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# Studies on free-radical scavenging activity and identification of active ingredients of different plant crude extracts of *Mentha piperita* collected from Sur, Sultanate of Oman

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#### ABSTRACT

**Objective:** To determine free radical scavenging activity and active chemical ingredients of different plant crude extracts of *Mentha piperita* (*M. piperita*).

**Methods:** The dried powder leaves of *M. piperita* were extracted with polar organic solvent by Soxhlet extractor. The crude extract and its fractions of hexane, chloroform, ethyl acetate and butanol crude extracts were prepared. The antioxidant activity of different crude extracts from *M. piperita* was carried out by DPPH method with minor modification, and the active chemical ingredients of different plant crude extracts of *M. piperita* were analyzed by gas chromatographymass spectrometry (GC–MS).

**Results:** Qualitative analysis of different polarities crude extracts by GC–MS found different types of active organic compounds. The antioxidant activity of different crude extracts were found to be in the order of chloroform extract> butanol extract> ethyl acetate extract> hexane extract> methanol extract. Majority identified compounds in the plant crude extracts by GC–MS were biologically active.

**Conclusions:** Therefore, the isolation, purification, identification and characterization of bioactive compounds from various crude extracts of *M. piperita* might have ecological significance.

#### **1. Introduction**

Mentha piperita L. (M. piperita) is the most important traditional medicinal plants available worldwide. It is

also found in the Sultanate of Oman. Locally it is known as peppermint belonging to the family of Lamiaceae. Traditionally, this plant is used by different ethnic communities of Sultanate of Oman for the treatment of cough and other cough related diseases<sup>[1]</sup>. It is native to European countries and Asia. Some varieties of M. *piperita* are indigenous to South Africa, South America, and Australia.Now it is a hybrid plant and cross between watermint and spearmint<sup>[2]</sup>. This plant grows well in all environmental conditions and the height is about 2–3 feet. It comes into flower from July through August,

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sprouting tiny purple flowers in whorls and terminal spikes. Dark green fragrant leaves grow opposite white flowers<sup>[3]</sup>. The whole plant is used as a flavoring agent for the preparation of food, gum, toothpaste and tea<sup>[4]</sup>. It is also used to soothe upset stomach and to aid digestion<sup>[5]</sup>. The crude extracts from *M. piperita* have been used as the therapeutic treatment of headaches, skin irritations, anxiety associated with depression, nausea, diarrhea, menstrual cramps and flatulence<sup>[4]</sup>. *In vitro* test shows that *M. piperita* crude extracts kill some types of bacteria, fungus and viruses. It suggests that it may have antibacterial, antifungal, and antiviral properties<sup>[6,7]</sup>.

Several scientific studies supported the use of *M. piperita* crude extract for indigestion and irritable bowel syndrome<sup>[8]</sup>. Recently, it has been also used to relax the muscles that allow painful digestive gas to pass<sup>[9–11]</sup>. The reported active main ingredient of *M. piperita* is menthol, which is an effective decongestant<sup>[9–11]</sup>. The menthol is also a good expectorant, meaning that it helps loosen phlegm and breaks up coughs<sup>[12–15]</sup>.

The previous study mentioned that the leaves and stems contained high concentrations of menthol type compounds that are used medicinally as a flavoring agent in food and cosmetics<sup>[16–18]</sup>. The literature search reveals that still no research has been done on Omani *M. piperita*. In addition, the available information with regard to correlating the phytochemical constituents and other activities of this plant is lacking. Therefore, the present study is to prepare different crude extracts collected from locally grown species and to identify their active chemical ingredients by gas chromatography– mass spectrometry (GC–MS) and evaluate their free radical scavenging activity by DPPH method.

#### 2. Materials and methods

#### 2.1. Sample collection

The healthy whole plants of *M. piperita* were collected from garden at Sur, Sultanate of Oman. The plants were harvested during the month of April 4, 2012 at 8 am. The collected samples were packed in a polyethylene bag and stored at 4  $^{\circ}$ C.

# 2.2. Preparation of samples

The collected plant samples were washed with fresh

water to remove all dust. The whole plant samples were dried under shade for complete dry. Approximately 60 g of plant samples were ground using a grinder (Super Deluxe, India) for 20 seconds. The small pieces of samples were homogenized in a grinder for 3 min to 40–60 mesh size. The dried plant sample was pulverized into powdered form.

### 2.3. Preparation of crude extracts

The powder sample (60 g) was extracted with methanol (200 mL, 72 h) by using a Soxhlet extractor. The solvent was evaporated by rotary evaporator (Yamato, Rotary Evaporator, Model–RE 801) to give gummy paste weight about 7.82 g. The 7 g crude extract was suspended in 100 mL of distilled water. Then it was extracted successively with hexane, chloroform, ethyl acetate and butanol to derive hexane (1.05 g), ethyl acetate (1.47 g), chloroform (0.37 g), butanol (1.45 g) and residual methanol fractions (2.13 g), respectively. The plant residue was re–extracted twice following the same procedure and filtered. The combined extracts were concentrated by using rotary evaporator and dried under vacuum.

#### 2.4. Radical scavenging activity using DPPH method

Free radical scavenging activity of different crude extracts with different polarities of M. piperita was estimated by the method described by Hossain with minor modification<sup>[19]</sup>. The different crude extracts of M. piperita (4 mL) in different concentrations (12.5, 25, 50, 100 and 200 mg/L, respectively) were taken in different test tubes. One milliliter of 0.1 mmol/L DPPH was dissolved in methanol, added to those tubes, and shaken vigorously. After shaking, all the test tubes were allowed to stand at 27 °C in a dark place for 45 min. The blank control was prepared in the same way without extract. The differences in the absorbance of the prepared samples were measured using UV spectroscopy at 517 nm. Radical scavenging activity of the tested different crude extracts was estimated as an inhibition percentage (%) and calculated by using the following formula,

Measurement of radical scavenging activity (%)

% Inhibition = 
$$\frac{A_{control} - A_{extract}}{A_{control}} \times 100$$

#### 2.5. GC-MS analysis

The GC-MS analysis of different polarities crude extracts from whole plant of *M. piperita* was performed using a Perkin Elmer Clarus 600 GC system equipped with a Rtx®-5MS fused silica capillary column (30 m×0.25 mm i.d., film thickness 0.25 µm). The GC system was coupled with a Perkin Elmer Clarus 600 MS. Gas chromatographymass spectroscopic detector of an electron ionization mode with ionization energy of 70 eV was used. Helium gas was used as a carrier gas at a constant flow rate of 1 mL/min. Mass transfer line and injector temperatures were set at 260 and 290 °C, respectively. The oven was programmed from 60 °C (hold 2 min) to 270 °C at 4 °C/min, then held isothermal for 20 min and finally raised to 290 °C at 10 °C/min. Diluted samples (1/100, v/v, in methanol) of 1  $\mu$ L were injected in the split mode with a split ratio of 120:1. The calculation of the percentage (%) of the crude extracts constituents was expressed as a percentage by peak area normalization.

#### 2.6. Identification of chemical constituents

The chemical ingredients in different crude extracts from the *M. piperita* were identified based on GC retention time on Rtx®-5MS fused silica capillary column, computer matching of mass spectra with those of standards (NIST 2005 v.2.0 and Wiley Access Pak v.7, 2003 of GC-MS systems)[19].

#### 3. Results

The samples of *M. piperita* were extracted with methanol by Soxhlet extractor and the solvent was evaporated by rotary evaporator to give a gummy paste. The percentage (%) yields of extraction from the whole plants powder of *M. piperita* were hexane, ethyl acetate, chloroform, butanol and residual methanol fractions, respectively.

# 3.1. Physical properties

The different crude extracts from the whole plant have different colors. The hexane crude extract was deep brown in color, ethyl acetate was pale yellow in color, chloroform extract was deep orange and the butanol extract was blackish in color.

#### 3.2. Chemical composition of different extracts

The hexane crude extract from the whole plant of local *M. piperita* was analyzed by using GC-MS. It contains 22 different organic compounds, representing 1.52% of the total extract. The identification of the active chemical compounds was done by GC-MS listed in Table 1 according to their elution order on Rtx®-5MS capillary column. The major active ingredients that were identified in hexane extract (shown in Table 1 and Figure 1) are pulegone (1.11%), carvone (9.76%), verbenone (2.79%), bicycle [7.2.0] undec-4-ene, eupatorin (2.12%), (2E)-3,7,11,15-tetramethyl-2-hexadecen-1ol (4.68%), hexahydrofarnesyl acetone (2.04%), (2E)-3,7,11,15-tetramethyl-2-hexadecen-1-ol (1.53%), methyl 14-methylpentadecanoate (4.14%), methyl linolenate (8.15%), phytol (24.16%), butyl palmitate (6.90%), 2,6,10,15tetramethylheptadecane (2.90%), 1-teracosanol (1.31%), teracosanol (4.36%), isobutyl stearate (4.27%), eicosane, 7-hexyl- (11.47%) and nonacosane (8.19%).



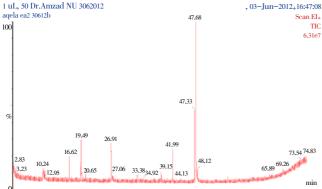
The ethyl acetate crude extract was analyzed by using GC-MS leading to the identification of a total of 7 different organic compounds using the same capillary column and conditions, representing 1.19% of the total extract from whole plant. The major chemical constituents that were identified in ethyl acetate crude extract by GC-MS (Figure 2 and Table 1) are carvone (6.47%), 3-eicosyne (2.85%), methyl 14-methylpentadecanoate (8.55%), (9E,12E,15E)-9,12,15-octadecatrien-1-ol (1.37%), sinensetin (1.42%), methyl linolenate (20.77%), phytol (54.18%) and methyl heptacosanoate (2.36%).

# Table 1

l composition of			

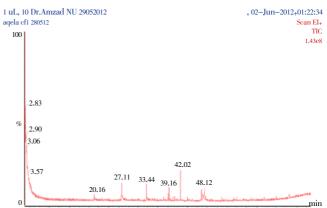
1 0	Retention time	Molecular	
Name of compounds	(min)	formula	Leave (%)
Hexane crude extract	(11111)	Iominuta	
Pulegone	16.510	C10H16O	0.8100
(+)-Carvone	16.690	$C_{10}H_{16}O$ $C_{10}H_{14}O$	7.0910
Verbenone	20.700	$C_{10}H_{14}O$ $C_{10}H_{14}O$	2.0290
Caryophyllene	23.900	$C_{10}H_{14}O$ $C_{15}H_{24}$	1.5750
3–Eicosyne	39.160	$C_{15}H_{24}$ $C_{20}H_{40}O$	3.4000
Hexahydrofarnesyl acetone	39.370	$C_{20}H_{40}O$ $C_{18}H_{36}O$	1.4880
3-Eicosyne	40.550	$C_{18}H_{36}O$ $C_{20}H_{40}O$	2.1240
Methyl 14–methylpentadecanoate (FA)	40.330	$C_{20}H_{40}O$ $C_{17}H_{34}O_2$	3.0110
Methyl linolenate (FA)	42.020	$C_{17}H_{34}O_2$ $C_{19}H_{32}O_2$	5.9230
Phytol	47.700	$C_{19}H_{32}O_2$ $C_{20}H_{40}O$	17.5520
Butyl palmitate (FA)	49.810		5.0130
Docosane	49.810	$C_{20}H_{40}O_2$ $C_{21}H_{44}$	2.1070
Octadecanol acetate	49.990 50.480	$C_{21}H_{44}$ $C_{24}H_{50}O$	0.9540
Tricosane			
	52.910	C <sub>24</sub> H <sub>50</sub>	3.1680
Isobutyl stearate (FA)	55.280	$C_{22}H_{44}O_2$	3.1050
Tetracosane	55.550	C <sub>26</sub> H <sub>54</sub>	8.0750
Pentacosane	58.110	C <sub>29</sub> H <sub>60</sub>	7.0620
Hexacosane	60.560	$C_{22}H_{44}O_2$	8.3330
Heptacosane	62.920	C <sub>28</sub> H <sub>48</sub>	5.7440
Octacosane	65.200	C <sub>34</sub> H <sub>50</sub> O	5.9510
Nonacosane	67.430	C <sub>35</sub> H <sub>52</sub>	3.4180
Triacontane	69.560	$C_{32}H_{54}O_2$	3.0740
Ethyl acetate extract	16 (24	сцо	6 7000
Carvone	16.624	$C_{10}H_{14}O$	6.7000
3-Eicosyne	39.149	C <sub>20</sub> H <sub>38</sub>	2.9600
Methyl 14-methylpentadecanoate (FA)	41.995	$C_{17}H_{34}O_2$	8.8600
Sinensetin	44.131	C <sub>18</sub> H <sub>32</sub> O	1.4270
Methyl linolenate (FA)	47.327	$C_{19}H_{32}O_2$	21.5080
Phytol	47.682	$C_{20}H_{40}O$	56.0940
Methyl isoheptadecanoate (FA)	48.117	$C_{28}H_{56}O_2$	2.4500
Chloroform extract	20.150	C II O	10.0402
Eupatorin	39.159	$C_{20}H_{40}O$	18.0483
Methyl 14-methylpentadecanoate (FA)	42.020	$C_{17}H_{34}O_2$	40.7430
Methyl linolenate (FA)	47.333	C <sub>18</sub> H <sub>31</sub> OCl	14.9235
Phytol	47.700	$C_{18}H_{36}O_2$	11.7032
Methyl isoheptadecanoate (FA)	48.123	$C_{20}H_{40}O_2$	14.5820
Butanol extract		6.11.0	60.4000
o-Tolualdehyde	15.744	C <sub>8</sub> H <sub>8</sub> O	60.1880
Methyl 14-methylpentadecanoate (FA)	42.035	$C_{17}H_{34}O_2$	39.8110
Methanol crude extract			
Pulegone	16.514	C <sub>10</sub> H <sub>16</sub> O	7.2040
Carvone	16.704	$C_{10}H_{14}O$	40.4600
Eugenol	21.416	$C_{10}H_{12}O_2$	9.0740
3-Eicosyne	39.170	$C_{17}H_{34}O_2$	7.7970
Methyl 14-methylpentadecanoate (FA)	42.030	C <sub>18</sub> H <sub>31</sub> OCl	18.0740
Methyl linolenate (FA)	47.347	C <sub>17</sub> H <sub>34</sub> O <sub>2</sub>	17.3900





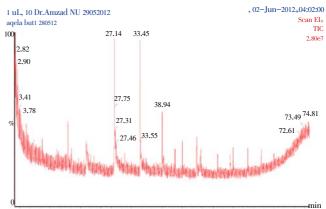
3.85 8.85 13.85 18.85 23.85 28.85 33.85 43.85 48.85 53.85 58.85 63.85 68.85 73.85 Figure 2. A typical gas chromatogram of the chemical constituents of ethyl acetate crude extract

The chloroform crude extract was analyzed by using GC-MS resulting in the identification of 4 different organic compounds using the same capillary column and conditions, representing 0.88% of the total extract. The chemical constituents that were found in chloroform plant crude extract (Figure 3 and Table 1) are identified as (2E)-3,7,11,15tetramethyl-2-hexadecen-1-ol (20.44%), methyl 14methylpentadecanoate (46.14%), linoleoyl chloride (16.90%) and methyl isoheptadecanoate (16.51%).



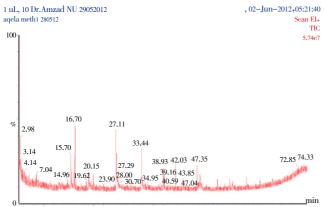
3.85 8.85 13.85 18.85 23.85 28.85 33.85 43.85 48.85 53.85 58.85 63.85 68.85 73.85 Figure 3. A typical gas chromatogram of the chemical constituents of chloroform crude extract.

The butanol plant crude extract was analyzed by using GC-MS, leading to the identification of 2 different organic compounds, representing 0.69% of the total extract. The major chemical constituents found in butanol crude extract (Figure 4 and Table 1) are o-tolualdehyde (27.97%) and methyl 14-methylpentadecanoate (18.50%).



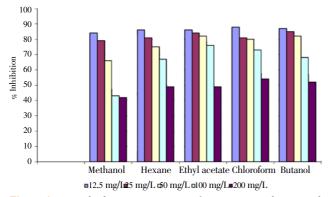
8.85 13.85 18.85 23.85 28.85 33.85 43.85 48.85 53.85 58.85 63.85 68.85 73.85 3.85 Figure 4. A typical gas chromatogram of the chemical constituents of butanol crude extract.

Finally the methanol crude extract was analyzed using GC-MS leading to identification of 5 different organic compounds representing 2.13% of the total extract. The major chemical constituents identified in methanol plant crude extract (Figure 5 and Table 1) are pulegone (6.00%), carvone (33.72%), eugenol (7.56%), methyl 14-methylpentadecanoate (15.06%) and lineoleoyl chloride (14.49%).



<sup>0</sup>3.85 8.85 13.85 18.85 23.85 28.85 33.85 43.85 48.85 53.85 58.85 63.85 68.85 73.85 Figure 5. A typical gas chromatogram of the chemical constituents of methanol crude extract.

The free radical scavenging activity of different polarities crude extracts was tested through DPPH method and the results are presented in Figure 6. The antioxidant activity in their interaction depends on oxidative free radicals. The mechanism of DPPH method is that the antioxidants react with the stable free radical *i.e.*,  $\alpha$ , $\alpha$ -diphenyl- $\beta$ picrylhydrazyl (deep violet colour) and convert it to  $\alpha,\alpha$ diphenyl- $\beta$ -picrylhydrazine with discoloration. The gradual discoloration indicates the scavenging capacities of the plant crude extracts. In the present study, the five different plant crude extracts extracted from locally grown M. piperita were able to decolorize DPPH and the free radical scavenging capacities of the plant crude extracts were found to be in the order of chloroform> butanol> ethyl acetate extract> hexane extract> methanol (Figure 6). The present experiment used DPPH as a control. It has been found that all aromatic and aliphatic organic compounds such as aromatic hydrocarbon, cysteine, glutathione, ascorbic acid, tocopherol, polyhydroxy and other aromatic amines such as *p*-phenylene diamine, *p*-aminophenol *etc.*, gradually reduce and decolorize by  $\alpha,\alpha$ -diphenyl- $\beta$ -picrylhydrazyl revealing their free radical ability<sup>[19,20]</sup>. In this study, it appears that perhaps the five different crude extracts from local M. piperita possess strong hydrogen donating capabilities to act as antioxidants.



**Figure 6.** Free radical scavenging activity of peppermint crude extracts by DPPH methods.

#### 4. Discussion

The different crude extracts of local M. piperita for respective bioactive compounds can be chosen on the basis of the above GC-MS analysis. Phytol is a major terpenoid found to be the most abundant in the hexane, ethyl acetate and chloroform crude extract. Hexane crude extract contains a lot of normal and cyclic hydrocarbons with high molecular weight. Ethyl acetate crude extract also contains methyl linolenate, phytol and some other aliphatic and aromatic chemical compounds such as flavonoids. Chloroform crude extract contains flavonoids eupatorin, (2E)-3,7,11,15-tetramethyl-2-hexadecen-1-ol and methyl 14-methylpentadecanoate; butanol crude extract contains o-tolualdehyde and methyl 14-methylpentadecanoate and methanol crude extract contains carvone, methyl 14-methylpentadecanoate and lineoleoyl chloride. All the major chemical compounds have previously been reported from a number of other plant species<sup>[19,20]</sup>. The major chemical compounds identified from different crude extracts by GC-MS are basically biologically active molecules reported in different plants crude extracts by the researchers<sup>[5,8,10-18]</sup>. All the predominant compounds in this plant crude extracts such as sinensetin, euparotin etc. are considered to be a part of plants' defense system, and as such have been included in a large group of protective molecules found in plants named 'phytoanticipins' or 'phytoprotectants'[17-19]. Therefore, the identified chemical compounds from various crude extracts of *M. piperita* by GC-MS might have some ecologically significant role. So far as known, this report is the first report to investigate in vitro antioxidant activities as well as chemical composition of different crude extracts of locally grown M. piperita using GC-MS. Hence, the entire *M. piperita* plant could be of use as a good source of antioxidant. Further studies are needed for the isolation and identification of individual bioactive compounds from the plant crude extracts of *M. piperita* and also in vivo studies are needed for better understanding of their mechanism of action as antioxidant.

# **Conflict of interest statement**

We declare that we have no conflict of interest.

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