

Review Article

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Therapeutic Potential of Secoisolariciresinol Diglucoside: A Plant Lignan

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ABSTRACT

Secoisolariciresinol diglucoside (SDG) is a plant lignan mainly found in dietary food and various plants. It belongs to a bioactive polyphenolic chemical class. SDG and its metabolites (mammalian enterolignan) are having various pharmacological activities, viz., antioxidant, partial agonist to estrogen receptor and inhibitor of tyrosine kinase and topoisomerase. Although, human studies are limited, its pharmacological actions explain its use in diabetes, atherosclerosis, breast cancer, colon cancer, prostate cancer and in cardiovascular disease.

Keywords: Secoisolariciresinol diglucoside, plant lignan, polyphenols, antioxidant, tyrosine kinase inhibitor, topoisomerase inhibitor.

INTRODUCTION

Plant lignans are biophenolic compounds and was first identified in 19th Century from woody tissues of trees. Several hundreds of lignans have been documented since then in roots, stem, cereals, oilseeds, nuts, legumes and fruits. ^[11] Nowadays, with the growing interest towards nutraceuticals, plant lignans are becoming important therapeutically active class of compounds because of their putative beneficial health effect such as antitumor, antioxidant, both estrogenic and antiestrogenic activity ^[2] and protection against coronary heart disease. ^[3] This plant lignans can be converted by intestinal bacteria into the mammalian lignan such as enterolignans, enterodiol and enterolactone. ^[4-6]

Secoisolariciresinol diglucoside (SDG) and matairesinol are the major lignans with traces of pinoresinol, lariciresinol and isolariciresinol found in roots, stem, cereals, oilseeds, nuts, legumes and fruits. ^[1-2] Flax seed (*Linum usitatissimum* L.) is the richest dietary source of lignans, with SDG as a major compound (% yield being 0.37%), besides that sesame seed, pumpkin seeds, cereals (triticle and wheat), leguminous plant (lentils, soyabeans), fruits (pears, prunes) and certain vegetables (garlic, asparagus, carrot) also contain traces of lignans, but concentration in flaxseed is about 1000 times as high as found in other food sources. ^[7]

PHARMACOKINETICS OF PLANT LIGNANS

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Table I: Lignans in selected food [8-9]		
Food and foods group		Secoisolariciresinol diglucoside content (µg/g)
Seeds	Flaxseed	3699
	Sunflower	6.1
	Caraway	2.21
	Pumpkin	213.7
	Soybean	2.73
Legumes	Peanut	3.33
	Pigeon pea	0.5
	Urad Dahl bean	2.4
Nuts	Walnut	1.63
Berries	Almond	1.07
	Blackberry	37.1
	Lingberry	15.1
	Strawberry	12.1
	Cranberry	15.1
	Red currant	1.6
Cereals	Oatmeal	0.1
	Oat bran	0.24
	Rye meal, whole grain	0.5
	Rye bran	1.32
Vegetables	Broccoli	4.14
	Garlic	3.79
	Carrots	1.92
Coffee and	Arabica coffee (instant)	7.16
Tea	Green tea	24.6
	Black tea	15.9

Bioavailability can be defined as the fraction of the ingested plant lignans that is absorbed and can be used for metabolic process (internal exposure) and storage in the body. Following metabolism of plant lignans in the human colon, the metabolites, enterodiol and enterolactone reach the circulation and target tissues. As metabolism is extensive, enterodiol and enterolactone might be more important for potential health effect. The bioavailability of lignans can be

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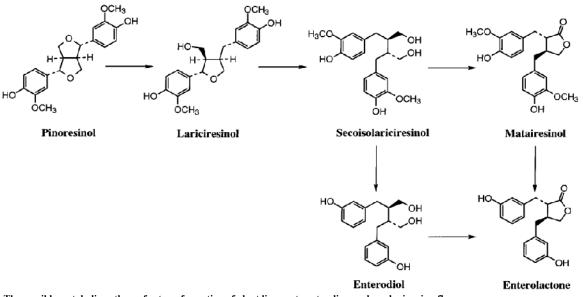
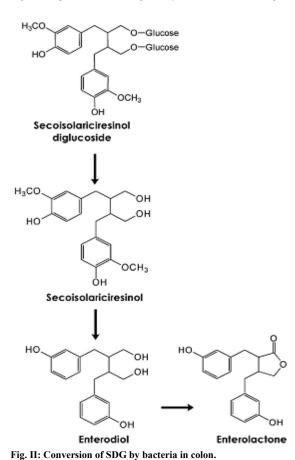
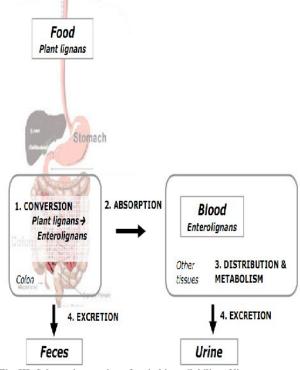


Fig. I: The possible metabolic pathway for transformation of plant lignans to enterolignans by colonic microflora.



form (aglycone or conjugated) of plant lignan, chronic exposure, and other host related factor like age and gender. $^{[10]}$





determined by several process: (a) Conversion of plant lignans to enterolignans in the human colon; (b) Absorption of enterolignans from the colon, which determines whether or not enterolignans become available in blood circulation; (c) Distribution and metabolism, which determines whether metabolites reach the target tissues where they can be have an effect or further metabolized. Finally, (d) Enterolignans are excreted from the body via faeces or urine (Fig. III).

Various factors may influence lignan bioavailability such as intestinal microflora, antibiotic use, food matrix, type and

After consumption of SDG, a small fraction is absorbed as such in the small intestine ^[11], and excreted in urine. ^[12-13] However, the largest fraction of the SDG is transported to colon and metabolized by intestinal flora (Fig. II). The importance of microflora in the metabolism of SDG was studied in germfree rats. ^[14-15] Although the intestinal bacteria play a crucial role in lignan metabolism, few studies had been published that identify organism involved in lignan breakdown. Two anaerobes (*Peptostreptococcus* and *Eubacterium*) catalyze the demethylation and dehydroxylation of SDG. ^[16] Recently, two microorganisms

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Peptostreptococcus productus and *Eggerthella inta* were isolated that were able to demethylate and dehydroxylate SDG and pinoresinol. However, concentrations of enterolactone are usually higher than enterodiol in humans. ^[17] One bacterial strain (ED-Mt61/PYCt-s6) was identified to be responsible for transformation of enterodiol to enterolactone. ^[18]

SDG and majority of lignan converted to enterolignans are absorbed by the large intestine into the blood stream or directly excreted via faeces. Plasma enterodiol and enterolactone circulate either as glucouronide and sulfate conjugates or as free forms. ^[19] They are excreted via urine or bile, in urine; enterodiol and enterolactone are excreted in conjugated forms; primarily as monoglucouronides (85 and 95% respectively) with small percentage being excreted as monosulfates (2-10%) and free aglycones (0.3-1%). ^[20-21] Conjugated enterolignans that are excreted via bile can undergo enterohepatic circulation (i.e. excreted through bile duct into the intestinal tract, further metabolized in the colon, and reabsorbed from the large intestine into the bloodstream). ^[15, 22] The mean residence time and half lives measured after single dose of SDG (0.9 mg SDG/kg bodyweight) in healthy men and women indicates that enterolignan accumulate in plasma when consumed 2-3 times a day and reach steady state. [23]

PHARMACOLOGICAL ACTIVITIES OF SECOISOLARICIRESINOL DIGLUCOSIDE (SDG)

Antioxidant activity: The plant lignans demonstrated extreme radical scavenging activity. In both lipid and aqueous, *in-vitro* model systems SDG and its metabolites appeared as antioxidant. All three lignans significantly inhibited the linoleic acid peroxidation at 10 and 100 μ M over a 24-48 h of incubation at 40°C. ^[24-25] However, it has been demonstrated that enterodiol and enterolactone were not effective in preventing H₂O₂ induced DNA damage in HT-29 cells and enterolactone did not reduced intracellular oxidative stress at similar concentration. ^[26]

Antiatherosclerotic effect: One of the most etiological reasons of atherosclerosis is the release of inflammatory mediators such as interleukin (IL-1), tumor necrosis factor (TNF), leucotriene B_4 (LTB₄). These all mediators are known to stimulate polymorphonuclear leukocytes (PMNLs) and monocytes to produce oxidative free radicals (OFRs). As the flaxseed is the richest source of the SDG, ^[6] supplementation of flaxseed was reduce the level of OFRs and hence, prevent the development of hypercholesterolemic atherosclerosis and aortic atherosclerosis by 46% markedly without lowering serum cholesterol. ^[27-30]

Anticancer activity: Secoisolariciresinol diglucoside and other lignans are phytoestrogen, because of their potential estrogenic and antioxidant activity had been studied for various cancer protective mechanisms. Supplementation of richest lignan source flaxseed reduced the epithelial cell proliferation by 38.8 - 55.4% and nuclear aberration by 58.8 - 65.9% in female rat. ^[31-32]. Enterolignans can bind to estrogen receptor α and β ^[33] and block or antagonize the effect of estrogen in some tissues. ^[34] Extensive work had been done to study the chemoprotective action of SDG and enterolignans on mammary cells. Supplementation of SDG reduced the risk of breast cancer, and showed antiproliferative effect on the breast, positive effect on lipoprotein profile and bone density in post menopausal

women. [24, 35-38] It showed the suppression of mammary tumorigenesis by its partial estrogenic activity, antioxidant activity or reduction of plasma insulin like growth factor-1. ^[38-41] Besides, its antioxidant and partial estrogenic activity of SDG and enterolignans also affect to beta glucouronidase activity, which may be the cause of protective effect against colon cancer. It has been observed that pretreatment of flaxseed decrease the risk of colon carcinogenesis, with reduced total number of aberrant crypts and foci significantly by 41-53% and 48-57% respectively. ^[38, 42-43] Metabolites of SDG also appeared to influence steroid metabolism in- vitro, not only by acting on steroid receptor but also by steroid metabolism, for e.g. sex hormone binding globulin synthesis ^[44-45] 5α -reductase and 17- β hydroxyl-steroid dehydrogenase. ^[46] On the virtue of these all property and inhibition of tyrosine kinase and topoisomerase contribute the lower incidence of prostate cancer. [47]

Anti-diabetic activity: It has been demonstrated that SDG prevented development of diabetes mellitus by 75%. The reactive oxygen species play an important role in development of debates mellitus (DM) therefore it was suggested that the antioxidant activity may be playing role for its antidiabetic activity. ^[48-49]

Effect on Cardiovascular system: There are several mechanisms by which SDG protect against cardiovascular diseases. It appeared beneficial role in endotoxic shock. ^[25, 50] Apart from reducing cholesterol level it also induced angiogenesis mediated cardioprotection by increased neovascularization in the peri-infarct zone, leading to less ventricular remodeling. Thus, SDG is a great clinical potential for treatment of ischemic heart disease. ^[51]

REFERENCES

- 1. Willtor SM, Smeds AI, Holmbom BR. Chromatographic analysis of Lignans (Review). J Chromatography A. 2006; 1112:64-77.
- Sicilia T, Niemeyer HB, Honig DM, Metzler M. Identification and stereochemical characterization of lignans in flaxseed and pumpkin seeds. J Agri Food Chem. 2003; 51:1181-1118.
- Smeds AI, Eklund PC, Sjoholm RE, Willfer SM, Nishibe S, Deyama T. Quantification of a broad spectrum of lignans in cereals, oil seeds and nuts. J Agri Food Chem. 2007; 55:1337-1346.
- Axelson M, Sjoevall J, Gustafsson BE. Origin of lignans in mammals and identification of precursor from plants. Nature.1982; 298:659-660.
- Borriello SP, Setchell KD, Axelson M, Lawson AM. Production and metabolism of lignans by the human faecal flora. J Appl Bacteriol. 1985; 58:37-43.
- Mazur WM, Uehara M, Wahala K, Adlercreutz H. Phytoestrogen content of berries and plasma concentration and urinary excretion of enterolactone after a single strawberry-meal in human subjects. Br J Nutrition. 2000; 83:381-387.
- Milder IEJ, Arts ICW, Putte B, Venema DP, Hollman PCH. Lignan content of Dutch plant foods: a database including lariciresinol, pinoresinol, secoisolariciresinol and matairesinol. Br J Nutrition. 2005; 93:393.
- Adlercreutz H, Mazur W. Phytoestrogen and western disease. Ann. Med. 1997; 29:95-120.
- 9. Mazur W. Phytoestrogen contents in food. Baillieres Clin Endocrinol Metab. 1998; 12:729-42.
- Rowald I, Faughnan M, Hoey L, Wahala K. Williamson G, Cassidy A. Bioavailability of Phytoestrogens. Br J Nutr. 2003; 89(Suppl): 838-852.
- Penalvo J L, Nurmi T, Haajnen K, Al-Maharik N. Determination of lignans in human plasma by liquid chromatography with coulometric electrode array detection. Anal Biochem. 2004; 332:384-393.
- Nurmi T, Voutilainen S, Nyyssonen K, Adlercreutz H, Salonen JT. Liquid chromatography method for plant and mammalian lignans in human urine. J Chromatography B. 2003; 798:101-110.
- 13. Bannwart C, Adlercreutz H, Wahala K, Brunow G, Hase T. Detection and identification of the plant lignans lariciresinol,

isolariciresinol and secoisolariciresinol in human urine. Clin Chimica Acta.1989; 180:293-301.

- 14. Bowey E, Adlercreutz H, Rowland I. Metabolism of isoflavones and lignans by the gut microflora: a study in germ free and human flora associated rats. Food Chem Toxicol. 2003; 41:631-636.
- Axelson M, Setchell KD. The excretion of the lignans in rats evidence for an intestinal bacterial source for this new group of compounds. FEBS Lett.1981; 123:337-342.
- Wang LQ, Meselhy MR, Li Y, Quin GW, Hattori M. Human intestinal bacteria capable of transforming secoisolariciresinol diglucoside to mammalian lignans, enterodiol and enterolactone. Chem Pharm Bull. 2000; 48:1606-1610.
- Clavel T, Henderson G, Alpert CA, Philippe C, Rigotter-Gois L, Dore J, Blaut M. Intestinal bacteria communities that produce active estrogen like compounds enterodiol and enterolactone in humans. Appl Environ Microbiol. 2005; 71:6077-6085.
- Clavel T, Henderson G, Engest W, Dore J, Blaut M. Phylogeny of human intestinal bactaria that activate the dietary lignans secoisolariciresinol diglucoside. FEMS Microbiol Ecol. 2006; 55:471-478.
- Adlercreutz H, Fotsis T, Lampe J, Wahala K, Makela T, Brunow G, Hase T. Quantitative determination of lignans and isoflavonoids in plasma of omnivorous and vegetarian women by isotope dilution gas chromatography – mass spectrometry. Scandinavian J Clin Lab Invest. 1993; 215:5-18.
- Adlercreutz H, Vanderwildt J, Kinzel J, Attalla H, Wahala K, Makela T, Hase T, Fotsis T. Lignans and isoflavonoids conjugates in human urine. J Steroid Biochem Mol Biol. 1995; 25:97-103.
- Axelson M, Setchell KD. Conjugation of lignans in human urine. FEBS Letters. 1980; 122:49-53.
- Knudsen KEB, Serena A, Kjaer AKB, Tetens L, Heinonen SM, Nurmi T, Adlercreutz H. Rye bread in the diet of pigs enhances the formation of enterolactone and increase its levels in plasma, urine and feces. J Nutrition. 2003; 133:1368-1375.
- Anneleen K, Iija CWA, Tom BV, Peter CHH. Pharmacokinetics of enterolignans in healthy men and women consuming a single dose of secoisolariciresinol diglucoside. J Nutrition. 2005; 135:795-801.
- Kitts DD, Vyan YV, Wijewickreme AN, Thompson LU. Antioxidant property of flaxseed lignans SDG and its mammalian lignan metabolites enterodiol and enterolactone. Mol Cell Biochem. 1999; 202:91-100.
- Prasad K. Antioxidant activity of secoisolariciresinol diglucoside derived metabolites enterodiol and enterolactone. International J Angiol. 2000; 9:220-225.
- Pool-Zobel BL, Adlercreutz H, Glei M, Liegibel UM, Sittlingon J, Rowland I, Wahala K, Rechkemmer G. Isoflavonoid and lignans have different potential to modulate oxidative genetic damage in human colon cells. Carcinogenesis. 2000; 21:1247-1252.
- Ogborn MR, Nitschmann E, Weiler HA. Flaxseed ameliorates intestinal nephritis in rat polycystic kidney disease. Kidney Int. 1919; 55:417-423.
- Prasad K. Dietary flaxseed in prevention of hypercholesterolemic atherosclerosis. Atherosclerosis. 1997; 132:69-76.
- Prasad K. Reduction of serum cholesterol and hypercholesterolemic atherosclerosis in rabbit by secoisolariciresinol diglucoside isolated from flaxseed. Circulation. 1999; 99:1355-1362.
- Prasad K. Hypercholesterolemic and antiatherosclerotic effect of flax lignan complex isolated from flaxseed. Atherosclerosis. 2005; 179:269-275.
- Serraino M, Thompson LU. The effect of flaxseed supplementation on early risk markers for mammary carcinogenesis. Cancer Lett. 1991; 60:135-142.

- 32. Serraino M, Thompson LU. The effect of flaxseed supplementation on the initiation and promotional stages of mammary tumorigenesis. Nutrition Cancer. 1992; 17:153-159.
- Muller SO, Simon S, Chae K, Metzler M, Korach KS. Phytoestrogens and their human metabolites show distinct agonistic and antagonistic properties on estrogen receptor α and β in human cells. Toxicol Sci. 2004; 80:14-25.
- Wang LQ. Mammalian phytoestrogens: enterodiol and enterolactone. J Chromatography B. 2002; 777:289-309.
- Haggans CJ, Olson BA, Thomul W. Effect of flaxseed consumption on urinary estrogen metabolites in postmenopausal women. Nutrition Cancer. 1919; 33:188-195.
- Tou JCL, Thompson LU. Exposure to flaxseed or its lignans components during different developmental stages influences rat mammary gland structures. Carcinogenesis. 1999; 20:1831-1835.
- Tou JCL, Chen J, Thompson LU. Flaxseed and its lignan precursor, secoisolariciresnol diglucoside affect pregnancy outcome and reproductive development in rats. J Nutrition. 1998; 128:1861-1868.
- Sung MK, Lautens M, Thompson LU. Mammalian lignan inhibit the growth of estrogen-independent human colon tumor cells. Anticancer Res. 1998; 18:1405-1408.
- Rickard SE, Yuan YV, Chen J, Thompson LU. Dose effect of flaxseed and its lignan on N-methyl-N-nitrosourea induced mammary tumorigenesis in rats. Nutrition Cancer. 1999; 35:50-57.
- Thompson LU, Seidl MM, Rickard SE, Orcheson LJ, Fong HHS. Antitumorigenic effect of mammalian lignan precursor from flaxseed. Nutrition Cancer. 1996; 26:159-165.
- Thompson LU, Rickard SE, Orcheson LJ, Seidl MM. Flaxseed and its lignan and oil component reduce mammary tumor growth at late stage of carcinogenesis. Carcinogenesis. 1996; 17:1373-1376.
- Serraino M, Thompson LU. Flaxseed supplementation and early markers of colon carcinogenesis. Cancer Lett. 1992; 63:159-165.
- Jenab M, Thompson LU. The influence of flaxseed and lignans on colon carcinogenesis and beta-glucuronidase activity. Carcinogenesis. 1996; 17:1343-1348.
- Schottner M, Gansser D, Spiteller G. Interaction of lignan with human sex hormone binding globulin (SHBG). J Biosci. 1997; 52:834-843.
- Adlercreutz H, Hockerstedt K, Bannwart C. A effect of dietary components, including lignan and phytoestrogens on enterohepatic circulation and liver metabolism of estrogen and on sex hormone binding globulin (SHBG). J Steroid Biochem Mol Biol. 1987; 27:1135-1144.
- 46. Evans BAJ, Griffliths K, Morton MS. Inhibition of 5α -reductase in genital skin fibroblast and prostate tissue by dietary lignans and isoflavonoids. J Endocrinol. 1995; 147:295-302.
- Tou JCL, Chen J, Thompson LU. Dose, timing and duration of flaxseed exposure alter reproductive indices and sex hormone in rats. J Toxicol Environ Health. 1990; 56:555-570.
- Prasad K. Oxidative stress as a mechanism of diabetes BB Prone rats: effect of SDG. Mol Cell Biochem. 2000; 209:89-96.
- Prasad K, Mantha SV, Muir AD, Westcott ND. Protective effect of secoisolariciresinol diglucoside against streptozotocin induced diabetes and its mechanism. Mol Cell Biochem. 2000; 206:141-150.
- Pattanik U, Prasad K. Oxygen free radicals and endotoxic shock: effect of flaxseed. J Cardiovascular Pharmacol Ther. 1998; 3:305-318.
- Penumathsa SV, Koneru S, Mahesh TV, Zhan L, Prasad K. Secoisolariciresinol diglucoside: Relevance to angiogenesis and cardioprotection against ischemia-reperfusion injury. J Pharmacol Exp Ther. 2007; 320:951-959.