for the old antibiotics colistin, temocillin, fosfomycin, mecillinam and nitrofurantoin. *Clin Microbiol Infect* 2015; **21**(10): 899-905.
- [21] Zykov IN, Sundsfjord A, Småbrekke L, Samuelsen Ø. The antimicrobial activity of mecillinam, nitrofurantoin, temocillin and fosfomycin and comparative analysis of resistance patterns in a nationwide collection of ESBL-producing *Escherichia coli* in Norway 2010–2011. *Infect Dis (Lond)* 2016; 48(2): 99-107.
- [22] Khameneh ZR, Afshar AT. Antimicrobial susceptibility pattern of urinary tract pathogens. *Saudi J Kidney Dis Transpl* 2009; 20(2): 251-3.