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In vitro Inhibition of Candida Species by Aqueous Garlic Extract in Gel and Lotion: Problem of Stability

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ABSTRACT

The inhibitory effect of aqueous garlic extract (AGE) against clinically significant microorganisms including Candida species is documented. The activity and stability of this extract in topical pharmaceutical formulations has received little attention. This communication reports on the stability and anticandidal activity of AGE in gel and lotion formulations. Gel and lotions containing 50, 100 and 200 mg garlic/ml were prepared and their physical as well as biological stability was established at 40°C. The anticandidal effect of all preparations was determined using the well diffusion method and Muller Hinton agar. Results indicated that both of the prepared formulations were physically stable but the freshly prepared lotion exhibited no anticandidal effect. Gels were effective inhibitors but this activity was lost with storage at 40°C in less than 9 days. It is concluded that lotion was not at all a suitable delivery system for AGE as it lost its activity during preparation. Whereas, freshly prepared gels were good inhibitors but this property was diminished upon storage. Comprehensive studies are needed to stabilize the AGE gel formulation over the expected shelf life.

Keywords: Aqueous garlic extract, Anticandidal effect, Formulation, Gel, Lotion, Biological stability.

INTRODUCTION

There is a copious amount of scientific literature that deals with garlic (*Allium sativum* L.) as an effective agent for the treatment of various illnesses. ^[1-2] Of special interest is the antimicrobial effect of garlic extract or garlic components and again this subject has been extensively investigated. ^[3-5] In fact there is enough evidence which indicates that garlic exhibits antimicrobial activity against a wide range of microorganisms comparable to or exceeds that of

*Corresponding author: Dr. Q. M. Abu Shaqra, Department of Allied Medical Sciences, Zarqa University College, Al- Balqa Applied University, P. O. Box 5341- Zarqa, 13111-Jordan; **Tel.:** +962-79-5009808; **E-mail:** qabushaqra@hotmail.com **Received:** 24 May, 2015; **Accepted:** 26 May, 2015 expensive antibiotics. ^[6] Even multi antibiotic resistant organisms were found to be susceptible to garlic. ^[7] This particular observation is important as it places cheep plants in the forefront as a possible alternative to the "by many" unaffordable modern antimicrobial drugs.

Garlic contains at least 33 sulfur compounds, several enzymes, minerals, vitamins, fiber and water. ^[8] The most abundant sulfur compound in garlic is alliin (Sallylcysteine sulfoxide), which is present at 10 and 30 mg/g in fresh and dry garlic, respectively. ^[9] Allicin which is the major antimicrobial component isolated from garlic, is formed when the garlic clove is crushed and the enzyme alliin lyase comes into contact with alliin. The main inhibitory effect of allicin may be due to its chemical reaction with thiol groups of various enzymes, e.g. alcohol dehydrogenase, thioredoxin reductase, and RNA polymerase. [10-11]

The antimicrobial effect of garlic extract incorporated into pharmaceutical formulations particularly those applied topically was the subject of few publications. Most important topical preparations are gels, lotions and creams. The addition of garlic volatile oil in the formulation of gels was tried by Bodhankar and Patil ^[12], whereas the addition of allicin into a readymade gel preparation was also reported. [11] Garlic extract or allicin were added to commercially available cream to establish the suitability of this formulation as a carrier of the antimicrobial activity exhibited by garlic and results proved encouraging. [13] The approach of triturating any component or extract with commercially available creams poses questions regarding the industrial feasibility of this practice and casts doubt about the long term physical stability of the preparation. It must be emphasized that as far as is known there has been no attempt to study the biological stability of garlic or allicin in formulated topical products.

No formulation can be proposed for commercial use without providing evidence to its physical and biological stability. The proposed preparation should be applicable for industrial scale production and the manufactured product should maintain its antimicrobial effectiveness over the expected shelf life. The objectives of this investigation were first to establish the in vitro activity of a simple gel and lotion formulations "with AGE" against clinical isolates of Candida species, second to determine the biological stability of these preparations over a period of time.

MATERIALS AND METHODS

Garlic aqueous extract preparation

Fresh garlic bulbs (Allium sativum L.) were purchased from a local supermarket in the city of Zarga, Jordan. The cloves were peeled and 500 g of the edible portion was homogenized in 1 liter of sterile distilled water using pistil and mortar. The homogenate was allowed to stand for 2 hours then filtered under vacuum through a 25 mm pore-size filter. The filtrate was collected in a flask and then stored at 4°C until used. This aqueous extract contained 500 mg of garlic/ml.

Gel formulation

The gel formulation was prepared by mixing 200 ml of aqueous garlic extract in 764 ml of sterile distilled water. To this mix 20 g of Carbomer 934 fine powder was added slowly while mixing and the gel was formed after the addition of 16 g Triethanolamine to the preparation while being agitated. This formulation contained 100 mg/ ml aqueous garlic extract. Three other preparations were also processed in the same manner to contain zero, 50 and 200 mg/ ml garlic extract.

Lotion Formulation

A simple lotion was prepared as follows: to a mix containing 763 ml sterile distilled water and 200 ml of garlic extract, 18 ml of triethanolamine was added, mixed, heated and maintained at 80°C. In another vessel 20 g cetyl alcohol, 10 g stearic acid and 10 g of beeswax were heated to 80°C. The prepared aqueous mix was added to the oily preparation while mixing at 80°C. Two other similar formulations were prepared; one with 200 g/ml and the other devoid of garlic extract.

Inoculums Preparation

A total of 6 Candida isolates were obtained from the laboratories of Jordan University hospital. Each isolate was grown on Sabouraud dextrose agar for overnight at 37°C before colonies were harvested and suspended in phosphate buffer pH 7.0. The turbidity of the suspension was adjusted to that of 0.5 MacFarland standards and thus contained 108 CFU/ml. [7] This suspension was further diluted with the same buffer so that the final concentration of cells would be 106 CFU/ml or as required.

Inhibition studies

To establish the anti candidal effect of aqueous garlic extract in formulation, Sabouraud dextrose agar plates were seeded with each clinical isolate under test from the dilution that contained 106 CFU/ ml. After drying, 4 wells in each plate were bored using a sterile cork borer of 6 mm in diameter. One hundred micro liters of each gel or lotion was separately dispensed into a well and plates were then incubated at 37°C for 24 hours before the zone of inhibition around each well was measured. For each AGE concentration and each test organism triplicate plates were prepared. The difference between the mean values was analyzed by Student's t test and *p* value of < 0.05 was considered as significant,

Time-kill investigation

In vitro kill rate of garlic extract against 2 clinical isolates of Candida species was performed following the technique described by Yen et al. [14] but with some modifications. A cell suspension of the Candida species under test was prepared as described above. One ml aliquot of this suspension was inoculated into 9 ml of the gel formulations prior to the addition of few drops of triethanolamine to ensure even distribution of the inoculum. At intervals, 0.5 ml quantity was withdrawn and transferred to a tube containing 4.5 ml sterile phosphate buffer pH 5.5, mixed and the number of surviving Candida was determined by plating 100 micro liters aliquot onto SDA.

Stability Study

Two gel formulations were made; one with 100 mg/ml AGE and the other with 200 mg/ml. Both preparations were incubated at 40°C and at intervals the persistence of the inhibitory effect of these formulations against C. albicans was determined as described above.

RESULTS AND DISCUSSION

This investigation was primarily concerned with the formulation of topical preparations for carrying AGE in presentable pharmaceutical form which can be manufactured at an industrial scale and this was Int. J. Pharm. Sci. Drug Res. May-June, 2015, Vol 7, Issue 3 (304-307) 305

achieved. The second concern was to determine if AGE in formulation can maintain its inhibitory effect against clinical isolates of Candida species and for how long. Soon after preparation, lotion formulation exhibited no antagonistic effect against Candida species tested in this study whereas; gel formulations were inhibitory to all isolates at 100 and 200 mg AGE/ml. This is clearly demonstrated in Table 1 which shows the zone of inhibition measured to various AGE concentrations incorporated into the gel preparation. The same table demonstrates that the gel prepared without any AGE exhibited no anticandidal effect. Therefore, any inhibition obtained must have been solely due to the added AGE.

Table 1: Susceptibility of Candida species to various concentrations of AGE in gel formulation (inhibition zone is the mean of 3 readings measured in mm)

Candida spp.	Zone of inhibition in mm.			
	200 mg/ ml	100 mg/ ml	50 mg/ ml	0 mg/ ml
C. albicans	23	18	12	Zero
C. tropicalis	26	16	15	Zero
C. glabrata	19	13	10	Zero
C. parapsilosis	14	9	Zero	Zero
C. krisei	12	8	Zero	Zero

The oil and water phases of lotion were mixed after heating each portion at about 80°C and this might explain why this formulation demonstrated no inhibitory effect against the Candida species tested. Sah et al [15] reported that aqueous extract derived from garlic cloves boiled in water at 100°C for 30 minutes, exhibited reduced antimicrobial properties as compared to that obtained by cold extraction. Dababneh and Al-Delaimy [16] indicated that heating garlic extract at 80°C to 90°C for 5 minutes completely destroyed its antimicrobial activity. Therefore, the heat treatment employed in the preparation of lotion was probably behind the lack of anticandidal activity in our preparation. Perhaps, this is why Cutler and Wilson ^[14] preferred to use a readymade cream for testing the antimicrobial effect of AGE rather than to prepare a cream formulation in the laboratory; rightly so to avoid heat treatment.

When the gel contained 50 mg/ml concentration of AGE, no inhibitory effect against *C. parapsilosis* or *C. krisei* was observed. It is important to note that the strength of garlic extract used in the formulations studied did not rely on the minimum inhibitory concentration of AGE reported in literature but rather depended on pragmatic data. Cutler *et al*, ^[11] used 500 mg Allicin/100 ml of gel to test for the antimicrobial effect whereas, Adejare *et al* ^[17] found that their clinical isolates of Candida species required as high as 200 mg/ml of AGE for effective inhibition. Therefore, these concentrations were used in this investigation.

Kill time of *C. albicans* and *C. parapsilosis* inoculated separately into gels with 50, 100, and 200 AGE is shown in Figure 1 (A & B). It is evident from figure 1A that complete inhibition of *C. albicans* was achieved in less than 4 hours regardless of the AGE concentration used.

Figure 1B shows that *C parapsilosis* was killed by the gels containing 100 mg/ml and 200 mg/ml in less than 5 hours. Thus, kill time is dependent on the AGE concentration used in formulation and the microbial strain tested. However, this particular observation is in agreement with those reported by others for plain AGE, the kill time for *C. albicans* was almost half the time reported by Iwalokun *et al* ^[7]. This difference could be attributed to the higher concentration of AGE used in the gel formulation employed in this study.

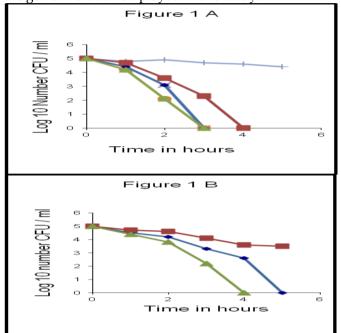


Fig. 1: Effect of different AGE concentrations in a freshly prepared gel formulation on the survival of *C. albicans* (Fig. 1A) and *C. parapsilosis* (Fig. 1B). \blacktriangle 200 mg/ ml, \bullet = 100 mg/ ml, \bullet = 50 mg/ ml, x = Plain gel with no AGE

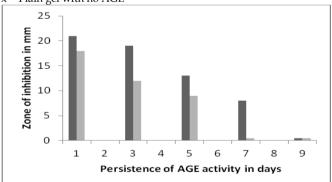


Fig. 2: Persistence of inhibitory effect in mm versus concentration of AGE in gel formulations (Dark column= 200 mg /ml, Light column= 100 mg /ml)

Stability study of any pharmaceutical product is performed to ensure the maintenance of product quality, safety and efficacy throughout the shelf life. ^[18] Figure 2 demonstrates that freshly prepared gels with 2 different concentrations of AGE was active inhibitor to the *C. albicans* studied, but this activity was reduced by almost 50% after 5 days of incubation at 40°C and completely lost after 9 days.

This means that the formulated gel was not biologically stable as the AGE antimicrobial activity disappeared from the preparation. Belguith *et al* ^[19] found that while

AGE stored at 4°C retained at least 90% of its original inhibitory potential for more than 10 days, the extract solution stored at 22°C lost the antibacterial activity slowly over a period of 6 to 10 days. Chong et al [20] investigated the Allicin stability in aqueous garlic extract by high performance liquid chromatography and demonstrated that when the garlic extract was incubated at 37°C, there was more than 50% loss of allicin over 5- day period. These findings are very close to those reported herein. Because Allicin plays a vital role in the antimicrobial activity of garlic, its degradation means loss of this activity. This is probably the case with our gel preparation which acted as growth inhibitor to all Candida species when it was just prepared but with the passage of time allicin was degraded and thus this activity was diminished. Block ^[21] attributed the loss of allicin biological activity to its transformation with time into more stable components: polysulfides and thiosulphonates. The biological halflife of allicin in water was determined by Fujisawa et al ^[22]; they found that the generated antibacterial activity and the detectable Allicin in crushed fresh garlic cloves declined in aqueous and ethanolic solutions upon storage at room temperature. The same authors calculated the biological and chemical half-lives to be 6 and 11 days, respectively. Gel formulations are aqueous preparations and therefore, the biological activity of garlic in such a formulation would be expected to follow the same trend as observed for aqueous garlic extract. This is exactly what was observed in this investigation.

Formulation of active substance into a suitable dosage form in addition to its stability and efficacy are components of the lengthy process of product development. It is concluded that lotion is not a good choice as a delivery system for carrying the anticandidal activity of AGE due the inactivation of garlic active molecules by heat. Formulation of AGE into a gel which can be produced at a commercial scale is presented but a long time might elapse before the problem of its biological stability can be resolved.

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REFERENCES

- Gebreyohannes G, Gebreyohannes M. Medicinal values of garlic: A review. International Journal of Medicine and Medical Sciences 2013; 5(9): 401-408.
- Chan JY, Yuen AC, Chan RY, Chan SW. A review of the cardiovascular benefits and antioxidant properties of allicin. Phytotherapy Research 2013; 27: 637–646.
- 3. Al-Astal ZY. Effect of storage and temperature of aqueous garlic extract on the growth of certain pathogenic bacteria. J Al Azhar Univ- Gaza 2003; 6: 11-20.
- Praba K, Kumaresan R. Efficacy of antimicrobial activity of aqueous garlic (*Allium sativum*) extract against different bacterial species. Journal of Chemical and Pharmaceutical Research 2014; 6(10): 677-679.

- 5. Sharanappa R, Vidyasagar GM. Anti-candida activity of medicinal plants. A review 2013 Inter Journal of Pharmacy and Pharmaceutical Sciences 5: Suppl 4. 9-16
- Srinivasan D, Lakshmanaperumalsamy P. In vitro Antibacterial activity and stability of garlic extract at different pH and temperature. Electronic Journal of Biology 2009; 5(1): 5-10
- Iwalokun BA, Ogunledun A, Ogbolu DO, Bamiro SB, Jimi-Omojola J. *In vitro* antimicrobial properties of aqueous garlic extract against multidrug-resistant bacteria and Candida Species from Nigeria. Journal of Medicinal Food 2004; 7 (3): 327-333.
- 8. Josling PA. The heart of garlic. Nature's aid to healing the human body, HEC Publishing Chicago Illinois, 2005, pp. 20.
- Lawson LD, Koch HP. The composition and chemistry of garlic cloves and processed garlic; in Garlic: the science and therapeutic application of *Allium sativum* L and related species. Edn 2, Baltimore: Williams and Wilkins, 1996. pp. 37–107.
- Ghannoum MA. Studies on the anticandidal mode of action of *Allium sativum* (garlic) Journal General Microbiology 1988;134: 2917-2924.
- 11. Cutler RR, Odent M, Hajj-Ahmad H, *et al.* In vitro activity of an aqueous allicin extract and a novel allicin topical gel formulation against Lancefield group B streptococci. Journal of Antimicrobial Chemotherapy 2009; 63 (1):151-154.
- Bodhankar, MM, Patil AT, Agnihotri VV. Development of topical formulations containing bioactive volatile oils: relative characterization of drug release for cream ointment and gel. Inter Journal of Pharmaceutical Technology 2011; 3(3): 3029-3038.
- Cutler RR, Wilson P. Antibacterial activity of a new, stable, aqueous extract of allicin against methicillanresistant Staphylococcus aureus. British Journal Biomedical Sciences 2004; 61(2): 71–74.
- 14. Yin MC, Chang HC, Tsao SM. Inhibitory effects of aqueous garlic extract, garlic oil and four diallylsulphides against four enteric pathogens. Journal of Food Drug Analysis 2002; 10: 12-126.
- Sah Pan, Al-Tamimi B, Al-Nassri N, Al-Mamari R. Effect of temperature on antibiotic properties of garlic (*Allium sativum* L.) and ginger (*Zingiber officinale* Rosc). African Journal of Biotechnology 2002; 11 (95): 16192–16195.
- Dababneh, BFA, Al-Delaimy KS. Inhibition of *Staphylococcus aureus* by garlic extract. Lebensm.-Wissenschaft und-Technology 1984; 17: 29-31.
- Adejare OY, Oduyebo OO, Oladele RO, Nwaokorie FO, Ogunsola FT. *In-vitro* antifungal effect of *garcinia kola* and garlic (*alliums sativum*) on vaginal isolates of candida. African Journal Clinical and Experimental Microbiology 2013; 14(3): 140-145.
- Bajaj S, Dinesh Singla D, Sakhuja N. Stability testing of pharmaceutical products. Journal of Applied Pharmaceutical Science 2012; 2 (03): 129-138.
- Belguith H, Kthiri F, Chati A, Abu Sofah A, Ben Hamida J, Landoulsi A. Study of the effect of aqueous garlic extract (*Allium sativum*) on some *Salmonella serovars* isolates. Emirate Journal of Food and Agriculture 2010; 22 (3): 189-206.
- 20. Chong K, Zamora MP, *et al.* Investigation of Allicin stability in aqueous garlic extract by high performance liquid chromatography method. Journal of Scientific Research & Reports 2015; 4(7): 590-598.
- 21. Block, E. The organosulfur chemistry of the genus Allium implications for organic sulfur chemistry. Angew and te Chemie. International Edition 104, 1992, 1158–1203.
- 22. Fujisawa H, Suma k, Origuchi K, *et al.* Biological and chemical stability of garlic-derived Allicin. J of Agriculture and Food Chemistry 2008; 56 (11): 4229- 4235.

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