Mortality of therapeutic fish *Garra rufa* caused by *Aeromonas sobria*

Juraj Majtán,*, Jaroslav Černy, Alena Ofúkaná, Peter Takáč, Milan Kožánek

1Institute of Zoology, Slovak Academy of Sciences, Dubravska cesta 984 506 Bratislava, Slovakia
2Department of Microbiology, Faculty of Medicine, Slovak Medical University, Limbova 1 283 303, Bratislava, Slovakia

**ARTICLE INFO**

**Article history:**
- Received 10 July 2011
- Received in revised form 5 August 2011
- Accepted 18 August 2011
- Available online 28 February 2012

**Keywords:**
- *Garra rufa*
- Ichthyotherapy
- *Aeromonas sobria*
- Antibiotic treatment
- Antibiotic susceptibility

**ABSTRACT**

**Objective:** To investigate a case of mass mortality of *Garra rufa* (*G. rufa*) from a fish hatchery farm in Slovakia. **Methods:** Causative bacterial agent was swabbing out of affected fish skin area and subsequently identified using commercial test system. Antibiotic susceptibility was determined by the disk diffusion method. **Results:** Infected *G. rufa* was characterized by abnormal swimming behaviour, bleeding of skin lesions and local haemorrhages. Despite of using recommended antibiotic treatment no improvement was achieved and *Aeromonas sobria* (*A. sobria*) was identified as a causative agent of fish mortality. Due to massive fish mortality, antibiotic susceptibility of pure isolated culture of *A. sobria* was evaluated employing eight antibiotics against human infections. *A. sobria* was resistant only against one antibiotic, namely ampicillin. **Conclusions:** These results indicate that *A. sobria* can act as a primary pathogen of *G. rufa* and may be a potential risk factor for immunodeficient or immunoincompetent patients during the ichthyotherapy.

1. **Introduction**

Ichthyotherapy, an alternative biotherapeutic method, has been shown to be effective in the treatment of patients with psoriasis and eczema[1–3]. The Kangal Spa in the Central Anatolia region of Turkey is the place where ichthyotherapy was applied for therapeutic purposes for the first time. Two different types of therapeutic fish, *Cyprinion macrostomus* and *Garra rufa* (*G. rufa*), have been identified and recommended for therapeutic purposes[4].

Nowadays, ichthyotherapy is becoming a more popular method for treatment of skin diseases as well as for improving of skin regeneration in healthy subjects. With increasing application of ichthyotherapy there is a potential risk of zoonotic infection transmission from fish to water or humans.

*Aeromonas* infections are probably the most common bacterial diseases diagnosed in cultured warmwater fish[5–7]. The common occurrence of this disease relates to stress conditions such as poor water quality, overcrowding, or rough handling. *Aeromonas* spp. are known to cause diarrhea, necrotizing fasciitis and sepsis in patients with hepatic diseases, diabetes mellitus and immunocompromised status[8–10]. In addition, skin and soft-tissue infections, including wound infections, are the second most frequent location of isolations of *Aeromonas* spp. in clinical samples, after the gastrointestinal tract[7,11,12].

A very recent study has provided the first suggestive evidence of successful colonization and infection by particular strain of *Aeromonas* spp. after transmission from water to human[13]. Moreover, several reported cases have supported this evidence. Recently, Manresa *et al* described two pediatric cases of *Aeromonas hydrophila* (*A. hydrophila*) folliculitis associated with use of recreational water facilities[14]. Similarly, Mulholland and Yong–Gee reported two young adult patients with a severe inflammatory folliculitis eruption attributed to possible contamination of a poorly maintained home spa bath with *A. hydrophila*[15].

In this study, a case of mortality of *G. rufa* from a fish hatchery farm in Slovakia was investigated. We demonstrate that *A. sobria*, resistant to current recommended treatment, caused the mass mortality of fish. Additionally, an antibiotic
susceptibility of isolated *A. sobria* was evaluated.

2. Materials and methods

2.1. Identification of causal agent of bacterial infection

For laboratory examination, four specimens of *G. rufa* with pronounced exophthalinus, bleeding of the eyes and fins, and locomotor ataxia were taken. For bacteriological analysis sterile cotton swabs were used to collect samples either from affected area of skin lesions and fins or from skin of clinically healthy fish. These samples were used to inoculate into Columbia sheep blood agar (MkB Test, Slovakia). The blood agar plates were incubated at 37 °C and 30 °C for a period of 2 days. Suspected single colonies on blood agar plates were re-streaked onto fresh media to obtain pure isolates. These isolates were identified using the Micro–LA test system (Erba Lachema, Czech Republic).

2.2. Antibiotic susceptibility test

Antibiotic susceptibility was determined by the disk diffusion method with 8 antibiotic disks according to the criteria of the Clinical and Laboratory Standards Institute (CLSI)[16] on Mueller–Hinton agar (Oxoid, Hampshire, UK). The following antimicrobial agents (disk content indicated in parentheses) were tested: amikacin (10 μg), ampicillin (10 μg), cefalotin (30 μg), cefepime (30 μg), cefotaxime (30 μg), cefazidine (30 μg), ceftriaxone (30 μg), gentamicin (10 μg). *Escherichia coli* ATCC 25922 was used as the control.

3. Results

3.1. Identification of Aeromonas sobria (*A. sobria*)

In all four specimens of *G. rufa* presenting skin ulcerations (Figure 1), bacteria can be isolated directly from skin lesions. The bacterial cultures were dominated by the appearance of white–grey colonies on blood agar plates. Using the Micro–LA test system, these bacteria were found to belong to the group of motile aeromonads. *A. sobria* isolated from affected fish was characterized by hemolytic colonies when grown at 30 °C. Only very weak growth was seen at 37 °C.

3.2. Antibiotic susceptibility

Isolated causative bacterial agent of mortality of *G. rufa*, *A. sobria*, was tested on antibiotic susceptibility using eight antibiotics against human infections. Among all antibiotics, *A. sobria* isolate was resistant only against one antibiotic, namely ampicillin.

4. Discussion

An increasing popularity of ichthyotherapy and establishment of numerous ichthyotherapeutic centres are accompanied by more frequent monitoring of water quality and microbial contamination due to possible risk of infection development in humans after water-to–human transmission.

Ichthyotherapy, an alternative and complementary treatment, reduces illnesses such as psoriasis[4]. The skin lesions in patients with psoriasis can be secondary infected with bacteria[17,18]. Many wound and skin infections that complicate skin lesions are caused by mixed bacterial flora. Aerobic and anaerobic; gram–negative and gram–positive organisms, whose origins are the oral, gastro–intestinal and skin flora may be present in such infections. Moreover, various microorganisms are associated with the provocation and/or exacerbation of psoriasis, but their roles in the disease pathogenesis are unknown[19,20]. Brook and colleagues isolated 36 different bacteria from secondary infected psoriasis lesions in 23 specimens[17]. The most predominant bacteria were aerobic or facultative bacteria.

Another source of secondary infections in psoriasis lesions could be water or fish itself in bathing tub during ichthyotherapy procedure[21]. In this study, we identified *A. sobria* as a causative bacterial agent which caused mortality of therapeutic fish *G. rufa*. Similarly, author of recent study described a case of mass mortality of *G. rufa* from a private fish hatchery farm in Korea caused by fish pathogenic bacteria *Citrobacter freundii*[22]. However, healthy individuals exposed to this bacteria are not very likely to get disease, immunodeficient or immunoincompetent people as well as people with other disease problems such as psoriasis and atopic dermatitis are at the highest risk. In a very recent study[13], authors provide the first suggestive evidence of successful colonization and infection by *Aeromonas* species after transmission from water to human. In addition, there is an evidence that proteolytic extracellular products from bacterial pathogen *A. hydrophila* have the ability to inhibit skin antimicrobial peptides[23]. This evidence suggests that *Aeromonas* spp. could disrupt microbial skin barrier through inhibition of antimicrobial peptides.

The presence of skin lesions, local haemorrhages and locomotor ataxia in *G. rufa* infected with *A. sobria* is in agreement with the signs of disease found in fish suffering
from aeromonad infections\textsuperscript{24}. Aeromonad infections have often been related to changing water temperature\textsuperscript{25}. Other factors, significantly related to increasing susceptibility of fish to aeromonad infections, are overcrowding, rough handling, transfer of fish, poor nutritional status and fungal and parasitic infections\textsuperscript{26}. In addition, fish which are in poor environments due to unsatisfactory water quality such as high nitrite levels, low levels of dissolved oxygen, or high levels of carbon dioxide are more susceptible to infection by \textit{Aeromonas} spp.

In conclusion, the findings of the present study provide the first evidence that \textit{A. sobria} is a causative bacterial agent of mass mortality of therapeutic fish \textit{G. rufa}. In addition, isolated \textit{A. sobria} was resistant to ampicilin but sensitive to several antibiotics against human infections.

**Conflict of interest statement**

We declare that we have no conflict of interest.

**Acknowledgements**

This work was supported by the Operational Program Research and Development and co–financed with European Fund for Regional Development (EFRD) with the grant number ITMS 26240220020 for establishment of biotherapeutic facility and technology proposal for production and development of bio–drugs.

**References**