**HEPATOPROTECTIVE AND IMMUNOMODULATORY PROPERTIES OF AQUEOUS EXTRACT OF CURCUMA LONGA IN CARBON TETRA CHLORIDE INTOXICATED SWISS ALBINO MICE**

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**Objective:** To evaluate the hepatoprotective and immunotherapeutic effects of aqueous extract of turmeric rhizome in CCl₄ intoxicated Swiss albino mice. **Methods:** First group of mice (n=5) received CCl₄ treatment at a dose of 0.5 mL/kg bw (i.p.) for 7 days. Second group was fed orally the aqueous extract of turmeric at a dose of 50 mg/kg bw for 15 days. The third group was given both the turmeric extract (for 15 days, orally) and CCl₄ (for last 7 days, i.p.). The fourth group was kept as a control. To study the liver function, the transaminase enzymes (SGOT and SGPT) and bilirubin level were measured in the serum of respective groups. For assaying the immunotherapeutic action of *Curcuma longa* (*C. longa*), non specific host response parameters like morphological alteration, phagocytosis, nitric oxide release, myeloperoxidase release and intracellular killing capacity of peritoneal macrophages were studied from the respective groups. **Results:** The result of present study suggested that CCl₄ administration increased the level of SGOT and SGPT and bilirubin level in serum. However, the aqueous extract of turmeric reduced the level of SGOT, SGPT and bilirubin in CCl₄ intoxicated mice. Apart from damaging the liver system, CCl₄ also reduced non specific host response parameters like morphological alteration, phagocytosis, nitric oxide release, myeloperoxidase release and intracellular killing capacity of peritoneal macrophages. Administration of aqueous extract of *C. longa* offered significant protection from these damaging actions of CCl₄ on the non specific host response in the peritoneal macrophages of CCl₄ intoxicated mice. **Conclusions:** In conclusion, the present study suggests that *C. longa* has immunotherapeutic properties along with its ability to ameliorate hepatotoxicity.

**1. Introduction**

The liver is the main organ responsible for the biosynthesis, uptake and degradation of proteins and enzymes. Liver function may therefore be reflected to some extent on the levels and/or the activities of these circulating biochemical compounds. The immune system is increasingly found to be involved in the development of several chronic illnesses, for which allopathic medicine has provided limited tools for treatment and especially prevention. In that context, it appears worthwhile to target the immune system in order to modulate the risk of certain chronic illnesses. Meanwhile, natural health products (NHPs) are generating renewed interest, particularly in the prevention and treatment of several chronic diseases. Herbal drugs are known to possess immunomodulatory properties and generally act by stimulating or suppressing both specific and non-specific immunity[1].

Turmeric is the dried rhizome powder of *Curcuma longa* (*C. longa*), a perennial herb of the Zingiberaceae family. *C. longa* is cultivated extensively in Asia (India and China). The major chemical principles of turmeric are curcuminoinds, which impart characteristic yellow colour to it.

Previous works have shown that *C. longa* inhibited the growth of activity of some bacteria and fungi[2,3]. Turmeric extract was found to possess in vitro free radical scavenging, ROS scavenging ability and cell proliferation activity in cell line[4]. Curcumin seems to regulate the immune function of mice in a dose-dependent fashion as curcumin treatment enhanced the phagocytosis of peritoneal macrophages and differentially regulates the proliferation of splenocytes[5]. The effect of curcumin in Alzheimer’s disease is mediated through the downmodulation of cytokine (i.e., TNF-α and IL-1β) and chemokine (i.e., MIP-1β, MCP-1, and IL-8) activity in peripheral blood monocytes and reduces
immunotherapeutic causes as well as ameliorating cytotoxic, antiproliferative, and anti-inflammatory activities of the hyaluronidase enzyme by the oil. Further, the oil produced was kept in the deep freezer at -20°C overnight and then subjected to freeze drying. Extract obtained by this method was be then weighed and stored at 22°C in desiccators until further use.

2.2. Animals

20 mice weighing approximately (20±1.0) g were taken and these mice were divided into four groups of five mice each. The first group was kept as control. The second group consisted of CCl4, at a dose of 0.5 mL/kg bw (i.p.) from the 8th day to 15th day of the experiment. In the third group, the mice were fed with extract of turmeric at a concentration of 50 mg/kg bw (orally) by feeding needle for 15 days. In the fourth group, the mice were given both turmeric extract (for 15 days, orally) and CCl4 (for last 7 days, i.p.). Animal experiments were in accordance with the instructions for the care and use provided by the institution at which the research was carried out.

2.3. Collection of blood

Blood was collected from the retro–orbital plexus of the animals and serum isolated from the blood samples. The animals were sacrificed on the 16th day of treatment. Peritoneal macrophages were isolated from the sacrificed mice.

2.4. Determination of serum bilirubin level

The formation of pink–colored azo–bilirubin by the reaction between bilirubin and a diazo reagent was utilized. In practice, one estimation was done with addition of alcohol and the other without alcohol. The former yields total bilirubin and latter conjugated bilirubin.

2.5. Estimation of serum glutamate oxaloacetate transaminase (aspartate transaminase) and serum glutamate pyruvate transaminase (alanine transaminase) levels in serum

For each enzyme, 0.1 mL of non–hemolyzed serum was mixed respectively with 0.5 mL of glutamic–oxaloacetic transaminase (SGOT or AST) substrate and 0.5 mL glutamic–pyruvate transaminase (SGPT or ALT) substrate and incubated for 1 h at 37°C. Then 0.5 mL of 2, 4–dinitrophenyl hydrazine solution was added to the respective test tubes and allowed to stand for 15 min at room temperature. Then, 5 mL of 0.4 (N) NaOH was added, mixed and kept at room temperature for 20 min. The intensity of the developed color was read at 540 nm after setting the instrument to zero density with water. The decrease in density represents the decrease in α–ketoglutarate from which the activity was calculated.

2.6. Isolation of peritoneal macrophages

On the 13th day, all the mice were injected with 50 μL of 3% starch (i.p.). After two days they were sacrificed by cervical dislocation. Following this 5 mL of ice–cold RPMI–1640 were injected (i.p.) in all the dead mice and the injected area was lavaged softly. A small perforation was made to withdraw the RPMI–1640 containing peritoneal fluid. The process was repeated to accumulate the remaining fluid by aspirating and collecting in plastic centrifuge tubes. The samples were then centrifuged for 30 min at 3 500 rpm. All the samples were then washed in RPMI–1640 twice and the pellet were collected separately and incubated for 2 h at 37°C. The supernatant was decanted and the adhered macrophages present in the micro centrifuge tubes were resuspended in 1 mL RPMI–1640. A portion of entire cell samples (100 μL) isolated from all the mice, were then aspirated and smeared on a glass slide and incubated for 1 h, fixed with 4.1% formaldehyde, kept for 30 min before staining with Giemsa
and all these assays were performed separately for each group of mice. After that the slides were washed, air dried and observed under microscope. The other portions of cell samples were kept intact for further assays[20].

2.7. Morphological alteration assay

100 µL sample cells in HBSS–BSA from all mice were taken separately and fixed in an equal volume of 2.5% glutaraldehyde in HBSS. After 10 min, cells were centrifuged at 2000 rpm for 5 min. The supernatant was removed and the pellet was resuspended in HBSS. Smears of cells were drawn on glass slides, air dried, fixed in methanol and stained with Giemsa. Cells were observed under oil immersion microscope. Any cell having intensive rough surface was scored as polarized and this was expressed as a percentage of the total number of cells counted[21].

2.8. In vitro cell adhesion assay

Cells were seeded separately for different groups in 96-well microtitre plates and allowed to adhere for different times. In time, wells were washed with HBSS, and then 100 µL of 0.5% crystal violet in 12% neutral formaldehyde, and 10% ethanol was added to each well and incubated for 4 h to fix and stain the cells. Wells were washed and air dried for 30 min. Crystal violet was extracted from the macrophage adhered in the wells by lysing with 0.1% SDS in HBSS. Absorbance was measured spectrophotometrically at 570 nm. Cell adhesion was expressed as increased absorbance at 570 nm[22].

2.9. Phagocytosis assay

100 µL of cells from all mice were allowed to adhere separately on glass slides whereas non adherent cells were washed out with DPBS (1×). To the glass slides containing adhered macrophages 10% heat killed Staphylococcus aureus was added and incubated for 3 h at 37 °C which were then washed with DPBS (1×) and dried. The cells were at last fixed in 50% methanol, stained with Giemsa, observed under oil immersion microscope and counted for number of bacterial cells ingested[23].

2.10. Myeloperoxidase release assay

200 µL of cells from different groups were taken into micro centrifuge tubes and stimulated with LPS (100 ng/mL) for 1 h at 37 °C and centrifuged at 13 000 rpm for 10 min. The supernatant thus obtained from different sets was recovered separately and kept at −20 °C until further use. The cell free supernatant was used for assay of the partial MPO release for different groups. The pellet that recovered from all the four groups were lysed in 0.01% SDS and then centrifuged again. The supernatant was recovered as before for total MPO release assay. Subsequently 100 µL of cell free supernatant as well as from lysis of cells were reacted with 100 µL of 0.5% crystal violet in HBSS–BSA containing 10% normal serum and incubated at 37 °C for 25 min. At 0 min and after 15, 30 and 45 min time intervals 0.1 mL sample was removed each time and treated with gentamycin. The above content was then plated onto nutrient agar petriplate and the number of viable intracellular bacteria was determined by counting the individual colony formed[26].

2.11. Nitric oxide release assay

100 µL of macrophage cells were isolated from respective groups from 10 cells/mL dilution and then suspended in DPBS–BSA. The cells were then stimulated with LPS (100 ng/mL) for 1 h at 37 °C[25].

2.12. Intracellular killing assay

In this procedure, 100 µL of bacterial cells were incubated separately for all groups with macrophages in a total volume of 1 mL DPBS–BSA and kept in the shaker for 20 min at 37 °C. Non ingested bacteria were then removed by differential centrifugation for 10 min at 10 000 rpm at 4 °C. Following that, two washes were given with ice-cold DPBS–BSA. The cells containing ingested bacteria were resuspended in DPBS–BSA containing 10% normal serum and incubated at 37 °C for 25 min. At 0 min and after 15, 30 and 45 min time intervals 0.1 mL sample was removed each time and treated with gentamycin. The above content was then plated onto nutrient agar petriplate and the number of viable intracellular bacteria was determined by counting the individual colony formed[26].

2.13. Statistical analysis

A one–tailed Student’s t test as well as ANOVA was performed to compare the mean values of control and turmeric extract group. The results were expressed as mean ± standard error mean and all the experiment was done in triplicates.

3. Result

3.1. Effect of C. longa on serum bilirubin level in CCl₄ intoxicated male albino mice

Figure 1 showed the effect of C. longa on bilirubin level in serum of CCl₄ intoxicated male albino mice. The control had shown the bilirubin level in serum of (11.34±0.41) mg/100 mL but after CCl₄ treatment, it increased to (43.65±0.81) mg/100 mL. Whereas after administration of C. longa at a dose of 60 mg/kg bw in CCl₄ intoxicated mice, the bilirubin level reduced to (14.65±0.69) mg/100 mL. The reduced level of bilirubin was also observed (10.92±0.37) mg/100 mL in control group after C. longa administration (Figure 1, P=***).
3.2. Effect of C. longa on serum SGOT (AST) level in CCl4 intoxicated male albino mice

Figure 2 showed the effect C. longa on SGOT (AST) level in serum of CCl4 intoxicated male albino mice. The control had shown the SGOT (AST) level in serum of (12.99±0.24) IU/L but after CCl4 treatment, it increased to (36.68±0.52) IU/L. Whereas after administration of C. longa at a dose of 60 mg/kg bw in CCl4 intoxicated mice, the SGOT (AST) level reduced to (15.22±0.36) IU/L. The reduced level of SGOT (AST) was also observed (13.54±0.36) IU/L in control group after C. longa administration (Figure 2; \( P=**** \)).

3.3. Effect of C. longa on serum SGPT (AST) level in CCl4 intoxicated male albino mice

Besides the measurements of serum SGOT level in CCl4 intoxicated male albino mice, it is also important to see the effects of C. longa on serum SGPT level (Figure 2). After CCl4 treatment in control mice, the serum SGPT level increased from (17.57±0.27) IU/L to (41.85±0.73) IU/L. However, administration of C. longa reduced the serum SGPT level to (18.14±0.46) IU/L. Administration of C. longa in control mice also showed some reduction of SGPT level i.e. up to (16.78±0.41) IU/L in serum (Figure 2; \( P=**** \)).

3.4. Effect of C. longa on morphology of peritoneal macrophages isolated from CCl4 intoxicated male albino mice

Morphology of macrophage plays a prominent part of their function. To demonstrate the effect of both CCl4 and C. longa on the number of differentiated macrophages, the macrophage cells deviated from spherical outline were measured and counted as morphologically altered macrophage. From this experiment, it was found that CCl4 treatment increased the number of undifferentiated macrophages from (28.04±0.75)% to (63.24±3.36)%. However the administration of C. longa in CCl4 intoxicated mice, the number of undifferentiated macrophages became almost normal level of (38.59±2.17)%. Administration of C. longa in control mice also showed some significant reduction of undifferentiated macrophages up to a level of (21.30±2.18)% (Figure 3; \( P=**** \)).

3.5. Effect of C. longa extract on the adhesion property of CCl4 intoxicated mice peritoneal macrophages

In vitro cell adherence assay may reflect the in vivo capacity for cellular adherence with treatment by C. longa extract hepatotoxic mice. In order to address this fundamental characteristic of macrophages, the cell adhesion property was assayed in vitro. Adhesion of cells increased gradually after C. longa administration (Figure 4; \( P=*** \)).

3.6. Effect of C. longa on phagocytosis capacity of peritoneal macrophages isolated from CCl4 intoxicated male albino mice

To evaluate the effect of C. longa on phagocytosis capacity of peritoneal macrophages, the phagocytosis index was measured. CCl4 administration in control mice reduced the phagocytosis index from (2 160.00±36.22) to (872.00±10.35). But after the administration of C. longa, the phagocytosis index of CCl4 intoxicated peritoneal macrophages increased to (2 104.00±10.81). Apart from increasing the phagocytosis index of CCl4 intoxicate peritoneal macrophages, C. longa treatment also increased the phagocytosis index of control peritoneal macrophages to the level of (2 783.4±9.4) (Figure 5; \( P=**** \)).
Figure 5. Effect of C. longa on phagocytosis of peritoneal macrophages in CCl4 intoxicated Swiss albino mice.

3.7. Effect of C. longa on myeloperoxidase (MPO) enzyme release from peritoneal macrophages isolated from CCl4 intoxicated male albino mice

MPO reduced the free radical level in our system. CCl4 caused an increase in the level of CCl4 production, but whether C. longa causes any alteration in the level of free radicals or not was the present objective of our study. CCl4 intoxication reduces the enzyme release from (63.56±0.33) to (42.82±1.25) as in the case of a control group. However, administration of C. longa extract in CCl4 intoxicated mice sufficed to protect the macrophage and induce (60.46±0.39)%.

Figure 6. Effect of C. longa on MPO enzyme release from peritoneal macrophages isolated from CCl4 intoxicated male albino mice.

3.8. Effect of C. longa on nitric oxide (NO) release from peritoneal macrophages isolated from CCl4 intoxicated male albino mice

Nitric oxide plays an important part to kill the microbial pathogen inside the macrophage by forming sodium hypochloride (NaOCl). CCl4 intoxication significantly lowered the nitric oxide release to (6.7±0.4) μM from (17.15±0.66) μM as in the case of control group. However, C. longa administration ameliorated the effect of CCl4 in CCl4 intoxicated male albino mice (Figure 7; P =***).

Figure 7. Effect of C. longa on NO release of peritoneal macrophages isolated from CCl4 intoxicated male albino mice.

3.9. Effect of C. longa on intracellular killing capacity of peritoneal macrophages isolated from CCl4 intoxicated male albino mice

Whether C. longa administration in CCl4 intoxicated mice has any effect in the killing capacity of peritoneal macrophages was the last and foremost important objective our study. Our results suggested that CCl4 intoxication in control mice reduced the intra cellular killing capacity by five folds but after administration of C. longa aqueous extract in CCl4 intoxicated mice restores the killing capacity of peritoneal macrophages (Figure 8; P =***).

Figure 8. Effect of C. longa extract on the killing capacity of peritoneal macrophages isolated from CCl4 intoxicated male albino mice.
4. Discussion

This study was undertaken to determine the effect of C. longa on liver function as well as the immune functional activities of peritoneal macrophages in CCl4 intoxicated Swiss albino mice. Our results provide evidence for the adverse effect of CCl4 on the liver functions when enzymes such as SGOT and SGPPT were found elevated in the serum. Since the estimation of serum enzymes such as SGOT and SGPPT was a good indicator for the pathological manifestation of jaundice, there was no doubt that CCl4 induces liver damages in these experimental animals. However, in vivo administration of C. longa aqueous extract causes hepatoprotective activities in CCl4 toxicity. The enzyme level goes back to the normal level, as in the case of control group. In our study, administration of C. longa was found to provide significant protection against a CCl4-induced increase in level of SGOT, SGPPT and serum bilirubin. CCl4 induces elevated levels may indicate liver damage, since the major site transamination reaction is in the liver. In transaminases, pyridoxal phosphate (PLP) the prosthetic group of the enzyme, forms a Schiff base intermediate which remains tightly bound to the enzyme by multiple non-covalent interactions. Turmeric treatment during CCl4 intoxication may enhance the above mentioned molecular mechanism of enzyme action.

When old red blood cells are destroyed the in the tissue macrophage system, the globin portion of the hemoglobin molecule is split off, and the heme is converted to biliverdin. In humans, most of the biliverdin is converted to bilirubin and excreted in the bile. The iron from the heme is reused for hemoglobin synthesis. The bilirubin is bound to albumin in the circulation. Some of its tightly bound, but most of it can dissociate in the liver, and free bilirubin enters liver cells, where it is bound to cytoplasmic proteins. It is next conjugated to glucuronic acid in a reaction catalyzed by the enzyme glucuronyl transferase. Each bilirubin molecule reacts with two uridine diphosphoglucuronic acid (UDPGLA) molecules to form bilirubin diglucuronide. This glucuronide, which is more water soluble than the free bilirubin, is then transported against a concentration gradient by a presumably active process into the bile canaliculi. A small amount of the bilirubin glucuronide escapes into the blood, where it is bound less tightly to albumin than is free bilirubin, and is excreted in the urine. Thus, the total plasma bilirubin normally includes free bilirubin plus a small amount of conjugated bilirubin. Most of the bilirubin glucuronide passes via the bile ducts to the intestine. When any hepatotoxicity appears, the total bilirubin (either free or conjugated bilirubin) rises in the blood and this may be due to five reasons i.e. 1) excess production of bilirubin; 2) decreased uptake of bilirubin into hepatic cells; 3) disturbed intracellular protein binding or conjugation; 4) disturbed secretion of conjugated bilirubin into the bile canaliculi; or 5) intrahepatic or extrahepatic bile duct obstruction. Our results showed after CCl4 intoxication in male albino mice, the total bilirubin rises in serum and it suggested that liver damages occur. However, administration of C. longa in CCl4 intoxicated mice restores the level of bilirubin in serum and this may be due to the obstruction of any above mentioned mechanism. Another consequence of this protective effect of C. longa may be due to the obstruction of all the bilirubin increasing mechanism by different compounds of the extract simultaneously.

Exposure of organisms to bacterial infection results in the activation of a variety of host defense mechanisms which include phagocytosis, degradative enzyme release and respiratory burst response. The first step in macrophage function is the activation or differentiation of macrophages. However intensive rough surface or almost spherical macrophages can be considered as abnormal macrophages because after getting encountered with the antigen, the macrophages must have perfect surface morphology. In our study, it was found that CCl4 intoxication increases this abnormal type of macrophages by two folds but after administration of C. longa aqueous extract in that CCl4 intoxicated mice, the macrophages become acquiring abnormal surface morphology so that they can be function normally. The mechanism involving this protection by C. longa extract probably somehow related to the production of monocyte colony stimulating factor or granulocyte–monocyte stimulating factor (M-CSF/GM-CSF).

In order for circulating macrophages to enter inflamed tissue or peripheral lymphoid organs, the cells must adhere to and pass between the endothelial cells lining the walls of blood vessels, a process called extravasation. Endothelial cells express leukocyte–specific cell adhesion molecules (CAMs). Some of these membrane proteins are expressed constitutively; others are expressed only in response to local concentrations of cytokines produced during an inflammatory response. Recirculating macrophage bears receptors that bind to CAMs on the vascular endothelium, enabling these cells to extravasate into the tissues. The increase in cell adherence in CCl4 intoxicated group after C. longa extract treatment might be primarily caused by an increase expression of cell adhesion molecules on the surface of macrophage cells.

As is evident from the phagocytic index after CCl4 intoxication, it can be suggested that CCl4 groups are more prone to infection, as they cannot phagocytose efficiently and, as a result, cannot clear out the invading microorganism, which may lead to a diseased state upon bacterial invasion. However, administration of C. longa increases the phagocytosis capacity of peritoneal macrophages isolated from CCl4 intoxicated mice and this may due to increased number of cells present in the peritoneum as well as alteration of the ability of those that remain functioning normally.

Macrophage secretes lysosomal proteolytic enzymes active at tissue pH that may be important in their ability to kill tumor and bacterial cells. As macrophage mature, there is a progressive rise in lysosomes and hydrolytic content. Continuous CCl4 treatment may also lead to immaturatation of macrophages in vivo, and it can be assumed that due to this exposure they have lesser lysosomal content and hence are less capable of secreting myeloperoxidase (MPO) and nitric oxide (NO). Due to this lesser lysosomal enzyme, these CCl4,
intoxicated peritoneal macrophages could not able to kill the intracellular *Staphylococcus aureus* and this was evident from our finding increased viability of *Staphylococcus aureus* in CCl4 treated group. However, treatment by *C. longa* extract during CCl4 intoxication may somehow alter the maturation of macrophages, so that they regain their lysosomal hydrolytic content and release more MPO and NO following activation and following this CCl4, intoxicated peritoneal macrophages could be able to regain their intracellular killing capacity.

From our results it can be suggested that turmeric would be effective in ameliorating jaundice with good efficacy and toleration. Additionally, its immunostimulatory effect prevents pathogenic insults in jaundice induced immunocompromised state.

**Conflict of interest statement**

We declare that we have no conflict of interest.

**References**


