

## Reduction of Aflatoxin M<sub>1</sub> during Production of Kashk, a Traditional Iranian Dairy Product

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### Abstract

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**Background:** Aflatoxin M<sub>1</sub> (AFM<sub>1</sub>) is a toxic compound that could be found in milk and dairy products. AFM<sub>1</sub> is the principle hydroxylated derivative of aflatoxin B<sub>1</sub> (AFB<sub>1</sub>), formed in liver and excreted into the milk in the mammary glands of both human and lactating animals that have been fed with AFB<sub>1</sub> contaminated diet. To the best of our knowledge, there is no report on stability of AFM<sub>1</sub> during production of Iranian traditional kashk (ITK). This study designed to assess the effect of ITK processing on AFM<sub>1</sub> content of cow's milk artificially contaminated with AFM<sub>1</sub>.

**Methods:** ITK making consisted of production of yogurt and strained yogurt, boiling the strained yogurt to make a curd and drying the resultant curd, according to common native procedure in Iran. AFM<sub>1</sub> content of initial yogurt, strained yogurt and dried kashk was determined using high performance liquid chromatography (HPLC). Statistical analyses were performed by Student's t-test and ANOVA using the SPSS 16.0 software package program.

**Results:** The mean concentration of AFM<sub>1</sub> in final ITK was determined as 0.118 µg/kg which was significantly ( $p < 0.05$ ) lower than that of initial milk, yogurt and strained yogurt samples. Also, AFM<sub>1</sub> content of yogurt, strained yogurt and final ITK product was 46.12%, 6.94% and 48.24%, respectively lower than that of initial milk ( $p < 0.05$ ).

**Conclusion:** This study showed that ITK processing can effectively degrade AFM<sub>1</sub> presented in initial raw milk and could be useful for minimizing AFM<sub>1</sub> content of highly contaminated raw milk in dairy industries.

### Introduction

Aflatoxins consist of four teratogenic, mutagenic and carcinogenic toxins including B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub>, and G<sub>2</sub>, which

are biosynthesized by toxicogenic strains of *Aspergillus flavus*, *A. parasiticus*, and *A. nomius* in various food commodities (Eslami et al., 2015; Fallah, 2010a; Fallah, 2010b; Iha et al., 2013; Mason et al., 2015).

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*A. flavus* exclusively produces B aflatoxins, while the two others produce B and G types. Aflatoxin M<sub>1</sub> (AFM<sub>1</sub>) is formed in liver and excreted into milk in the mammary glands of both human and lactating animals that have been fed with aflatoxin B<sub>1</sub> (AFB<sub>1</sub>) contaminated diet (Creppy, 2002; FAO, 2001; Kamkar et al., 2008; Khodadadi et al., 2014). The rate of alteration of AFB<sub>1</sub> to AFM<sub>1</sub> are various from 0.5 to 5% for lactating animals (Neal et al. 1998). AFM<sub>1</sub> has been classified as group 1 of human carcinogens (IARC, 2002). Among consumers, infants and young children use high amount of dairy products and therefore, effect of AFM<sub>1</sub> in these groups make great concern for public health. The accepted maximum level of AFM<sub>1</sub> content in milk and dairy products has been reported 0.05 µg/kg (EC, 2006b). There is some published data indicated high content of AFM<sub>1</sub> in milk and dairy products of Iran (Fallah, 2010a; Fallah, 2010b; Fallah et al., 2011; Ghazani, 2009; Heshmati, 2010; Heshmati and Milani, 2010; Iha et al., 2013). AFM<sub>1</sub> can be detected in dairy products if the initial milk is contaminated to this toxin (Bakirci, 2001). Unfortunately, a little reduction of AFM<sub>1</sub> occurs during normal heat processing and current food preservation methods (El Khoury et al., 2011; Fallah, 2010a; Iha et al., 2013).

South East Asia including, Iran, Iraq, Syria, and Turkey are important area for production and consumption of traditional fermented milk (Chandan, 2006). Due to nutritional benefits of yogurt, its consumption rate has been increased, recently (McKinley, 2005). Dried yogurt is normally obtained from dehydration of natural yogurt to keep the product stable and extend its shelf life. There are many kinds of dried yogurt and its derivatives worldwide. “Kashk” is local name of a popular and traditional dried yogurt in Iran. It is obtained from natural/plain yogurt when concentrated, boiled, shaped into flat rolls and dried. This product is very nutritious; it contains 95.6% total solid, 54.4%, protein 7.9% fat, and 29.5% lactose content (Tamime and Robinson, 2007).

Many researchers have attempted to study the effect of food processing on the residue of AFM<sub>1</sub> in milk and dairy products. To the best of our knowledge, there is lack of data about the variation of AFM<sub>1</sub> during processing of dried yogurt and its derivatives such as Iranian traditional kashk (ITK). Thus, the main aim of this study was to evaluate the stability of AFM<sub>1</sub> during the production and processing of ITK using high performance liquid chromatography (HPLC) assay.

## Materials and methods

### AFM<sub>1</sub> standard preparation

AFM<sub>1</sub> standard was purchased from sigma (Sigma Chemical Co., St Louis, MO). Working solution of AFM<sub>1</sub> was achieved by suitable dilution in acetonitrile.

To obtain final concentration of 0.1 to 10 ng/ml, different portions of the standard solutions were evaporated, and then diluted with mobile phase.

### Sample preparation and extraction

The samples were shaken for about 2 min to obtain homogeneous mixtures. Briefly, 10 g kashk and yogurt samples, 10 g celite (Sigma–Aldrich, 5876), and 80 ml dichloromethane (HPLC grade, Merck) were blended using Ultra Turrax (Junke and Kunkel, GmbH, Germany) at 24000 rpm for 3 min in order to form slurry. The slurry was filtered with Whatman no. 1 filter paper. Collected filtrate was evaporated to dryness under vacuum at 35–40 °C using laboratory type evaporator (Heidolph Rotary Evaporator VV–2000, Germany). The obtained residue was dissolved in a mixture (30:50:20 v/v/v) of methanol (HPLC grade, Merck), water and hexane (extra pure, Merck). The aqueous phase was separated using a separator funnel. An aliquot (35 ml) of the filtrate from each sample was passed through an immunoaffinity column (Aflaprep M, R-Biopharm Rhone Ltd., Glasgow, Scotland, UK) containing monoclonal antibodies specific to AFM<sub>1</sub> (Montaseri et al., 2014).

### Immunoaffinity column purification and isolation

Each test solution obtained from former step was passed through an immunoaffinity column, at a slow steady volume having flow rate of 1 ml/min. Then, column was secured on a vacuum manifold conditioned with 5 ml of phosphate-buffered saline. After double washing of the column with 10 ml of ultrapure water, AFM<sub>1</sub> was removed from the column with acetonitrile. The eluate was then evaporated under nitrogen flow (Dubnof Bath BSD/D). The residue was dissolved again in mobile phase, collected in HPLC vials (Supleco, Bellefonte, PA, USA), and finally injected to HPLC system (Montaseri et al., 2014).

### HPLC analysis

Setup of HPLC was carried out according to the published procedure by Montaseri (2014). Briefly, Column (Reverse phase ODS<sub>2</sub>–5 µm, 250 m × 4.6 m C18 Column TSK–GEL<sup>®</sup> TosoHas), Guard Column (Guard Column NovaPak<sup>®</sup> C18 Waters), and Mobile phase (acetonitrile: methanol: H<sub>2</sub>O; 20:20:60) were used. The flow rate of 1 ml/min, and injection volume of 20 µl were adjusted. Fluorescence detector was waters 2475 fluorescence detector, excitation 360 nm, emission 440 nm, and Gain and emission units full scale (EUFS) were 10 and 1000, respectively. Retention time was considered 3.5 min. HPLC System was Waters Breeze 1525 HPLC Pump, Waters 1525 Binary HPLC Pump, Waters Column

Heater, Waters Bus SAT/IN, Waters Bus Lace, Waters Breeze Software.

#### Validation of HPLC analytical method

The HPLC assay was validated based on the selectivity, linearity, sensitivity, accuracy and precision. The selectivity of the assay was determined by evaluation of blank and spiked samples at different levels of 0.1–10 ng/mg. The linearity was analyzed by constructing 9-point calibration curve at content of 0.1 to 10 ng/mg. Each concentration was injected four times and then evaluated by linear regression analysis and stated as correlation coefficient ( $R^2$ ). The precision of the assay was determined by limits of detection (LOD) and quantification (LOQ). The LOD, as the lowest content of AFM<sub>1</sub> which can be detected above the baseline, was determined by three fold analysis of spiked non-contaminated samples. LOQ, as the lowest content of analyzed sample, can be achieved with acceptable relative standard deviation (RSD) in within and between run procedure. To determine the accuracy (recovery), blank samples were spiked with suitable content of AFM<sub>1</sub> working standards to produce amounts of 0.02, 0.05 and 0.2 µg/kg. The recovery values were analyzed by the evaluation of three spiked samples with HPLC after extraction and immunoaffinity column clean-up mentioned previously (EC, 2006a; Montaseri et al., 2014).

#### Cow's milk artificially contaminated with AFM<sub>1</sub>

Skimmed cow's milk (dry matter, 9%) artificially contaminated with AFM<sub>1</sub> at level of 0.25 µg/l according to Montaseri et al. (2014) and heated to 42 °C. The AFM<sub>1</sub> content of the initial milk was under the limit of detection (0.01 ng/ml). A commercial starter culture (V1, direct vat set type, Chr. Hansen, Horlom, Denmark) with a mix of *Streptococcus thermophilus* and *Lactobacillus bulgaricus* (1:1) was used for yogurt making. According to manufacturer's recommendation the lyophilized starter culture was activated in sterilized skimmed milk and 4 ml of activated culture added to 1 l AFM<sub>1</sub> contaminated milk. After aseptically distributing in sterile plastic bottles, the incubation step was started until the pH of the samples reaches to 3.90 ± 0.02. At the end of the fermentation stage, each sample was quickly cooled in an ice bath and stored at 4 °C. Strained yogurt was produced from plain yogurt, for this purpose, yogurt was placed in cheesecloth and hung overnight at 4 °C.

#### Production of dried kashk from strained yogurt

According to ITK making method, the obtained strained yogurt was heated in a water bath at 95 °C for 4 h. After cooling, the resultant curd was divided into

small rounded pieces of kashk and dried to reach a moisture content of about 10%. AFM<sub>1</sub> content of initial yogurt, strained yogurt and dried kashk was determined. All the experiments were carried out in triplicate.

#### Statistical analysis

Statistical analyses of the obtained data were performed by Student's t-test and ANOVA using the SPSS, Inc, Chicago, IL (version 16.0) software package program. Probability ( $p$ ) values of less than 0.05 were considered as significant.

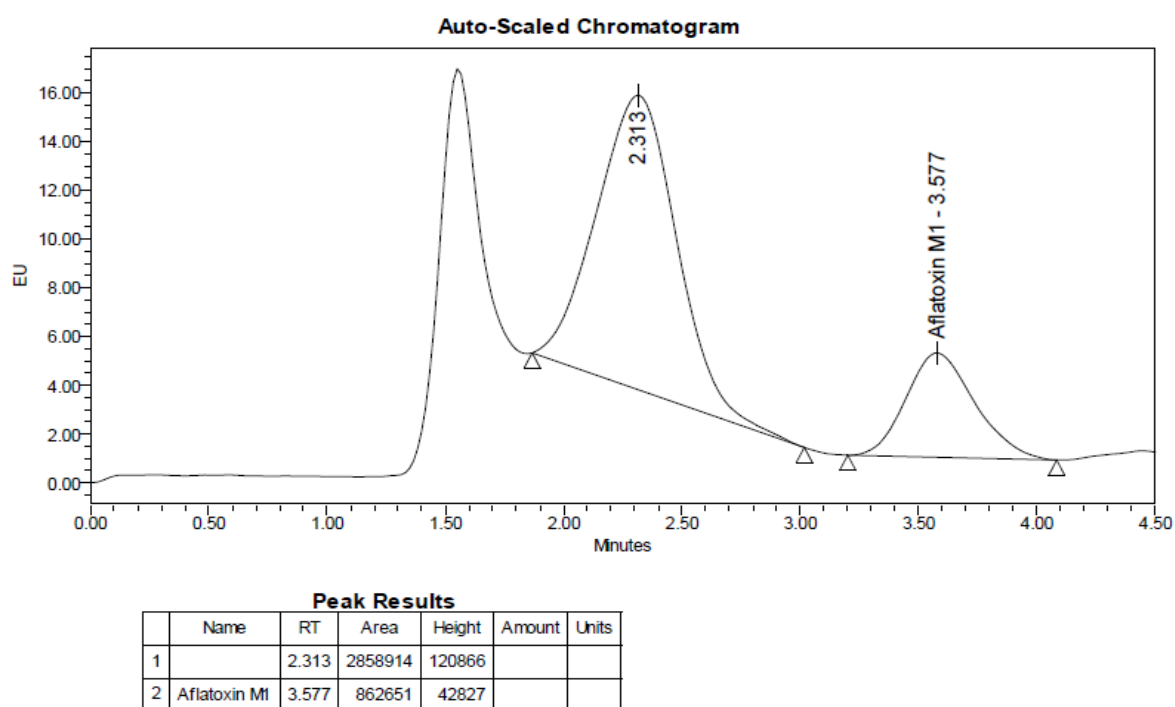
#### Results

In this study, selectivity of the method was confirmed using immunoaffinity column for clean up as well as a selective fluorescence detector. As shown in Fig. 1, no interfering peak was observed at the retention time of AFM<sub>1</sub> (3.5 min). LODs determined as the lowest concentration of AFM<sub>1</sub> that can be completely detected above the baseline signal and were 0.003 and 0.006 ng/g for yogurt and kashk samples, respectively. Also, LOQs were 0.01 and 0.019 ng/g for yogurt and kashk samples, respectively. Method validation data for AFM<sub>1</sub> determination in yogurt and kashk are indicated in Table 1.

As seen in Table 2, the mean concentration of AFM<sub>1</sub> in final ITK was determined as 0.118 µg/kg which was significantly ( $p < 0.05$ ) lower than that of initial milk, yogurt and strained yogurt samples. Also, AFM<sub>1</sub> content of yogurt, strained yogurt and final ITK product was 46.12%, 6.94% and 48.24%, respectively which were lower than that of initial milk ( $p < 0.05$ ).

#### Discussion

In this study, we found that there was a significant reduction in AFM<sub>1</sub> content during ITK production stages. Previously, Govaris et al. (2002) investigated the change of AFM<sub>1</sub> in yogurt samples that contaminated artificially with 0.05 and 0.1 µg/kg during 4 week storage at different pH levels of 4 and 4.6. They showed that stability of AFM<sub>1</sub> in test samples were pH dependent; in pH 4, the AFM<sub>1</sub> level was decreased significantly unlike pH 4.6; and at the end of storage time, AFM<sub>1</sub> showed significant decrease at both concentration levels. In contrast, Iha et al. (2013) reported that yogurt storage up to 28 days did not change the level of AFM<sub>1</sub>. It has been shown that factors such as low pH and fermentation of organic acids or other fermentation by-products may change the AFM<sub>1</sub> content in yogurt (Govaris et al., 2002). During yogurt production, decrease in pH level resulting from fermentation, changes the proteins structures e.g. casein. This phenomenon consequently resulted in the association of



**Fig. 1:** Chromatogram for sample of milk artificially contaminated with 0.1 ( $\mu\text{g/l}$ ) AFM<sub>1</sub>

**Table 1:** Method validation data for AFM<sub>1</sub> determination in yogurt and kashk

Matrix	Spiking level ( $\mu\text{g/kg}$ )	AFM <sub>1</sub> found (ng/kg)	SD ( $\mu\text{g/kg}$ )	Recovery (%)	RSD (%)
yogurt	0.05	48	0.005	96	6.5
	0.2	185	0.004	92.5	6.3
	0.5	470	0.005	94	6.7
kashk	0.05	38	0.008	76	9.1
	0.2	150	0.010	75	8.7
	0.5	365	0.011	73	8.9

**Table 2:** Changes in AFM<sub>1</sub> concentration during production of Iranian traditional kashk

Sample	Concentration of AFM <sub>1</sub> $\mu\text{g/Kg}$ (mean $\pm$ SD)	AFM <sub>1</sub> variation (%)	
		In comparison to initial spiked milk	In comparison to previous step
Artificially contaminated milk	0.245 $\pm$ 0.005		
Yogurt	0.132 $\pm$ 0.017	-46.12	
Strained yogurt	0.228 $\pm$ 0.006	-6.94	+72.72
Dried kashk	0.118 $\pm$ 0.006	-48.24	-51.84

AFM<sub>1</sub> or other toxins with casein (Brackett and Marth, 1982). As illustrated in Table 2, the concentration of AFM<sub>1</sub> in strained yogurt was significantly higher than yogurt sample. In agreement to this result, Govaris et al. (2002) showed that the concentration of AFM<sub>1</sub> in strained yogurt was higher than its correspondence plain yogurt. This could be explained by the effect of increasing the concentration of total solids of yogurt when removing its water content.

The mean concentration of AFM<sub>1</sub> in ITK (0.118  $\mu\text{g/kg}$ ) obtained in the present work was significantly lower than milk, yogurt and strained yogurt samples. Previous stud-

ies showed that the concentration of AFM<sub>1</sub> increased after physical water removing from dairy product such as cheese (Kamkar et al., 2008) and strained yogurt (Govaris et al., 2001). On the other hand, heat treatment of milk and dairy product had no remarkable effect on the concentration of AFM<sub>1</sub> indicated by Bakirci (2001) and Govaris et al. (2002). Contrary to mentioned finding, the obtained result of this study showed that the concentration of AFM<sub>1</sub> significantly decreased after transforming the milk in comparison to ITK. It may be explained by the synergistic effect of the combination of heat treatment and low pH during ITK making process that

lead to strongly degradation of AFM<sub>1</sub>. Montaseri et al. (2014) stated that processing and storage of probiotic yogurt may decrease the AFM<sub>1</sub> content of initial milk. However, probability of AFM<sub>1</sub> reduction during producing and processing of probiotic ITK is not still reported in scientific database.

## Conclusion

AFM<sub>1</sub> is a hazardous component that can seriously threaten the public health by dairy consumption. It is necessary to search methods capable of removing or inactivating AFM<sub>1</sub> in dairy products. The results of this study showed that ITK processing can effectively degrade AFM<sub>1</sub> presented in initial raw milk and could be useful for minimizing AFM<sub>1</sub> content of highly contaminated raw milk in dairy industries. More studies are needed to assess the effects of the other aspects of ITK processing on AFM<sub>1</sub> stability.

## Conflicts of interest

All the authors of this article state that they had no conflicts of interest.

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## References

- Bakirci I. (2001). A study on the occurrence of aflatoxin M<sub>1</sub> in milk and milk products produced in Van province of Turkey. *Food Control*. 12: 47-51.
- Brackett R.E., Marth E.H. (1982). Association of aflatoxin M<sub>1</sub> with casein. *Zeitschrift für Lebensmittel-Untersuchung und Forschung*. 174: 439-441.
- Chandan R.C. (2006). History and Consumption Trends. Manufacturing yoghurt and fermented milks. Blackwell Publishing Ltd, UK. pp: 5-400.
- Creppy E.E. (2002). Update of survey, regulation and toxic effects of mycotoxins in Europe. *Toxicology Letters*. 127: 19-28.
- El Khoury A., Atoui A., Yaghi J. (2011). Analysis of aflatoxin M<sub>1</sub> in milk and yoghurt and AFM<sub>1</sub> reduction by lactic acid bacteria used in Lebanese industry. *Food Control*. 22: 1695-1699.
- Eslami M., Mashak Z., Heshmati A., Shokrzadeh M., Mozaffari Nejad A.S. (2015). Determination of aflatoxin B<sub>1</sub> levels in Iranian rice by ELISA method. *Toxin Reviews*. 34: 125-128.
- European Commission (EC). (2006a). Regulation (EC) No. 401/2006 of 23 February 2006, laying down the methods of sampling and analysis for the official control of the levels of mycotoxins in foodstuffs. *Official Journal of the European Union*. 70: 12-34.
- European Commission (EC). (2006b). Regulation (EC) No. 1881/2006 of 19 December 2006, setting maximum levels for certain contaminants in foodstuffs. *Official Journal of the European Union*. 364: 5-24.
- Fallah A.A. (2010a). Aflatoxin M<sub>1</sub> contamination in dairy products marketed in Iran during winter and summer. *Food Control*. 21: 1478-1481.
- Fallah A.A. (2010b). Assessment of aflatoxin M<sub>1</sub> contamination in pasteurized and UHT milk marketed in central part of Iran. *Food and Chemical Toxicology*. 48: 988-991.
- Fallah A.A., Rahnama M., Jafari T., Saei-Dehkordi S.S. (2011). Seasonal variation of aflatoxin M<sub>1</sub> contamination in industrial and traditional Iranian dairy products. *Food Control*. 22: 1653-1656.
- Food and Agriculture Organization (FAO). (2001). Safety evaluation of certain mycotoxins in food. Prepared by the fifty-sixth meeting of the joint FAO/WHO expert committee on food additives (JECFA). FAO food and nutrition paper 74, Food and Agriculture Organization of the United Nations, Rome, Italy.
- Ghazani M.H.M. (2009). Aflatoxin M<sub>1</sub> contamination in pasteurized milk in Tabriz (North West of Iran). *Food and Chemical Toxicology*. 47: 1624-1625.
- Govaris A., Roussi V., Koidis P.A., Botsoglou N.A. (2002). Distribution and stability of aflatoxin M<sub>1</sub> during production and storage of yoghurt. *Food Additives and Contaminant*. 19: 1043-1050.
- Heshmati A. (2010). Occurrence of aflatoxin M<sub>1</sub> in Iranian white cheese. *Iranian Journal of Food Science and Technology*. 7: 117-122.
- Heshmati A., Milani J.M. (2010). Contamination of UHT milk by aflatoxin M<sub>1</sub> in Iran. *Food Control*. 21: 19-22.
- Iha M.H., Barbosa C.B., Okada I.A., Trucksess M.W. (2013). Aflatoxin M<sub>1</sub> in milk and distribution and stability of aflatoxin M<sub>1</sub> during production and storage of yoghurt and cheese. *Food Control*. 29: 1-6.
- International Agency for Research on Cancer (IARC). (2002). IARC monograph on the evaluation of carcinogenic risk to humans, some traditional herbal medicines, some mycotoxins, naphthalene. Lyon, France.
- Kamkar A., Karim G., Shojaee Aliabadi F.S., Khaksar R. (2008). Fate of aflatoxin M<sub>1</sub> in Iranian white cheese processing. *Food Chemical and Toxicology*. 46: 2236-2238.
- Khodadadi M., Khosravi R., Allahresani A., Khaksar Y., Rafati L., Barikbin B. (2014). Occurrence of aflatoxin M<sub>1</sub> in pasteurized and traditional cheese marketed in southern Khorasan, Iran. *Journal of Food Quality and Hazards Control*. 1: 77-80.
- Mason S., Arjmandtalab S., Hajimohammadi B., Khosravi Arsanjani A., Karami S., Sayadi M., Oryan A. (2015). Aflatoxin M<sub>1</sub> contamination in industrial and traditional yogurts produced in Iran. *Journal of Food Quality and Hazards Control*. 2: 11-14.
- McKinley M.C. (2005). The nutrition and health benefits of yoghurt. *International Journal of Dairy Technology*. 58: 1-12.
- Montaseri H., Arjmandtalab S., Dehghanzadeh G., Karami S., Razmjoo M.M., Sayadi M., Oryan A. (2014). Effect of production and storage of probiotic yogurt on aflatoxin M<sub>1</sub> residue. *Journal of Food Quality and Hazards Control*. 1: 7-14.
- Neal G.E., Eaton D.L., Judah D.J., Verma A. (1998). Metabolism and toxicity of aflatoxins M<sub>1</sub> and B<sub>1</sub> in human-derived in vitro system. *Toxicology and Applied Pharmacology*. 151: 152-158.
- Tamime A.Y., Robinson R.K. (2007). Tamime and Robinson's Yoghurt: Science and Technology. Third edition. CRC Press, Chicago. pp: 348-467.