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## Bioaccumulation of cadmium in gills and muscles of shellfish from Pulicat lake, Tamil Nadu, India

Kalyanasundaram Dhinamala<sup>1</sup>, Munuswamy Pushpalatha<sup>1</sup>, Mohamed Meeran<sup>2</sup>, Subramanian Arivoli<sup>3</sup>, Samuel Tennyson<sup>1</sup>, Rajasingh Raveen<sup>1\*</sup>

<sup>1</sup>Department of Zoology, Madras Christian College, Chennai 600 059, Tamil Nadu, India

<sup>2</sup>Department of Zoology, Hajee Karutha Rowther Howdia College, Uthamapalayam 625 533, Tamil Nadu, India

<sup>3</sup>Department of Zoology, Thiruvalluvar University, Vellore 632 115, Tamil Nadu, India

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### ABSTRACT

**Objective:** To evaluate the presence of heavy metal cadmium in six species of shellfish at Pulicat Lake, Tamil Nadu, India.

**Methods:** Six species of shellfish, *Fenneropenaeus indicus*, *Fenneropenaeus monodon*, *Fenneropenaeus semisulcatus*, *Scylla serrata*, *Clibanarius longitarsus* and *Meretrix casta* (*M. casta*) in Pulicat lake, Tamil Nadu, India were analysed for the presence of cadmium in the gills and muscles from January 2011 to December 2012.

**Results:** The results showed seasonal variations in the uptake of cadmium. Very high accumulation of cadmium was found in the gills and muscles of *M. casta* during post monsoon, summer, premonsoon and monsoon. The corresponding values of cadmium present in the gills of *M. casta* were 1.59, 1.56, 1.48 and 1.46 µg/g in 2011 and 1.16, 1.25, 1.15 and 1.14 µg/g in 2012. Whereas for muscles, they were 1.14, 0.11, 0.96 and 0.80 µg/g in 2011 and 0.49, 0.34, 1.05 and 1.20 µg/g in 2012.

**Conclusion:** The results of the present study has shown that the accumulation of cadmium found in the gills and muscles were high in *M. casta* when compared to other species of shellfish. Thus, the consumption of the shellfish is safe, but does not exclude bioaccumulation risk in their meat. This present study has highlighted the need for estuarine biomonitoring to avoid possible contamination of shellfish and its consumers. The overall scenario of the shellfish accumulating high levels of cadmium indicates that the Pulicat lake is polluted with undesirable elements and the risk of consuming the meat of shellfish by man and other carnivores may lead to their toxicity. Stringent control measures are necessary to control the pollution of this precious lake to reduce the bioaccumulation of toxic metals in organisms.

## 1. Introduction

The estuaries are considered to be essential components of the marine ecosystem and estuary is a zone which is transitional between the riverine and marine environments. Estuaries effectively trap nutrients to provide resources to aquatic organisms and to humans for commercial and entertainment activities. Oysters and clams prefer this area for their settlement whereas other fin fishes and shell fishes also prefer estuaries to act as their nursery grounds[1].

Heavy metals are present in marine environments due to natural developments and man-made causes[2,3]. The pollution of natural water bodies by trace elements affects the aquatic organisms and creates environmental and human health hazards[4,6]. Heavy metals such as

arsenic, cadmium, chromium, copper, lead, mercury, tin and zinc are among the most dangerous pollutants in the marine environment[7-9]. Heavy metal accumulation in water bodies surrounding urban and industrial areas are mainly due to effluents that are discharged from a number of factories involved in the production of biocides, pigments and paints, ore processing, electroplating, tanning and textile dyeing[10]. Heavy metals are one of the most serious pollutants in the environment due to their toxicity, persistence and ability to concentrate along the food chain[11]. Rapid growth in human population, intensive agriculture and industrial activities have eventually led to increased amount of pollutants especially heavy metals that threaten the estuarine environment[12]. Most organisms require minute quantities of essential heavy metals viz., copper, iron, manganese and zinc for essential processes especially for growth[12,13]. When certain limits are exceeded, it becomes toxic to the organisms[11]. Certain metals such as cadmium, chromium and lead are non-essential and exhibit toxicity even at low concentrations[14]. The sediment dwelling biota are exposed to heavy metals due to their ingestion of surrounding waters, sediments and through the food chain[15]. High levels of

\*Corresponding author: Rajasingh Raveen, Department of Zoology, Madras Christian College, Chennai 600 059, Tamil Nadu, India.

Tel: +91 9840463064

E-mail: raveenraja2002@gmail.com

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heavy metals in sediments indicate anthropogenic pollution than due to natural causes[16,17]. Certain marine invertebrates are considered as bioindicators to indicate the presence of heavy metals as the contamination in these organisms depicts a direct relationship to that of the environment[18]. Consumption of bivalves can be severely limited due to the presence of heavy metals since they cause injurious effects to human health[19]. A precise estimation of heavy metals in the environment and in the biota will provide the degree of metal contamination in that particular place[20]. Availability of cadmium may vary due to natural and anthropogenic causes in different habitats[21,22]. In view of the aforementioned factors, the current study was undertaken to evaluate the variation of the trace metal cadmium in the gill and muscle tissues of six species of shellfish viz., *Fenneropenaeus indicus* (*F. indicus*), *Fenneropenaeus monodon* (*F. monodon*), *Fenneropenaeus semisulcatus* (*F. semisulcatus*), *Scylla serrata* (*S. serrata*), *Clibanarius longitarsus* (*C. longitarsus*) and *Meretrix casta* (*M. casta*) present in Pulicat lake, Tamil Nadu, India.

## 2. Materials and methods

### 2.1. Study area

Pulicat lake ( $13^{\circ}24' - 13^{\circ}47' \text{ N}$ ,  $80^{\circ}03' - 80^{\circ}18' \text{ E}$ ) is the second largest brackish water body of India with an area of 18440 hectares and is located 40 km north of Chennai. The length of this lake is about 60 km and varies in breadth (0.2 to 17.5 km). Pulicat lake is drained by four rivers, the Swarnamukhi, the Kalangi, the Aranair and the Royyala Kalava apart from many minor inflows. Industrial and domestic waste are brought into this lake by the Buckingham canal and finally to the Bay of Bengal[23]. Local climate, riverine inflow and the neritic waters from the Bay of Bengal influence the hydrological characters of Pulicat lake. Many euryhaline species are present in this lake which act as breeding grounds for many organisms and certain fishes[24]. Untreated effluents from industries and urban areas are considered to be point sources of pollution[23,25,26].

### 2.2. Collection of specimens

Six shellfish species viz., *F. indicus*, *F. monodon*, *F. semisulcatus*, *S. serrata*, *C. longitarsus* and *M. casta* were collected from Pulicat lake, Tamil Nadu, India on a monthly basis for a period of two years from January 2011 to December 2012. The collected organisms were brought to the laboratory in an ice box and were stored at  $4^{\circ}\text{C}$  until analyses. The organisms were thoroughly washed with running tap water to eliminate mud and other debris and were subsequently rinsed with double-distilled water. Rust free stainless steel kit was used to dissect the animal. Care was taken to avoid external contamination of the samples.

### 2.3. Determination of metals in animals

The gills and muscles were used to estimate cadmium content. The analysis was carried out using the method suggested by Watting[27]. Analytical grade reagents were used. For analysing cadmium, the samples were oven dried at  $60^{\circ}\text{C}$  for 24 h. The dried sample (0.5 g) was taken and ground with a mortar and pestle. Using nitric and perchloric acid (3:1), the ground samples were digested. After adding the acids, the samples were kept in the hot plate at  $120^{\circ}\text{C}$  until white

residues were formed. Finally the residue was dissolved in 10 mL of distilled water and then filtered. The filtered sample was aspirated into the atomic absorption spectrophotometer and the reading was recorded. The solution was then diluted and filtered through a 0.45  $\mu\text{m}$  nitrocellulose membrane filter. Determination of cadmium in samples was carried out by inductively coupled plasma atomic emission spectroscopy (Optima 2100 DV, Perkin-Elmer, USA).

## 3. Results

Bioaccumulation is the net uptake of chemicals through probable routes such as diet and respiration, thereby resulting in cytotoxicity. Most studies on bioaccumulation of trace metals in the marine environment have focused on animals nearer to the lower end of the food web, since these animals could be directly utilized for consumption by both human and livestock, and can serve to transfer heavy metals to carnivores[28].

Industrialization and exploitation of mineral resources around the globe has caused heavy accumulation of toxic trace metals in the environs. Increasing levels of numerous toxic heavy metals in the environment has evoked serious concern among the masses. The transfer of heavy metals through the food chain is attributed to the migration, concentration and solubility in marine waters. Bottom dwellers such as polychaetes and molluscs are more prone to accumulation of high concentrations of heavy metals[10]. Trace metal accumulation in the various tissues of aquatic organisms depends on the route of entry which may be from the water medium or from other chemical sources. Therefore, accumulation of trace metals in the tissues of such aquatic fauna is due to their presence in water. The gills function as the major pathway for intake of heavy metals from the aquatic medium by the external mucous membrane and also due to specific characters of the cell wall[29-31].

The accumulations of cadmium in the gills of shellfish are exhibited as seasonal as well as species specific variations. During post monsoon, the gills and muscles tend to accumulate a high amount of cadmium in *M. casta* whereas for other organisms it is highly reduced in both the years. During summer, there is moderate uptake of cadmium by all the shellfish species except *F. monodon* and *C. longitarsus* in both the years. Premonsoon exhibits a rise in the accumulation of cadmium in all the six organisms for both years, especially *M. casta* which shows a very high build-up of cadmium in both gills and muscles. Similarly in monsoon, there is an overall increase in the cadmium accumulation in the gills and muscles for both the years. Among the six shellfish species studied, *M. casta* had accumulated the highest amount of cadmium (Figures 1 and 2).

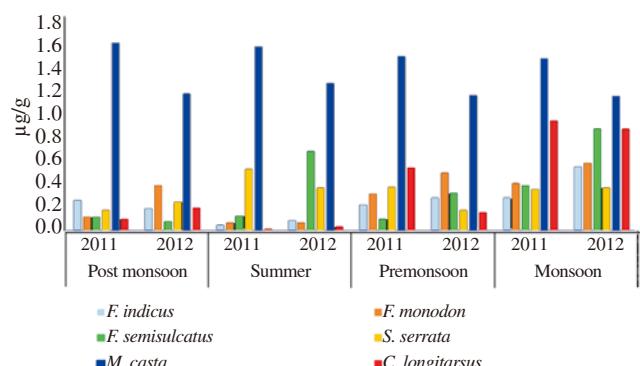
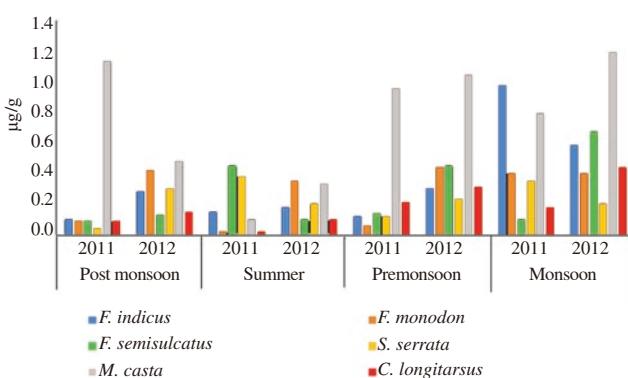


Figure 1. The presence of cadmium in gills of shellfish species.



**Figure 2.** The presence of cadmium in muscles of shellfish species.

#### 4. Discussion

Cadmium is present in the coastal and estuarine environments and does not have any biological functions in organisms. It gets accumulated in marine invertebrates especially crustaceans in the detoxified form[32-35]. However, its toxicity is very high to the aquatic animals and can affect their physiology even at low levels[36-40].

Besides, scanty reports are present to illustrate the accumulation of cadmium in the penaeid decapods which are of economic and ecological importance in the tropics and subtropics[41-43]. Marine bivalves which are exposed to different concentrations of cadmium tend to concentrate this heavy metal in their tissues acquired from sediments and water to a very high magnitude above their exposure concentrations[44-46]. However, high cadmium concentrations result in acute toxicity, whereas cellular and physiological processes are affected even at low, environmentally realistic concentrations[39,40,47-49].

Cadmium is widely distributed in the earth's crust. Its main sources are from cadmium rich soils from which there is runoff to the river, leaching of rocks and deposition of diatoms in the sediments[21,45]. It is mainly used as pigments in plastics, as stabilizers and in electroplating. Smelting and mining of lead and zinc produce cadmium as a byproduct. Cadmium is also used in the manufacture of nickel-cadmium batteries. Use of commercial fertilizers, insecticides and fungicides containing cadmium increases its concentration in the soil. Exposure to alloys used in dentistry, motor oil and automobile exhausts are lesser known sources of cadmium which generally affect the kidneys, liver, lungs, brain, bones and the placenta[50]. High concentration of cadmium leads to severe health hazards to humans and is a potential toxin along with mercury. Consumption of food or intake of water with high cadmium levels leads to vomiting, irritation of the stomach, diarrhoea and sometimes even death. Ingesting low amounts of cadmium during an extended time can result in its accumulation in the kidneys, thereby causing kidney damage and also fragile bones. Studies on human and rats have pointed out that cadmium and its compounds may be potential carcinogens[10].

In conclusion, the result of the present study has shown that the accumulations of cadmium found in the gills and muscles were high in *M. casta* when compared to other species of shellfish. Thus, the consumption of the shellfish is safe, but do not exclude bioaccumulation risk in crustacean meat. This present study has

highlighted the need for estuarine biomonitoring to avoid possible contamination of shellfish and its consumers. The overall scenario of the shellfish accumulating high levels of cadmium indicates that the Pulicat lake is polluted with undesirable elements and the risk of consuming the meat of shellfish by man and other carnivores may lead to their toxicity. Stringent control measures are necessary to control the pollution of this precious lake to reduce the bioaccumulation of toxic metals in organisms.

#### Conflict of interest statement

We declare that we have no conflict of interest.

#### References

- Rajendran N, Sanjeevi SB, Khan SA, Balasubramanian T. Ecology and biodiversity of Eastern Ghats-Estuaries of India. *EPTRI-ENVIS Newslett* 2004; **10**(4): 1-12.
- Connell DW, Lam P, Richardson B, Wu R. *Introduction to ecotoxicology*. Oxford: Blackwell Science Ltd; 1999, p. 71.
- Franca S, Vinagre C, Cacador I, Cabral HN. Heavy metal concentrations in sediment, benthic invertebrates and fish in three salt marsh areas subjected to different pollution loads in the Tagus Estuary (Portugal). *Mar Pollut Bull* 2005; **50**: 993-1018.
- Cajaraville MP, Bebianno MJ, Blasco J, Porte C, Saarasquete C, Varengo A. The use of biomarkers to assess the impact of pollution in coastal environments of the Iberian Peninsula: a practical approach. *Sci Total Environ* 2000; **247**: 295-311.
- Ravera O. Monitoring of the aquatic environment by species accumulator of pollutant: a review. *J Limnol* 2001; **60**(Suppl 1): S63-78.
- Otchere FA. Heavy metals concentrations and burden in the bivalves (*Anadara (Senilia) senilis*, *Crassostrea tulipa* and *Perna perna*) from lagoons in Ghana: model to describe mechanism of accumulation/excretion. *Afr J Biotechnol* 2003; **2**(9): 280-7.
- Goldberg ED. *The health of the oceans*. Paris: United Nations Educational, Scientific, and Cultural Organization; 1976, p. 172.
- Phillips DJH. *Quantitative aquatic biological indicators: their use to monitor trace and organochlorine pollution*. London. London: Applied Science Publishers; 1980.
- Cossa D. A review of the use of *Mytilus* spp. as quantitative indicators of cadmium and mercury contamination in coastal waters. *Oceanologica Acta* 1989; **12**(4): 417-32.
- Gopinathan KM, Amma SR. Bioaccumulation of toxic heavy metals in the edible soft tissues of green mussel (*Perna viridis* L.) of Mahe region. Pondicherry: Government of Pondicherry; 2008, p. 1-32. [Online] Available from: <http://dste.puducherry.gov.in/gopinath.pdf> [Assessed on 5 October, 2016]
- Beldi H, Gimbert F, Maas S, Scheifler R, Soltani N. Seasonal variations of Cd, Cu, Pb and Zn in the edible mollusc *Donax trunculus* (Mollusca, Bivalvia) from the Gulf of Annaba, Algeria. *Afr J Agric Res* 2006; **1**: 85-90.
- Ndome CB, Ekaluo UB, Asuquo FE. Comparative bioaccumulation of heavy metals (Fe, Mn, Zn, Cu, Cd, Cr) by some edible aquatic mollusc from the Atlantic coastline of South Eastern Nigeria. *World J Fish Mar Sci* 2010; **2**: 317-21.
- Çulha M, Bat L, Türk-Çulha S, Gargaci A. Some mollusk species on

- the hard-bottom of Sinop Peninsula (Central Black Sea). *Turkish J Aquat Life* 2007; (5-8): 242-57.
- [14] Astudillo LR, Yen IC, Bekele I. Heavy metals in sediments, mussels and oysters from Trinidad and Venezuela. *Rev Biol Trop* 2005; **53**(Suppl 1): S41-53.
- [15] Luoma SN. Can we determine the biological availability of sediment-bound trace metals. *Hydrobiologia* 1989; **176**: 379-96.
- [16] Davies CAL, Tomlinson K, Stephenson T. Heavy metals in river Tees estuary sediments. *Environ Technol* 1991; **12**: 961-72.
- [17] Chang JS, Yu KC, Tsai LJ, Ho ST. Spatial distribution of heavy metals in bottom sediment of Yenshui river, Taiwan. *Water Sci Technol* 1998; **38**(11): 159-67.
- [18] Rainbow PS. Trace metal concentrations in aquatic invertebrates: why and so what? *Environ Pollut* 2002; **120**(3): 497-507.
- [19] El-Moselhy KM, Yassien MH. Accumulation patterns of heavy metals in venus clams, *Paphia undulata* (Born, 1780) and *Gastrarium pectinatum* (Linnaeus, 1758), from lake Timsah, Suez Canal, Egypt. *Egypt J Aquat Res* 2005; **31**(1): 13-28.
- [20] Bryan GW, Langston WJ, Hummerstone LG, Burt GR. *A guide to the assessment of heavy metal contamination in estuaries using biological indicators*. Plymouth: Marine Biology Association of the United Kingdom; 1985, p. 1-92.
- [21] World Health Organization. Land/sea boundary flux of contaminants: contributions from rivers. Geneva: World Health Organization; 1987; **32**: 1-172.
- [22] Nriagu JO, Sprague JB. *Cadmium in the aquatic environment*. New York: John Wiley and Sons Inc.; 1987.
- [23] Batvari BP, Kamala-Kannan S, Shanthi K, Krishnamoorthy R, Lee KJ, Jayaprakash M. Heavy metals in two fish species (*Carangoides malabaricus* and *Belone stranglurus*) from Pulicat lake, north of Chennai, southeast coast of India. *Environ Monit Assess* 2008; **145**: 167-75.
- [24] Ramadevi K, Indra TJ, Ragunathan MB. Fishes of Pulicat lake. *Rec Zool Surv India* 2004; **102**(3-4): 33-42.
- [25] Periakali P, Padma S. Mercury in Pulicat lake sediments, east coast of India. *J Ind Assoc Sedimentol* 1998; **17**: 239-44.
- [26] Padma S, Periakali P. Physico-chemical and geochemical studies in Pulicat lake, east coast of India. *Indian J Mar Sci* 1999; **28**: 434-7.
- [27] Watting RJ, Emmerson WD. A preliminary pollution survey of the Papenkuijs river, Port Elizabeth. *Water Sci Afr* 1981; **7**: 211-5.
- [28] Bhargavan BPV. Haematological responses of green mussel *Perna viridis* (Linnaeus) to heavy metals copper and mercury[Dissertation]. Cochin: Cochin University of Science and Technology; 2008, p. 163-74.
- [29] Ghosh TK, Kshirsagar DG. Selected heavy metals in seven species of fishes from Bombay offshore areas. Proceedings of the National Academy of Sciences of India. Section B. Biol Sci 1993; **63**(3): 350-11.
- [30] Amiard JC, Amiard-Triquet C, Berthet B, Metayer C. Comparative study of the patterns of bioaccumulation of essential (Cu, Zn) and non-essential (Cd, Zn) trace metals in various estuarine and coastal organisms. *J Exp Mar Biol Ecol* 1987; **106**: 73-89.
- [31] Vijayaraman K. Physiological responses of the freshwater prawn, *Macrobrachium malcolmsonii* to the heavy metals cadmium, copper, chromium and zinc [Dissertation]. Thiruchirappalli: Bharathidasan University; 1994.
- [32] Rainbow PS, White SL. Comparative strategies of heavy metal accumulation by crustaceans: zinc, copper and cadmium in a decapod, an amphipod and a barnacle. *Hydrobiologia* 1989; **174**: 245-62.
- [33] Rainbow PS. Phylogeny of trace metal accumulation in crustaceans. In: Langston WJ, Bebianno MJ. *Metal metabolism in aquatic environments*. London: Chapman & Hall; 1998, p. 285-319.
- [34] Rainbow PS. Trace metal concentrations in aquatic invertebrates: why and so what? *Environ Pollut* 2002; **120**: 497-507.
- [35] Wang WX. Interactions of trace metals and different marine food chains. *Mar Ecol Prog Ser* 2002; **243**: 295-309.
- [36] Byczkowski JZ, Sorenson JRJ. Effects of metal compounds on mitochondrial function: a review. *Sci Total Environ* 1984; **37**: 133-62.
- [37] Viarengo A. Heavy metal cytotoxicity in marine organisms: effects on Ca<sup>2+</sup> homeostasis and possible alteration of signal transduction pathway. In: *Advances in comparative and environmental and physiology*. 1994; **20**: 85-109.
- [38] Stohs SJ, Bagchi D. Oxidative mechanisms in the toxicity of metal ions. *Free Radic Biol Med* 1995; **18**: 321-36.
- [39] Sokolova IM. Cadmium effects on mitochondrial function are enhanced by elevated temperatures in a marine poikilotherm, *Crassostrea virginica* Gmelin (Bivalvia: Ostreidae). *J Exp Biol* 2004; **207**: 2639-48.
- [40] Sokolova IM, Sokolov EP, Ponnappa KM. Cadmium exposure affects mitochondrial bioenergetics and gene expression of key mitochondrial proteins in the eastern oyster *Crassostrea virginica* Gmelin (Bivalvia: Ostreidae). *Aquat Toxicol* 2005; **73**: 242-55.
- [41] Holthuis LB. *Shrimps and prawns of the world: an annotated catalogue of species of interest to fisheries*. Rome: Food and Agriculture Organization of the United Nations; 1980, p. 271.
- [42] Grey DL, Dall W, Baker A. *A guide to the Australian penaeid prawns*. Darwin: Northern Territory Government Printing Office; 1983, p. 140.
- [43] Perez-Farfante I, Kensley B. Penaeoid and sergestoid shrimps and prawns of the world: Keys and Diagnoses for the Families and Genera (Memoires du Museum National d'Histoire Naturel). Paris: Museum National d'Histoire Naturelle; 1997, p. 1-233.
- [44] Roesjadi G. Environmental factors: response to metals. In: Kennedy VS, Newell RIE, Eble AF, editors. *The eastern oyster Crassostrea virginica*. College Park: University of Maryland; 1996, p. 515-37.
- [45] Frew RD, Hunter KA, Beyer R. Cadmium in oysters and sediments from Foveaux Strait, New Zealand. In: Macaskill RB, editor. *Proceedings of the Trace Element Group of New Zealand*. Hamilton: Waikato University Press; 1997, p. 1-23.
- [46] Engel DW. Accumulation and cytosolic partitioning of metals in the American oyster *Crassostrea virginica*. *Mar Environ Res* 1999; **47**: 89-102.
- [47] Anderson RS, Oliver LM, Jacobs D. Immunotoxicity of cadmium for the eastern oyster [*Crassostrea virginica* (Gmelin, 1791)]: effects on hemocyte chemiluminescence. *J Shellfish Res* 1992; **11**: 31-5.
- [48] Roesjadi G, Brubacher LL, Unger ME, Anderson RS. Metallothioneine mRNA induction and generation of reactive oxygen species in molluscan hemocytes exposed to cadmium *in vitro*. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 1997; **118**: 171-6.
- [49] Auffret M, Mujdzic N, Corporeau C, Moraga D. Xenobiotic-induced immunomodulation in the European flat oyster, *Ostrea edulis*. *Mar Environ Res* 2002; **54**: 585-9.
- [50] Children's Environmental Health Network. Training manual on pediatric environmental health: putting it into practice. Emeryville: Children's Environmental Health Network; 1999.