

CODEN (USA): IAJPBB

ISSN: 2349-7750

INDO AMERICAN JOURNAL OF

PHARMACEUTICAL SCIENCES

Available online at: http://www.iajps.com

Research Article

FT-IR STUDIES OF ETHANOLIC EXTRACT OF SARACA ASOKA

Asha Jyothi V*1 and Dr. Satyavati D2

¹ Department of Pharmacology Shadan Womens College of Pharmacy, Khairatbad, Hyderabad.

²Brilliant college of Pharmacy, Koheda, Near Ramoji film city, Hyderabad.

Abstract:

In the present study an attempt has been made to establish FT-IR profile and identify the functional components of Saraca asoka. FTIR method was performed on a Thermo Scientific Spectrophotometer system which was used to detect the characteristic peak values and their functional groups. The results of Saraca asoka barks FTIR analysis confirmed the presence of N-H stretching bond, alkenes group, aldehyde group, and carboxylic group which shows major peaks at 3421-3398 cm^{-1,} 3010cm^{-1,} 2926-2894 cm^{-1,} and 1697 cm⁻¹ respectively. The results of the present study produced the FTIR spectrum profile for the medicinally important plant Saraca asoka.

Keywords: Saraca asoka, FTIR, Spectroscopy, Functional groups and Phytoestrogens.

Corresponding author:

Asha Jyothi V,

Department of Pharmacology, Shadan Womens College of Pharmacy, Khairatbad, Hyderabad.

E mail: ashajyothivadlapudi@gmail.com



Please cite this article in press as Asha Jyoti and Satyavati, **FT-IR Studies of Ethanolic Extract of Saraca Asoka**, Indo Am. J. Pharm. Sci, 2016; 3(2).

INTRODUCTION:

Fourier transform infrared spectrometry is a physicochemical analytical technique that does not resolve the concentrations of individual metabolites but provides a snapshot of the metabolic composition of a tissue at a given time¹. FTIR can be employed to determine the structure of unknown composition and the intensity of the absorption spectra associated with molecular composition or content of the chemical group [1, 2]. The FT-IR method measures the vibrations of bonds within chemical functional groups and generates a spectrum that can be regarded as a biochemical or metabolic "fingerprint" of the sample. By attaining IR spectra from plant samples, it might possible to detect the minor changes of primary and secondary metabolites [2,3]. At present, particularly in phytochemistry, FT-IR has been exercised to identify the concrete structure of certain plant secondary metabolites [4-6]. But, on pharmacognosy front FT-IR is still a novel tool to characterize and identify the commercial components from the adulterant. FT-IR method has been successfully utilized in the characterization of bacterial, fungal and plant species [7-19]. FT-IR is one of the most widely used methods to identify the chemical constituents and elucidate the compounds structures, and has been used as a requisite method to identify medicines in Pharmacopoeia of many countries [20]. At the pharmacological front FT-IR is helpful tool to identify the compounds responsible for respective pharmacological activities.

Saraca L. is a genus in the family Fabaceae (legume family) of about eventy plant species of tree native to the lands rom India, Chinaand Ceylon to Malaysia and Celebes . The trees are grown in warm humid climates, and prefer a moist well-drained soil with plenty of organic matter. Typically, these trees are accustomed to the shade of other trees. Most species particular of Saraca are trees characteristic of streams. The species Saraca asoca is believed to be the tree under which Buddha was born. In ancient times this plant was used menstrual irregularities. Red saraca is the provincial tree of Yala province, Thailand [21].

MATERIALS AND METHODS:

Collection and Processing of Plant Material:

Saraca asoka was procured from the authorized botanist Dr. Madhukar Reddy of Heritage bionaturals, Habsiguda, Hyderabad. Shade dried samples were grounded to fine powder using pulverizer. The powdered samples were then stored in a refrigerator for further use.

Extraction of Plant Material:

The powdered barks of *Saraca asoka* were extracted using ethanol with gentle stirring for 72 h separately at room temperature. The extracts were then filtered through Whatmann No. 1 filter paper and concentrated using rotaevaporator.

FTIR Spectroscopic Analysis:

The FT-IR spectrophotometer used was Shumatzu at Osmania University. KBr is an important sample matrix for FTIR scanning. The KBr used was of IR grade (SD Fines). About 500 mg of KBr was placed into a mortar and grind it until there is no evidence of crystalinity. The KBr powder was transferred into the drying box at a temperature of 40°C. 10 mg of solid sample was placed into the mortar and again grind it until a fine powder is formed. Weigh 1mg of solid fine powder of sample (as per requirement of the die) and 200mg of dry fine powder of KBr. Weighed quantities were transferred into a mortar and mix well with the help of a spatula. Bottom and top portion of KBr were assembled at press assembly and one of the 13 mm die with the polished surface up inside the press. The KBr sample mixture was transferred to KBr press assembly. Second die was transferred inside the KBr press assembly with polished side down so that KBr sample mixture was sandwiched between the polished surfaces of the each die. The KBr was transferred to press assembly to press. Vacuum line was connected to evacuate air from the KBr press assembly with a vacuum pump. The die is slowly compressed in KBr press assembly until a pressure of 2000 kg/cm2 is achieved on gauge with the vacuum on. Making sure that pressure release valve is closed. After 60 s, slowly the pressure release valve is open to release the pressure and also the vacuum line is disconnected. The disc is checked if it is translucent and the sample is homogenously distributed in the disc²⁰. The prepared disc is then subjected for scanning between 500-4000⁻¹ cm.

RESULTS AND DISCUSSION:

The FTIR spectrum was used to identify the functional group of the active components based on the peak value in the region of infrared radiation. The barks powder and ethanolic extracts evaporated powder of *Saraca asoka* was passed into the FTIR and the functional groups of the components were separated based on its peak ratio.

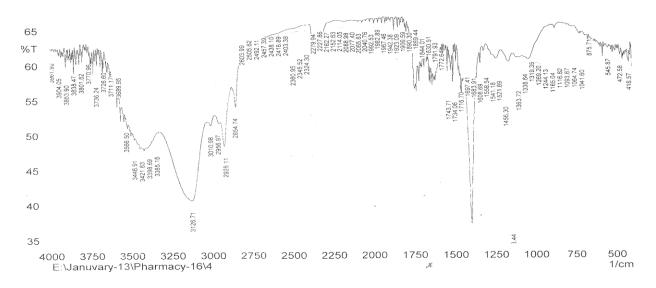


Fig:1 Showing the Peaks of FT-IR Scan of Ethanolic Extract Saraca Asoka.

The results of *Saraca asoka* barks FTIR analysis confirmed the presence of N-H stretching bond, alkenes group, aldehyde group, and carboxylic group which shows major peaks at 3421-3398 cm⁻¹. 3010cm⁻¹, 2926-2894 cm⁻¹, and 1697 cm⁻¹ respectively. Whereas the prominent peaks at 3838 cm⁻¹, 1859 cm⁻¹, 1791 cm⁻¹ and a broad plateau were unknown. The results of the present study produced the FTIR spectrum profile for the medicinally important plant *Saraca asoka*.

CONCLUSION:

Many researchers applied the FTIR spectrum as a tool for distinguishing closely associated plants and other organisms [1-19]. The results of the present study coincided with the previous observations observed by various plant biologist and taxonomist. The results of the present study developed novel phytochemical marker to identify the medicinally important plant. Further advanced spectroscopic studies are required for the structural elucidation and identification of active principles present in the leaves of *Saraca asoka*.

REFERENCES:

- 1. Griffiths PR, de Haseth JA. Fourier transform infrared spectroscopy. New York: Wiley, 1986.
- 2. Surewicz WK, Mantsch HH, Chapman D. Determination of Protein Secondary Structure by Fourier Transform Infrared Spectroscopy: A Critical Assessment. Biochemistry 1993; 32(2): 389-393.
- 3. McCann MC, Hammouri M, Wilson R, Belton P, Roberts K. Fourier transform infrared microspectroscopy is a new way to look at plant cell walls. Plant Physiol. 1992; 100: 1940- 1947.

- 4. Yang J, Yen HCE. Early Salt Stress Effects on the Changes in Chemical Composition in Leaves of Ice Plant and Arabidopsis. A Fourier Transform Infrared Spectroscopy Study. Plant Physiology 2002; 130: 1032-1042.
- 5. Stehfest K, Toepel J, Wilhelm C. The application of micro-FTIR spectroscopy to analyze nutrient stress-related changes in biomass composition of phytoplankton algae. Plant Physiology and Biochemistry 2005; 43: 717-726.
- 6. Ivanova DG, Singh BR. Nondestructive FTIR monitoring of leaf senescence and elicitininduced changes in plant leaves. Biopolymers 2003; 72(2): 79-85.
- 7. Helm D, Labischinski H, Schallehn G, Naumann D. Classification and identification of bacteria by Fourier transform infrared spectroscopy. J. Gen. Microbiol. 1991; 137:69–79.
- 8. Timmins EM, Howell SA, Alsberg BK, Noble WC, Goodacre R. Rapid differentiation of closely related *Candida* species and strains by pyrolysismass spectrometry and Fourier transform-infrared spectroscopy. J. Clin. Microbiol. 1998; 36: 367–374.
- 9. Goodacre R, Shann B, Gilbert RJ, Timmins EM, Mcgovern AC, Alsberg BK, Kell DB, Logan NA. Detection of the dipicolinic acid biomarker in *Bacillus* spores using Curie-point pyrolysis mass spectrometry and Fourier transform-infrared spectroscopy. Anal. Chem. 2000; 72:119–127.
- 10. Johnson HE, Broadhurst D, Goodacre R, Smith AR. Metabolic fingerprinting of salt-stressed tomatoes. Phytochemistry 2003; 62:919–928.
- 11. Martín JA, Solla A, Woodward S, Gil L. Fourier transform-infrared spectroscopy as a new method for

- evaluating host resistance in the Dutch elm disease complex. Tree Physiology 2005; 25, 1331–1338.
- 12. Liu H, Sun S, Lv G, Chan KKC. Study on Angelica and its different extracts by Fourier transform infrared spectroscopy and two-dimensional correlation IR spectroscopy. Spectrochimica Acta Part A 2006; 64: 321–326.
- 13. Duraes N, Bobos I, Ferreira Da Silva E. Chemistry and FT-IR spectroscopic studies of plants from contaminated mining sites in the Iberian Pyrite Belt, Portugal. Mineralogical Magazine 2008; 72(1): 405–409.
- 14. Wei ZL, Dong L, Tian ZH. Fourier transform infrared spectrometry study on early stage of cadmium stress in clover leaves. *Pak. J. Bot.* 2009; 41(4): 1743-1750.
- 15. Thenmozhi M, Bhavya PK, Rajeshwari Sivaraj. Compounds identification using HPLC and FTIR in *Eclipta alba* and *Emilia*
- *sonchifolia*. International Journal of Engineering Science and Technology 2011; 3(1):292-298.

- 16. Janakiraman N, Sahaya Sathish S, Johnson M. UV-VIS and FTIR Spectroscopic Studies on *Peristrophe bicalyculata* (Retz.) Nees. Asian Journal of Pharmaceutical and Clinical Research 2011; 4(4): 125-129.
- 17. Dorado J, Almendros G, Field JA, Sierra-Álvarez R. Infrared spectroscopy analysis of hemp (*Cannabis sativa*) after selective delignification by Bjerkandera sp. at different nitrogen levels. Enzyme Microb. Technol. 2001; 28:550–559.
- 18. Gidman E, Goodacre R, Emmett B, Smith AR, Jones DG. Investigating plant–plant interference by metabolic finger printing. Phytochemistry 2003; 63: 705–710.
- 19. Hori R, Sugiyama J. A combined FT-IR microscopy and principal component analysis on softwood cell walls. Carbohydr. Polym. 2003; 52: 449–453.
- 20. Kumar S, Joshi P, Rajvanshi A, Scanning Of Forensic Samples On Ftir Sample Preparation, The.Internet.J.For.Sci. 2008. (4)1:1-4.
- 21. https://en.wikipedia.org/wiki/Saraca_asoca.